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**Distribution, Ecology, and
Postglacial Dispersal
of Certain Crustaceans and Fishes
in Eastern North America**

Michael J. Dadswell

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Dedication

I wish to dedicate this work to the late Mr. W. Van Vliet. Bill was a close friend and colleague as well as an exceptional field biologist.

He introduced me to the use of otter trawls from small boats and constructed the trawl that I used throughout my study. Bill died in October 1968 while scuba diving in Heney Lake, Quebec, during the course of his studies on deepwater sculpins for his doctorate.

On a nouvellement trouvé dans 246 localités de l'est de l'Amérique du Nord une ou plusieurs des espèces suivantes de crustacés: *Mysis relicta*, *Pontoporeia affinis*, *Gammaracanthus loricatus*, *Limnocalanus macrurus*, *Senecella calanoides*, et (ou) le *Myoxocephalus quadricornis*, qui est un poisson. En conséquence, on sait maintenant qu'une de ces espèces ou davantage, se rencontrent dans 327 localités à l'est du 82^e parallèle. Leur distribution se confine dans certains bassins qu'occupaient jadis de vastes lacs glaciaires communicants, ainsi que leurs déversoirs, ou qu'envahissaient des mers saumâtres et froides à mesure que se retirait la calotte glaciaire. La *Mysis*, la *Pontoporeia*, le *Limnocalanus*, la *Senecella*, et le *Myoxocephalus* se dispersèrent principalement à travers le réseau de lacs glaciaires, tandis que le *Gammaracanthus* se répandit en empruntant les eaux marines qui recouvraient certaines parties du continent. Le *Limnocalanus macrurus* est, de toutes ces espèces, celle que l'on retrouve le plus à l'est dans les eaux douces d'Amérique du Nord.

Les paramètres physico-chimiques des lacs, tant à l'intérieur qu'à l'extérieur de la limite des lacs glaciaires, étaient les mêmes et on ne peut guère leur attribuer l'absence des crustacés qu'on remarque dans les lacs au-delà de cette limite. Toutefois, dans ceux des lacs où les crustacés sont présents, la profondeur maximale, la température, la quantité d'oxygène en solution, le contenu minéral, l'acidité, la pénétration de la lumière et la matière organique en solution ont une interaction qui détermine la composition spécifique de la faune des crustacés. La présence de ces derniers se trouvait limitée surtout par de basses tensions d'oxygène et de hautes températures de fond. C'est la *Mysis relicta* qui tolère le mieux un écart appréciable de ces facteurs, tandis que la *Pontoporeia affinis* y est très sensible.

On a établi expérimentalement que la *Mysis relicta* s'accommode d'une température et d'un indice de salinité plus élevés que ceux que peut supporter la *Senecella calanoides*.

Ces animaux ont probablement évolué dans les mers saumâtres arctiques pour envahir les eaux douces d'Amérique du Nord, soit à l'occasion d'une inondation marine, soit en migrant vers l'intérieur dans les eaux refoulées en bordure des calottes envahissantes. La redistribution postglaciaire de ce groupe (à l'exception du *Gammaracanthus loricatus*) a pu commencer il y a environ 14,500 ans, à partir d'un certain nombre de refugia au sud des Grands Lacs et se poursuivre vers le nord à la suite des glaces en retraite dans les réseaux de lacs glaciaires. Des indices de salinité plutôt élevés dans la mer Champlain originelle ont bloqué la dispersion de certains crustacés, les empêchant de se répandre vers l'est. Le *Gammaracanthus loricatus* semble s'être dispersé depuis l'Arctique en suivant la côte atlantique et en pénétrant dans la vallée du Saint-Laurent. Sauf certains mouvements secondaires vers l'aval dus au rétablissement de l'équilibre isostatique des voies fluviales, la dispersion dans l'est de l'Amérique du Nord s'est terminée il y a environ 6,000 ans, en même temps que disparaissait le dernier réseau de lacs glaciaires dans le centre nord du Québec.

Ces animaux qu'on nomme souvent "reliques de l'époque glacio-marine" sont-ils de véritables reliques dans le sens que donnent à ce terme Darlington ou Ekman? Voici un aspect de leur évolution sur lequel l'ouvrage tente de jeter quelque lumière.

Summary

One or more of the crustaceans *Mysis relicta*, *Pontoporeia "affinis"*, *Gammaracanthus loricatus*, *Limnocalanus macrurus*, *Senecella calanoides*, and/or the fish *Myoxocephalus quadricornis* were found in 246 new localities in eastern North America. One or more of these species are now known from 327 localities east of the 82nd parallel of longitude. Their distributions are restricted to basins in areas formerly occupied by the large, interconnected glacial lakes and their spillways, or by cold, brackish seas during the retreat of the last ice sheet. *Mysis*, *Pontoporeia*, *Limnocalanus*, *Senecella*, and *Myoxocephalus* dispersed primarily through the glacial-lake systems, *Gammaracanthus* by means of marine submergence. *Limnocalanus macrurus* has the most easterly known range of any of the species in the fresh waters of North America.

Physicochemical parameters of lakes inside and outside the glacial-lake boundaries were similar, and cannot be considered responsible for the absence of the crustaceans from lakes outside. In lakes containing the crustaceans, however, maximum depth, temperature, dissolved oxygen, total hardness, pH, light penetration, and dissolved organic matter interact to influence the species composition of the crustacean community. Occurrence of the crustaceans was limited primarily by low oxygen tensions and high bottom temperatures. *Mysis relicta* is the species most tolerant of a wide range of factors, *Pontoporeia "affinis"* the least.

It was experimentally determined that *Mysis relicta* tolerates higher temperatures and salinities than *Senecella calanoides*.

These animals probably evolved in brackish arctic seas and have invaded North American fresh water either by marine inundations or by transportation inland by proglacial waters ponded in front of advancing ice sheets. Postglacial redispersal of the group (except *Gammaracanthus loricatus*) probably began about 14,500 B.P. from a number of refugia south of the Great Lakes, and followed the ice northward in the glacial-lake systems. Higher salinities of the early Champlain Sea acted as a dispersal barrier to some of the crustaceans, limiting their eastward movement. *Gammaracanthus loricatus* apparently dispersed from the Arctic via the Atlantic Coast into the St. Lawrence Valley. Except for some minor downstream movement, caused by isostatic readjustment of drainage systems, dispersal in eastern North America ceased about 6,000 B.P. with the termination of the last glacial-lake systems in north-central Quebec.

Whether these animals, which are often referred to as "glacio-marine relicts", are true relicts in either Darlington's or Ekman's sense is discussed.

Biographical Note

Born in Dafoe, Saskatchewan, in 1944, Dr. Michael J. Dadswell graduated from Carleton University, Ottawa, with an Honours B.Sc. in 1967, and after a work and study trip to the Great Barrier Reef, Australia, he returned to Carleton and began doctoral studies in invertebrate biology and zoogeography. He received his Ph.D. in 1973 and joined The Huntsman Marine Laboratory, New Brunswick, as a research scientist in resource potential. His research has been primarily in the zoogeography and ecology of fresh- and saltwater mysids, plankton ecology, fisheries biology, and physical limnology. At present he is investigating the biology and population dynamics of shortnose sturgeon (*Acipenser brevirostris*) in the St. John River, New Brunswick.

Acknowledgements

It gives me pleasure to acknowledge the following persons for their contributions to this study. Messrs. Chris Morry, Don Moxley, Don Rivard, Jim Kelly, and Gary Laver assisted in the field and provided comradeship during the long trips. The late Bill Van Vliet built the otter trawl used throughout this study. Mr. R.J. Mott of the Geological Survey of Canada provided outboard motors and the inflatable rubber boat used when surveys were conducted by aircraft. Pilots of Fecteau Aerien, Laurentian Air Services, Brochu Air, Kipawa Air Services, and Lakeland Air Services flew in fair weather and foul. Biologists of the Quebec Service de la Faune, Ontario Ministry of Natural Resources, New York State Department of Conservation, Vermont Fish and Game Department, and the Maine Fish and Game Department readily provided information on their respective districts. Mr. W.J. Traversy of the Inland Waters Directorate, Environment Canada, arranged for the complete chemical analysis of my water samples. Dr. E.L. Bousfield and Mr. C.H. Douglas of the National Museum of Natural Sciences provided the illustrations of *Mysis relicta* and *Pontoporeia "affinis"* (Plate 1). Many persons identified or confirmed identifications of specimens: Dr. A.H. Clarke, Jr., and Rev. H.B. Herrington (molluscs); Dr. E.L. Bousfield (peracaridan crustaceans); Dr. D.E. McAllister and Dr. W.B. Scott (fishes); Dr. D.J. Faber (planktonic crustaceans); and Dr. R.W. Davies (leeches).

I especially thank my supervisors, Dr. H.F. Howden and Dr. E.L. Bousfield, for their encouragement during this study and for the stimulating discussions that have influenced many of my ideas. Dr. Bousfield suggested the topic of this research paper.

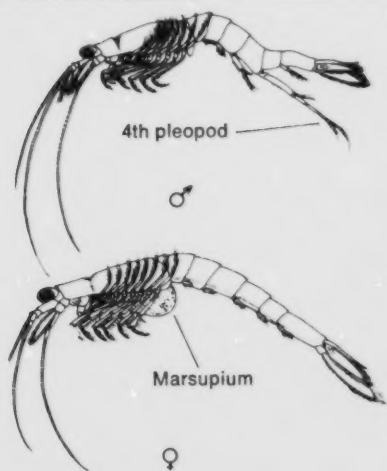
Dr. V.K. Prest helped immeasurably in my understanding of the recession of the last ice sheet and the history of the glacial Great Lakes.

I also thank Mr. J.-P. Cuerrier, who first stimulated my interest in aquatic biology, and who has helped me in so many ways during the last five years.

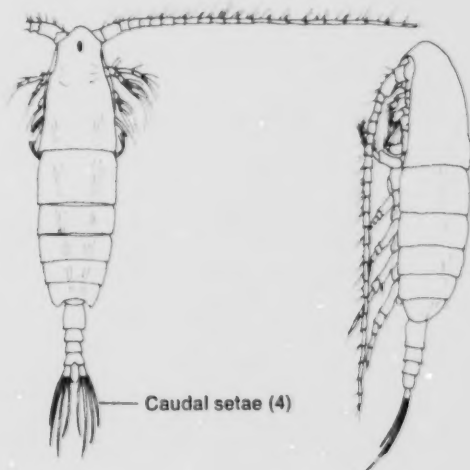
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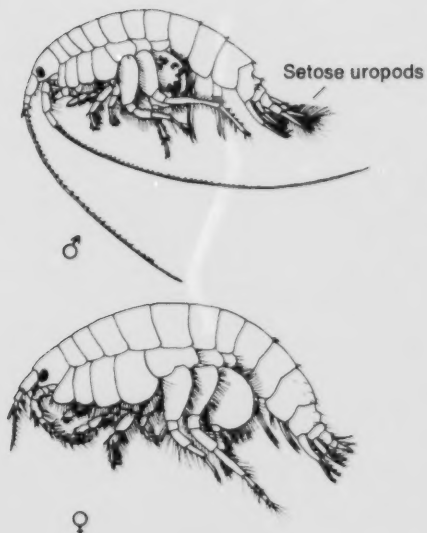
Drs. E.L. Bousfield, H.F. Howden, V.K. Prest, and D.A. Smith critically read the manuscript.



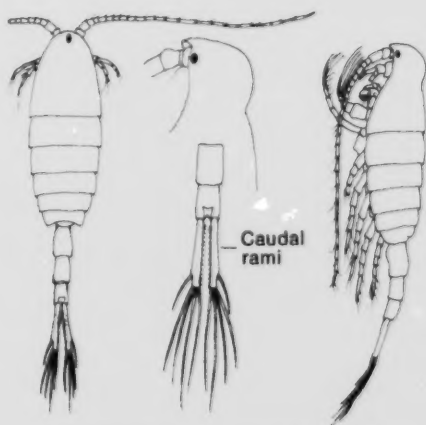
Mysis relicta, × 5



Senecella calanoides, × 25



Pontoporeia "affinis", × 7



Limnocalanus macrurus, × 25
(after Gurney 1923)

Deepwater crustaceans dispersed mainly by glacial lakes in eastern North America

General Introduction

The crustaceans *Mysis relicta* Lovén, *Pontoporeia affinis* Lindstrom, *Gammaracanthus loricatus* Sabine, *Limnocalanus macrurus* Sars, *Saduria entomon* (L.), and the fish *Myoxocephalus quadricornis* (L.) are the North American representatives of a group often referred to as "glaciomarine relicts" (Ricker 1959; Johnson 1964). On the basis of their present taxonomic concept (Holmquist 1959, 1970), these species, or perhaps species complexes (that is, *Pontoporeia "affinis"*), have a Holarctic distribution in brackish portions of arctic seas and in lakes, mainly in the glaciated regions of Eurasia and North America. *Senecella calanoides* Juday is apparently confined to fresh waters of North America, but is often included in this group because it has a similar habitat and distribution (Martin and Chapman 1965).

In view of the wide distribution of this group of animals and their apparent success in fresh water, it seems inappropriate to call them "relicts". *Mysis*, *Pontoporeia*, *Limnocalanus*, *Senecella*, and *Myoxocephalus* form a distinctive deepwater community (Henson 1966) in lakes throughout most of the North American mainland formerly covered by the Laurentide Ice Sheet. They are the main grouping dealt with in this work, and for my purposes will be known as the "deepwater community" or simply the "community".

The presence of these animals in North American lakes was noted as early as 1870 (Smith 1871), but zoogeographical research on them proceeded so slowly that when Ricker (1959) summarized their distributions on this continent, he could find only 43 localities mentioned in the literature. Recently, knowledge of their distributions has been increased in the Arctic (Johnson 1964; Holmquist 1966) and in Ontario (Martin and Chapman 1965; Hamilton 1971). Working in Algonquin Park, Ontario, Martin and Chapman (1965) were the first to determine the precise distributions of these species and relate them to the former extent of glacial lakes.

In North America, the ecology of these species has received greater attention than their zoogeography, but most studies have been confined to animals living in deep oligotrophic lakes (Larkin 1948; Green 1965; Carter 1969; Brownell 1970), and emphasize only low temperatures and high levels of dissolved oxygen as necessary for their survival. On the other hand, it has been shown that these animals can tolerate both high temperatures and

low oxygen levels (Juday and Birge 1927; Lasenby 1971), and it is suspected that other factors may influence their survival in a lake (Holmquist 1959).

The present distribution of any organism depends on a number of factors, including its dispersal mechanisms, dispersal routes open to it in the past, and the present availability of its required habitat. In this study I have attempted to outline the distributions of these species in eastern North America, to determine which environmental factors limit the occurrence of the individual species, and to demonstrate further the relationship between their distribution patterns and the extent of postglacial waters. These exact biological and zoogeographical data should provide a firm base on which inferences about the dispersal history of these organisms can be made.

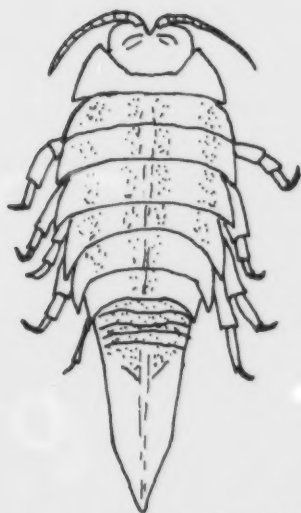
In recent years, fisheries biologists have come to realize the importance of the crustaceans (especially *Mysis relicta*) in the food chains of deepwater communities and their usefulness in fisheries management (Cuerrier and Schultz 1951; Dryer, Erkkila, and Tetzloff 1965; Van Vliet and Qadri 1970; Rawson 1961). For this reason, *Mysis relicta* has been widely introduced into North American lakes, from which it was originally absent (Sparrow, Larkin, and Rutherglen 1964; Linn and Frantz 1965; Schumacher 1966). Consequently, an exact determination of the natural distribution of this deepwater community would be invaluable to future zoogeographical studies. Also, identification of the environmental limiting factors of these organisms should provide fisheries biologists with criteria for selecting the lakes into which these animals could be successfully introduced.

Description of Organisms Studied

The animals studied can be divided into two groups: those whose postglacial dispersal in eastern North America was restricted mainly to glacial lakes, and those which apparently dispersed into lakes of eastern North America via postglacial marine inundations. The first group, the "deepwater community", includes the mysid *Mysis relicta*, the deepwater amphipod *Pontoporeia "affinis"*, the large deepwater copepods *Limnocalanus macrurus* and *Senecella calanoides*, and the sculpin *Myoxocephalus quadricornis* (Plates 1, 3). The second group consists of the amphipod



Gammaracanthus loricatus, $\times 3$



Saduria entomon, natural size

Crustaceans occurring only in former marine-inundated areas.
(Both after Segerstråle 1962.)

Gammaracanthus loricatus, which is known from only one freshwater locality within the study area, and the isopod *Saduria entomon*, which is unknown in the study area (Plate 2).

Mysis relicta, *Limnocalanus macrurus*, and *Myoxocephalus quadricornis* have the widest ranges, and are found in brackish and fresh waters mostly in the glaciated portions of Eurasia and North America (Map 1). The *Pontoporeia* "affinis" complex, which may be represented by different species in the fresh waters of Eurasia and North America, has nearly as extensive a distribution (Map 1). *Gammaracanthus loricatus* and *Saduria entomon* are largely confined to arctic brackish-water localities (Lomakina 1952; Segerstråle 1962), and are found in fresh water only in areas of previous marine inundation (Map 1). *Senecella calanoides* is apparently confined to fresh waters of North America (Map 1). Although Wilson (1959) stated that *Senecella* is known from Siberia, she did not cite a source, and the only primary Russian reference that I have been able to locate (Slastnikov 1940) lists only *Lenicella calanoides*, which may or may not be the same species.

The three deepwater fishes, *Cottus ricei*, *Pungitius pungitius*, and *Percopsis omiscomaycus* (Plate 3), are included in later discussions on postglacial dispersal because their dispersal seems to have been restricted mainly to glacial lakes (Dadswell 1972), and they can be used as subsidiary indicators of ecological conditions of former dispersal routes. They perhaps occupy a comparable zoogeographical position in North America with respect to the "glaciomarine relicts" as the amphipod *Pallasea quadrispinosa* does in Eurasia (Ekman 1920).

Biology of Dispersal

The biological characteristics of an organism, which govern its possible means of dispersal, are an important consideration when discussing the organism's zoogeography. Certain characteristics of these animals limit their dispersal to movement primarily through bodies of standing water.

None of the crustaceans have a diapause period during their life cycle. In both *Mysis* and *Pontoporeia*, the eggs undergo direct development in a marsupium and are not subject to passive dispersal. *Limnocalanus* and *Senecella* shed their eggs in deep water, usually under the ice during winter, and the

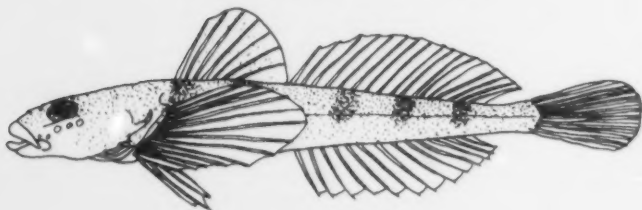
eggs develop immediately (Roff 1972). It is unlikely that any of the adult crustaceans are resistant enough to desiccation to undergo long-distance, passive aerial dispersal; however, very short-distance wind dispersal may be possible, especially in arctic coastal situations. Strong winds, such as tornadoes, usually occur during the day, when these animals are occupying deep water in southern Canadian latitudes (see below). If passive dispersal has actually occurred to any great extent, one would expect the distribution patterns of these animals to show it. They do not.

Usually these animals are light-avoiding and occupy deep water during daylight. *Mysis relicta*, by vertical migrations, maintains itself at light levels of 10^{-1} lux (Beeton 1960). This results in a spatial separation of *Mysis* from most agencies of passive dispersal during daylight (that is, waterfowl, wind), even when surface waters are cold. *Pontoporeia* is completely benthic, and only adult males commonly leave the bottom for more than short periods (McNaught and Hasler 1966). In fresh water, *Limnocalanus* and *Senecella* usually remain in the hypolimnion during daylight (Wells 1960; Carter 1969). This light-avoidance tendency, which may be less prevalent in *Limnocalanus* (MacKay 1924; Grainger 1965), would likely eliminate most possibilities of these animals coming in contact with agents of passive dispersal, at least in the southern parts of their ranges.

In general these animals swim poorly against currents, especially in fresh water. *Mysis* is positively rheotactic and can maintain its position against tidal currents in estuaries (Holmquist 1963). However, Summerhayes and Elton (1923) state that *M. relicta* was unable to swim up the smallest rapids in streams between beach ponds on Spitsbergen Island. More critical evidence was obtained by Dormaar (1970), who showed that *M. relicta* cannot swim against currents greater than 10 cm per second ($\frac{1}{2}$ mile per hour), and that swimming for even short periods in fresh water causes severe osmotic stress to develop in this mysid. Since *Limnocalanus* and *Senecella* are planktonic, they probably possess little or no ability to swim against currents (Hutchinson 1967). The marine form of *Myoxocephalus quadricornis* is known to ascend streams in the Arctic (Johnson 1964), but the lake form has never been captured in streams. Possibly the deepwater lake form is physiologically different from its



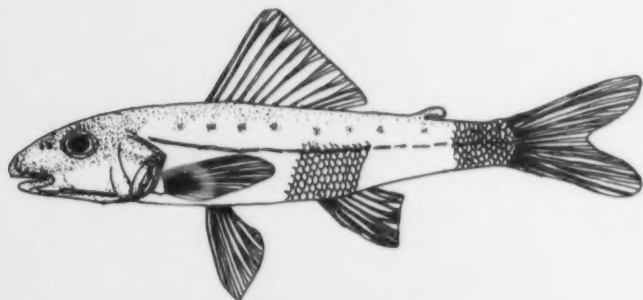
Myoxocephalus quadricornis



Cottus ricei



Pungitius pungitius



Percopsis omiscomaycus

The deepwater fishes of eastern North America that are dispersed mainly by glacial lakes.
All natural size (after McPhail and Lindsey 1970).

marine relative and does not venture from its deepwater habitat. The other deepwater fishes (*Cottus ricei*, etc.), which also occur in cool streams, have distribution patterns that clearly indicate that upstream dispersal has occurred (Dadswell 1972). Regardless, the now well-known distribution pattern of these animals in eastern North America suggests that active upstream movements have not played a major role in their dispersal.

Taxonomic Positions of North American Populations

Research on "glaciomarine relicts" began in 1861 when Lovén described some animals from the deep waters of Swedish lakes and noted that they were the same as, or closely related to, species living in the nearby Baltic Sea. Since then, the nomenclature and the concept of the individual species involved have varied with the views of each author.

Mysis relicta was described by Lovén as separate from the marine *M. oculata*, but for many years most authors considered it to be only a subspecies of *oculata* (Ekman 1920; Pennak 1953). Holmquist (1959) revised the genus *Mysis* and upheld *relicta*. She also compared North American and Eurasian freshwater populations and concluded that they were all the same species, *M. relicta* (in agreement with Tattersall 1951). In fact, Holmquist remarks, and she is supported by Fürst and Nyman (1969), that *relicta* is quite phenotypically conservative for an animal with so many widely and long-isolated populations.

The freshwater *Pontoporeia* of North America was originally described as *P. hoyi*. Segerstråle (1937) revised the genus and concluded that one species, *P. "affinis"*, occurred in brackish and freshwater localities of Eurasia and North America. Bousfield (personal communication) now believes the North American freshwater populations to be a different species from at least the Baltic "*affinis*". At the present time, however, the species concepts are not yet clear, and until they become so, it is better to maintain, with reservation, *P. "affinis"* for the eastern North American freshwater populations.

Gammaracanthus has alternately been considered as two species (*lacustris* in fresh water, *loricatus* in salt water) or as just one species, *loricatus*, in both salt and fresh water. At present the two-species concept is accepted by most authors, even though an ap-

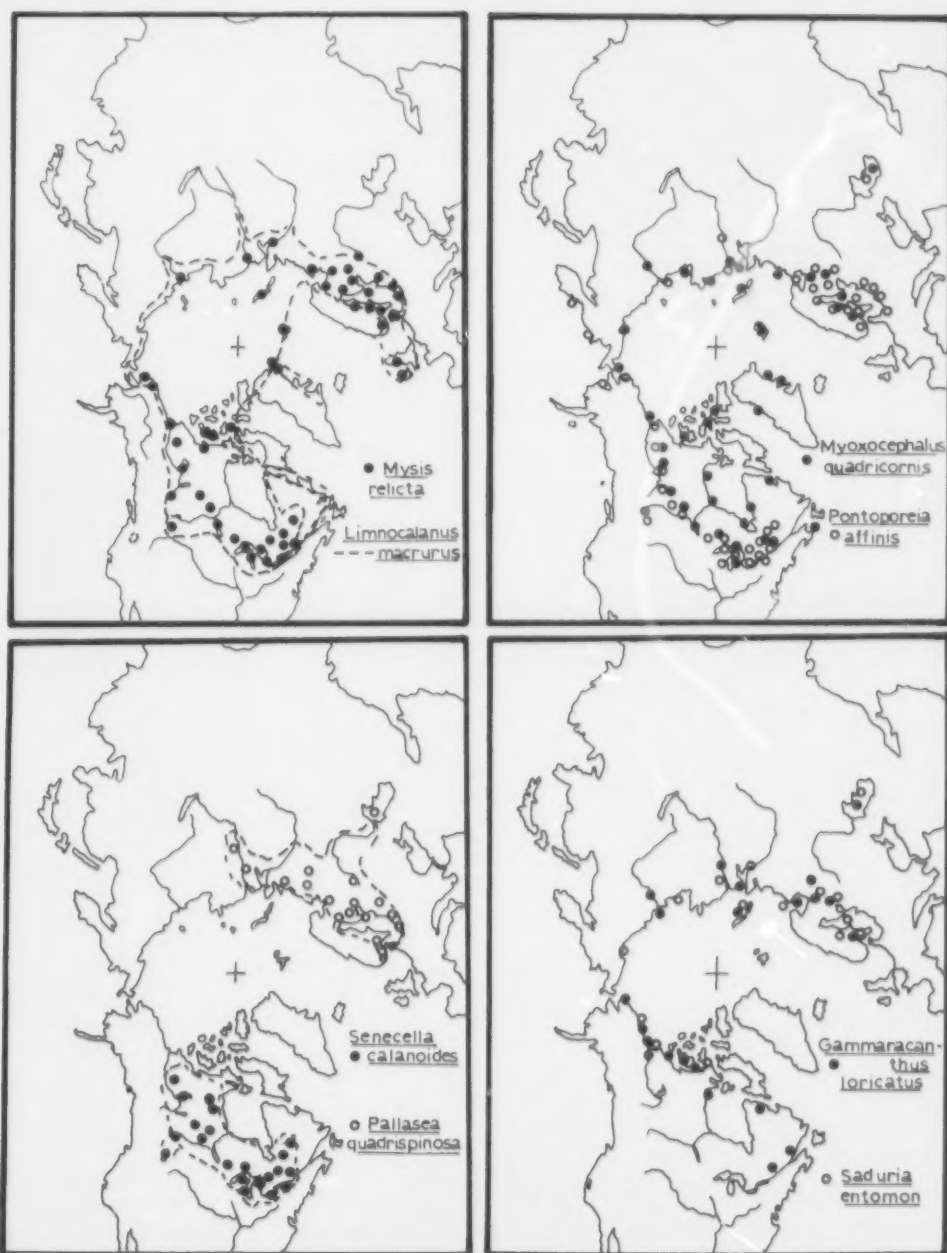
parently continuous character cline links the fresh- and saltwater populations (Lomakina 1952; Johnson 1964). The single specimen that I captured from fresh water in eastern North America (Heart Lake, No. 287) conforms to Lomakina's description (1952) of the estuarine form *G. loricatus aestuariorum* (Gnathopod I/II ratio of 0.92). The Heart Lake specimen differed somewhat from the typical marine *G. loricatus* from the Saguenay estuary, but since its characters were within the range of *loricatus* as given by Lomakina, and since it was an immature animal, I have referred it to *G. loricatus*.

Ekman (1920) and Gurney (1923) considered *Limnocalanus* in fresh water to be a subspecies of the marine *L. grimaldii*. Lindquist (1961) thought the freshwater populations of Eurasia and North America were *L. macrurus*, a distinct species, although there was a continuous character cline linking the freshwater *macrurus* and the marine *grimaldii*. Recently, Holmquist (1970) revised the genus and concluded that only one species, *L. macrurus*, occurs in salt and fresh waters of Eurasia and North America. Regardless of the controversy, *macrurus* is the senior name (Holmquist 1970).

Senecella calanoides is very distinct and has had a stable taxonomic history.

Myoxocephalus quadricornis is considered by most authors to be the same species in salt and fresh waters of Eurasia and North America (Segerstråle 1962; McPhail and Lindsey 1970). McAllister (1961) distinguished the North American freshwater animals as *M. thompsonii* on the basis of the loss of tubercles and body plates, as well as the smaller adult size of the freshwater forms. Intermediate forms, however, exist in arctic lakes with a short history of separation from the sea (McPhail and Lindsey 1970), and Pleistocene fossils of this species in Scandinavia indicate that populations alternately lost and regained bony structures in response to changing salinities in the postglacial Baltic Sea (Segerstråle 1957). It seems reasonable, therefore, to maintain *Myoxocephalus quadricornis* for the North American freshwater populations until this problem has been resolved.

Comparisons among the freshwater populations of *Mysis relicta*, *Limnocalanus macrurus*, *Senecella calanoides*, and *Myoxocephalus quadricornis* from eastern North America revealed little morphological variation be-



Map 1
Distribution of the "glaciomarine relicts" in Eurasia and North America (after Segerstråle 1957; Ricker 1959; Zenkevitch 1963; Johnson 1964; Holmquist

1966). *Pallasea quadrispinosa* is restricted to the fresh waters of Eurasia.

tween themselves or from their accepted descriptions. *Pontoporeia "affinis"* from within the study area, however, was morphologically quite variable. Populations of both *Mysis* and *Pontoporeia* differed in adult size depending on the ecological favourableness of the lake in which they were living. In general the adults were smaller in less favourable localities. It is well known that physicochemical characteristics can affect the growth of freshwater organisms (Jewell 1935; Hutchinson 1967).

Zoogeographical and ecological studies demand good systematics, since closely related species may differ markedly in various aspects of their biology, and confusion of one species with another can lead to erroneous conclusions. In this case, though, what may prove to be a series of taxonomic complexes behaves as a zoogeographical unit, and its dispersal is restricted in a similar way throughout the Holarctic. Therefore, in view of the prevailing taxonomic confusion surrounding these animals, and because there is little knowledge concerning their environmentally induced morphological changes, it seems best at this time to maintain the current nomenclatural usage (with reservations in the case of *P. "affinis"*) when referring to the populations of eastern North America.

Introduction

The importance of former glacial lakes and marine inundations to the dispersal of aquatic organisms during the retreat of the last ice sheet has been stressed by numerous authors. Dymond (1939) related the presence of landlocked smelt and arctic char in lakes of the Ottawa Valley to the intrusion of the Champlain Sea into that area. Many authors have attributed the presence of the goldeye (*Hiodon alosoides*) in northern Ontario and Quebec to the connection that existed between glacial Lakes Agassiz and Barlow-Ojibway (Radforth 1944; McPhail and Lindsey 1970). Turvey (1968) related the distribution of *Diaptomus reighardi* (Copepoda) in Ontario to the known extent of glacial lakes. He found *D. reighardi* in 23 of the 119 lakes he examined that were inside the glacial-lake boundaries, but only in 2 of 107 localities outside the boundaries. Dadswell (1972) showed that the distribution of four species of deep-water fishes in eastern North America was closely associated with the occurrence of glacial lakes. Ricker (1959) suggested that the occurrence of *Mysis relicta* and *Pontoporeia affinis* in North America was related to former glacial-lake coverage. Martin and Chapman (1965) demonstrated the exact relationship between the former extent of glacial Lake Algonquin and the present distribution pattern of the "glaciomarine relicts". Ekman (1920) and Segerstråle (1957) have demonstrated a similar relationship between the distribution of the "relicts" and the extent of former glacial waters in Scandinavia.

Collecting Methods

A wide study area (82°W–57°W, 53°N–41°N) (Map 2) was chosen to provide maximum knowledge of the community's distribution in relation to a number of glacial lakes and marine inundations. Map 2 shows the known maximum extent of all glacial waters formed during the retreat of the last ice sheet, which with some modification is basically the extent established by Prest, Grant, and Ramp-ton (1968).

Fieldwork was carried out between May and December over a four-year period (1969–72). Each region was examined beforehand, using 1:125,000 or 1:250,000 topographic maps, and study lakes were selected according to their geographical situations and eleva-

tions with respect to former glacial lakes and marine shorelines, depth (if known), and accessibility. In areas where glacial-lake and marine shorelines were poorly known, large numbers of lakes were surveyed to delimit the precise upper elevational limit of the deepwater community.

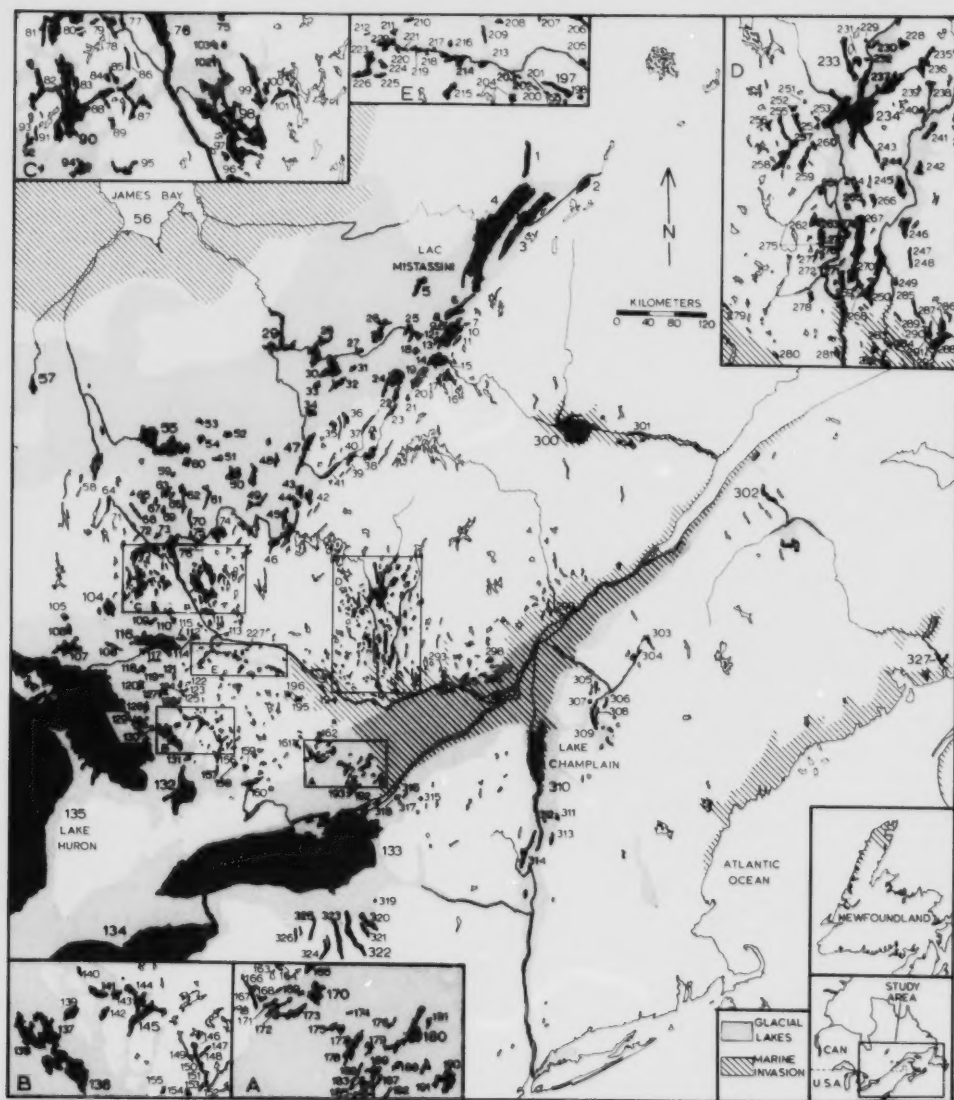
The main sampling gear was a small otter trawl, 2.5 m across the footrope and 5.5 m long (Figure 1). Mesh size was 38 mm throughout, with an inner 2-m bag of 3-mm mesh in the codend. The trawl was towed along the bottom with a ratio of tow rope to depth of 3:1, at speeds from ½–3 kph. This trawl was especially effective in collecting the larger benthic and epibenthic organisms (that is, *Mysis*, *Pontoporeia*, and fish). Trawling was done from a small boat (14 ft) propelled by a 9.5 h.p. outboard motor. In lakes accessible only by airplane, a 12-ft inflatable rubber boat capable of handling the 9.5 h.p. outboard motor was used.

The sampling procedure in each lake was standardized as follows. Vertical temperature and oxygen profiles were taken from the deepest part of the lake, and from these the best trawling depth for the capture of the crustaceans was selected. Two slow-speed tows of 5–7 minutes duration were then made at the selected depth, usually 20–50 m. If the crustaceans were present in the lake, they were usually captured in the first tow. All trawling was done during daylight.

Limnocalanus and *Senecella* were collected with a simple plankton net (No. 6 mesh, mouth diameter 32 cm), using moderately fast (1 meter per second) vertical hauls. At least two hauls were taken in the deepest part of each lake, one from just above the bottom, the second after the net was allowed to settle into the mud for a few seconds. The second haul often captured *Pontoporeia*, and was sometimes the only way to take *Senecella*.

If no *Pontoporeia* were collected with the plankton net or the trawl, two grabs were made with an Ekman dredge (23 × 23 × 23 cm) in depths of 15–20 m.

The otter trawl was used for 614 of the 629 lakes sampled. The additional 15 lakes were sampled with the plankton net only, after initial work had indicated that only *Limnocalanus* occurred in the area. Usually a lake could be sampled in two to three hours; normally two lakes were sampled per day, or as many as seven on days when aircraft were used.



Map 3
Localities (solid lakes and numbers) in which one or more species of the deepwater community have been found. Numbers refer to localities listed in Table 1. Open lakes are ecologically acceptable lakes that lack the community.

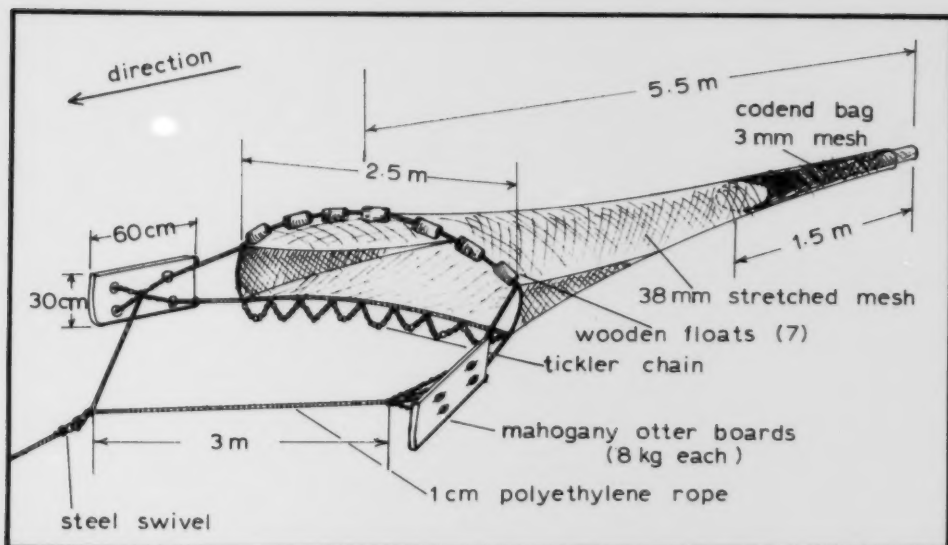


Figure 1
Small otter trawl used to collect *Mysis*, *Pontoporeia*, and the fishes. Netting is made of nylon. Trawl

was towed with a ratio of tow rope to depth of 3:1.

Benthic crustaceans were preserved immediately after capture in 10% formalin and transferred to 70% ethanol within 24 hours. Fish were preserved in 10% formalin, and plankton samples in 5% formalin. Sorting and identification of material were usually done the evening of collection. Organisms other than the crustaceans were referred to specialists for identification.

Collecting Results

One or more of the "glaciomarine relicts" were found in 246 new localities. They are now known from 327 localities east of the 82nd parallel of longitude in North America (Map 3, Table 1).

The distribution of these animals is closely related to the known extent of postglacial waters (Map 3). The community was found in 325 (91%) of the 355 lakes considered to be within the glacial-lake or marine boundaries. It was found in only three (0.6%) localities that may prove to be outside the boundaries (Map 3: Weslemkoon, No. 161; Temiscouata, No. 302; St. John estuary, No. 327), which, however, contained only *Limnocalanus*.

Mysis relicta was captured most often and is now known from 263 localities within the

study area (Map 4A). *L. macrurus* is known from 143 localities (Map 4B), *P. "affinis"* from 151 localities, *G. loricatus* from one freshwater and one saltwater locality (Map 5A), *S. calanoides* from 143 localities, and *Myoxocephalus quadricornis* from one saltwater and 10 freshwater localities (Map 5B). The record of *Gammaracanthus* from Heart Lake (Map 3, No. 287) is the first from fresh water south of the Arctic in North America.

The dispersal ability of other deepwater animals that use, but do not solely depend on, glacial lakes for dispersal (that is, are capable of active upstream movement or passive dispersal), is clearly shown in their present distribution patterns (Maps 6, 7A, B). Approximately 30% of the localities where these animals are known to occur lie outside the glacial-lake boundaries (Dadswell 1972). *Pisidium conventus*, which is commonly found in association with the deepwater community and has been considered part of this group by some authors (Henson 1966), is particularly capable of dispersal beyond glacial-lake boundaries (Map 7A). So far it is the only member of the Great Lakes profundal community (*sensu* Henson 1966) that is known from Newfoundland.

Certain locality records that have persisted in the literature for some time must be discarded. The record of *Pontoporeia* (Norton 1909) from Chamberlain Lake, Maine (Map 20, No. 667), is apparently wrong. Intensive sampling by me and by biologists of the Maine Fish and Game Department failed to find any of the deepwater community in this lake. It is possible that the specimen of *Pontoporeia* supposedly from this lake was actually taken by Kendall (the collector of the specimen) from Lake Champlain, where *Pontoporeia* is known to occur (Henson, Potash, and Sundberg 1966), and was mislabeled, or the Champlain and Chamberlain labels were confused.

The record of *Mysis* from Upper Saranac Lake, New York (Map 20, No. 694) (Greene 1930), is apparently also wrong. Intensive collecting in this lake failed to discover any members of the deepwater community. This lake, however, contains a massive population of *Chaoborus*, and they may have been mistaken for *Mysis*. When these midges are found in fish stomachs (source of the original record), their transparent bodies with the large black hydrostatic organs look superficially like chewed-up mysids.

The population of *Pontoporeia*, referred to as *P. affinis* by Ricker (1959) and known from one locality in the St. Lawrence estuary (Bousfield 1955), is now considered to be distinct from the North American freshwater form (Bousfield, personal communication). This form is apparently adapted to conditions of high salinity (20–30‰) (Bousfield and Laubitz 1972), and so far as is known, it has been unable to establish populations in fresh water in eastern North America.

The validity of the *Senecella* record from Lake Ontario (Davis 1966) is in doubt. The original and only record of it from this lake came from a fish's stomach. Intensive sampling by Patalas (1969) and some collections examined by Juday (1925) failed to reveal *Senecella*. It should be considered as absent from this lake until clearly shown to be present.

Thirty-seven of the lakes sampled by Martin and Chapman (1965) were rechecked, and my results were in close agreement with theirs. I did not find the animals in any lake in which they did not find them, and more intensive sampling revealed only one new crustacean species in each of three lakes.

No discrepancies occurred between my findings and the published records from other lakes that were rechecked — Simcoe, Champlain, Keuka, George, etc. (Ricker 1959).

To check the efficiency of my collecting methods, 55 of the study lakes were visited two or more times. In only eight lakes (14%) were species of the deepwater community captured that were not obtained during the first visit, and no additional species were taken in any lake after the second visit (Table 2). The only three study lakes where the community is known, and in which at least one species was not found during the first visit, had very low levels of hypolimnion oxygen and contained very small populations of the crustaceans. I was able to capture crustaceans from this type of lake only because of the efficiency of the otter trawl. Most of the lakes inside the glacial-lake-marine boundaries in which I was unable to find any of the crustaceans had very low levels of hypolimnion oxygen (Appendix 1, Nos. 328–57). This type of lake was avoided when sampling outside the glacial-lake-marine boundaries.

Three of the lakes in which only *Limnocalanus* was found (Saint-Jean, Temiscouata, Aylmer) were trawled extensively (5–10 times) at all depths to make certain the other crustaceans did not occur there.

Table 1

Table 1

Localities in which one or more of the six "glaciomarine relicts" were found

Number of locality indicates position of record on Map 3. Plus sign (+) indicates presence of species; minus sign (-) indicates species not found. Unless otherwise specified, all localities are lakes. Previously published records for the Algonquin

Park area are from Martin and Chapman (1965); for Stony Lake from Lasenby (1971); for the Finger Lakes from Birge and Juday (1921); for the Great Lakes from Ricker (1959) and Davis (1966); and for Lake Nipissing from Langford (1938).

No. Locality	<i>Mysis relicta</i>	<i>Pontoporeia "affinis"</i>	<i>Limnocalanus macrurus</i>	<i>Senecella calanoides</i>
1. Baudeau	+	-	-	+
2. Bethoulat	+	-	-	-
3. Albanel	+	-	+	-
4. Mistassini	+	+	+	+
5. Opataca	+	-	-	-
6. Waconichi	+	+	+	-
7. Lymburner	+	+	+	-
8. Bourbeau	+	+	-	-
9. Gilman	+	+	+	-
10. Chibougamau	+	+	+	+
11. Armitage	+	-	-	-
12. Antoinette	+	+	-	+
13. Caché	+	+	-	-
14. Obatogamau	+	-	+	-
15. Malo	-	-	+	-
16. Rohault	-	-	+	-
17. Gabriel	+	-	+	-
18. Presqu'île	+	-	+	-
19. Caopatrina	+	-	-	-
20. de la Surprise	+	+	+	+
21. Sylvie	+	-	-	-
22. Hébert	+	-	-	-
23. Lacroix	+	-	-	-
24. Doda	+	+	+	+
25. Opemisca	+	-	-	+
26. la Trêve	+	-	-	+
27. Renault	+	-	-	-
28. Goeland	+	-	-	-
29. Mattagami	+	-	-	-
30. Waswanipi	+	-	+	+
31. Bachelor	+	-	-	-
32. Pustimica	+	+	+	+
33. Madeleine	+	-	-	-
34. Quevillon	+	-	-	-
35. Cuvillier	+	-	-	-
36. Wetetnagami	+	+	-	+
37. Maseres	+	+	-	+
38. Megiscane	+	-	-	-
39. Valmy	+	-	-	-
40. Berthelot	+	-	-	-

Distribution in Eastern North America

No.	Locality	<i>Mysis</i> <i>relicta</i>	<i>Pontoporeia</i> "affinis"	<i>Limnocalanus</i> <i>macrurus</i>	<i>Senecella</i> <i>calanoides</i>
41.	Faillon	+	-	-	+
42.	Matchi-Manitou	+	+	-	+
43.	Gueguen	+	-	-	-
44.	Villebon	+	-	-	+
45.	Granet	+	-	-	-
46.	Grand Victoria	+	-	-	-
47.	Parent	+	-	-	+
48.	Fiedmont	+	-	-	+
49.	Mourier-Lemoine	+	-	-	-
50.	Priessac	+	-	-	-
51.	Lois	+	-	-	-
52.	Chicobi	+	-	-	-
53.	Turgeon	+	-	-	-
54.	Macamic	+	-	-	-
55.	Abitibi	+	-	-	-
56.	James Bay*†	?	?	?	?
57.	Remi	+	-	-	-
58.	Papakomeka	+	-	-	-
59.	Clarice	+	-	-	-
60.	Duparquet	+	-	-	-
61.	Caron	+	-	+	+
62.	Opasatica	+	+	+	+
63.	Larder	+	-	-	-
64.	Mistinikon	+	-	-	-
65.	Round	+	-	-	+
66.	Raven*	+	+	+	-
67.	St. Anthony	+	-	-	-
68.	Kinogami	+	-	+	-
69.	Wendigo	+	+	+	-
70.	Rémigny	+	+	+	+
71.	Mountain	+	-	+	+
72.	Mendelssohn	+	-	-	+
73.	Hammond	+	+	-	-
74.	Simard	+	-	-	+
75.	des Quinze	+	-	+	+
76.	Temiskaming	+	+	+	+
77.	Gilles	+	+	-	-
78.	Bay	+	-	+	+
79.	Kitt	+	-	-	+
80.	Barter	+	-	-	+
81.	Lady Evelyn	-	-	+	-
82.	Obabika (upper)	+	+	-	+
	(lower)	+	-	-	-
83.	Kokoko	+	-	-	-
84.	Kanichee	+	-	+	-
85.	Net	+	-	+	+
86.	Rib	+	-	-	-
87.	Rabbit	+	-	+	+
88.	Herridge	+	-	-	+
89.	Jumping Caribou	+	-	-	+
90.	Temagami	+	+	-	+

Table 1

No.	Locality	<i>Mysis relicta</i>	<i>Pontoporeia "affinis"</i>	<i>Limnocalanus macrurus</i>	<i>Senecella calanoides</i>
91.	Manitou	+	+	-	-
92.	Waswiakashi	+	+	+	-
93.	Cucumber	+	-	-	-
94.	Red Cedar	+	-	-	+
95.	Marten	+	-	-	+
96.	Beauchêne	-	-	-	+
97.	Tee	+	+	+	+
98.	Kipawa	+	+	+	+
99.	Ostabonique	+	-	-	+
100.	Cooks	+	+	-	+
101.	Pomeroy	+	-	-	+
102.	Saint-Amand	+	+	-	+
103.	Argentier	+	+	+	-
104.	Wanapitei	+	+	-	+
105.	Windy	+	+	-	+
106.	Fairbank*	+	+	-	+
107.	Panache	+	-	+	+
108.	Trout	+	-	-	+
109.	Tomiko	+	-	-	+
110.	Tilden	+	-	-	+
111.	Marin	+	+	+	+
112.	Talont†	+	+	+	+
113.	Smith†	+	-	+	+
114.	Turtle†	+	+	+	+
115.	Trout	+	+	-	+
116.	Nipissing†	+	-	+	-
117.	Restoule	+	-	-	+
118.	Waukimakog	+	-	+	+
119.	Fowke	+	+	-	-
120.	Wawashkesh	+	+	-	+
121.	Eagle	-	-	-	+
122.	Bernard	+	+	-	-
123.	Sand†	+	+	-	+
124.	Pickarel	+	-	+	-
125.	Bethunet	+	-	-	+
126.	Doe	+	-	+	+
127.	Ahmict†	+	-	+	+
128.	Manitouwabing	+	-	-	+
129.	Mill	+	-	+	+
130.	Blackstone	+	+	-	+
131.	Kashe	+	-	-	-
132.	Simcoe	+	-	-	-
133.	Ontario*†	+	+	+	-
134.	Erie*†	+	+	+	-
135.	Huron*†	+	+	+	+
136.	Muskoka	+	-	-	+
137.	Rousseau	+	-	-	+
138.	Joseph	+	+	-	+
139.	Skeleton	+	+	-	-
140.	Buck	+	-	-	-

Distribution in Eastern North America

No.	Locality	<i>Mysis relicta</i>	<i>Pontoporeia "affinis"</i>	<i>Limnocalanus macrurus</i>	<i>Senecella calanoides</i>
141.	Vernon	+	-	-	+
142.	Mary	+	+	-	+
143.	Fairy	+	-	-	+
144.	Peninsula	+	-	-	+
145.	Lake of Bays	+	+	-	+
146.	Halls	-	-	+	-
147.	Maple†	+	+	-	+
148.	Beech	-	+	-	+
149.	Boshkung	+	+	+	+
150.	Twelvemile†	+	+	+	+
151.	Mountaint†	+	+	+	+
152.	Horseshoe	-	+	+	+
153.	Minden	+	+	+	+
154.	Bob	+	+	-	-
155.	Big Trout†	+	-	-	-
156.	Gull	+	+	+	+
157.	Shadow	-	-	+	-
158.	Crystal	+	+	+	-
159.	Mississagua	-	-	+	-
160.	Stony†	+	-	+	-
161.	Weslemkoon	-	-	+	-
162.	Mackie	+	-	-	-
163.	Wensley	+	-	-	-
164.	Big Ohlman	+	-	-	-
165.	Palmerston	+	+	-	-
166.	MacKavoy	+	+	-	-
167.	Mazinaw	+	+	+	+
168.	Shabomeka	+	+	-	-
169.	Plevna	+	-	+	-
170.	Cross	+	+	+	-
171.	Mississagagon	+	+	-	-
172.	Kaswakamak	+	+	-	+
173.	Clarendon	+	+	+	-
174.	Silver	-	-	+	-
175.	Sharbot	+	+	-	-
176.	Pike	+	+	-	-
177.	Crowe	+	+	+	-
178.	Bobs	+	+	+	-
179.	Wolfe	-	+	-	-
180.	Big Rideau	+	+	+	+
181.	Otter	-	+	-	-
182.	Canoe	+	+	+	-
183.	Desert	+	+	+	-
184.	Knowlton	+	+	-	-
185.	Birch	+	+	+	-
186.	Clear	+	+	-	-
187.	Buck	+	+	+	-
188.	Indian	-	-	+	-
189.	Devil	+	+	+	-
190.	Charleston (BW)	+	+	+	-
	(RB)	+	+	+	+

Table 1

No.	Locality	<i>Mysis relicta</i>	<i>Pontoporeia "affinis"</i>	<i>Limnocalanus macrurus</i>	<i>Senecella calanoides</i>
191.	Red Horse	+	+	+	-
192.	Loughbourough	+	-	-	-
193.	Sydenham	-	+	-	-
194.	Muskrat	+	+	+	-
195.	Golden	+	-	-	+
196.	Round	+	+	-	+
197.	Grand†	+	-	+	+
198.	St. Andrews†	+	-	+	+
199.	Guthrie†	+	+	+	+
200.	Little Carcajou†	-	+	+	+
201.	Carcajou†	+	-	+	+
202.	Greenleaf†	+	-	+	+
203.	Eustache†	+	+	-	+
204.	White Partridge†	+	+	-	+
205.	Cartier†	-	-	-	+
206.	Ottawa River	+	-	+	+
207.	McSourley†	+	+	-	-
208.	Waterloo†	+	+	-	-
209.	Wendigo†	+	-	-	+
210.	Papineau†	+	-	-	-
211.	Lauder†	+	-	+	+
212.	Guillemette†	+	-	-	+
213.	Radiant†	+	-	-	+
214.	Cedar*†	+	+	-	+
215.	Hogan†	+	-	-	+
216.	Gilmour†	+	+	-	-
217.	Laurie†	+	+	-	+
218.	Gouinlock†	+	+	-	-
219.	Cauchon†	+	+	-	-
220.	Whitebirch†	+	-	-	-
221.	Mink†	+	+	+	+
222.	Kioshkokwi†	+	+	-	+
223.	Wilkes†	+	+	-	+
224.	Threemile†	+	+	-	-
225.	Biggar†	+	+	-	+
226.	Waskigomog†	+	+	-	+
227.	Big Gibson†	+	-	-	-
228.	Nottawissi	+	-	+	+
229.	Kettle	+	+	-	+
230.	Crevier	+	-	-	-
231.	Marguerite	+	+	-	+
232.	Cobble	-	-	-	+
233.	Petawaga	+	+	-	+
234.	Baskatong	+	+	+	+
235.	Chopin	+	-	-	+
236.	Polonais	+	-	+	+
237.	Piscatosine	+	-	+	+
238.	Tapani	-	-	+	-
239.	Chinard	-	-	+	-
240.	Gravel	-	-	+	-

Distribution in Eastern North America

No.	Locality	<i>Mysis</i> <i>relicta</i>	<i>Pontoporeia</i> "affinis"	<i>Limnocalanus</i> <i>macrurus</i>	<i>Senecella</i> <i>calanoides</i>
241.	Moreau	-	-	+	-
242.	des Écorces	-	-	+	-
243.	Pope	-	-	+	-
244.	Gatineau	-	-	+	-
245.	des Îles*	-	-	+	-
246.	du Cerf	-	-	+	-
247.	Corbeau	-	-	+	-
248.	Serpent	-	-	+	-
249.	O'Neil	-	-	+	-
250.	Poisson Blanc	-	-	+	-
251.	Leamy	+	-	-	+
252.	Serpent	+	-	+	-
253.	Quinn	+	-	-	-
254.	de la Vieille	+	-	+	-
255.	Tomasine	+	-	+	+
256.	Savary	+	-	+	-
257.	Rond	+	-	+	+
258.	Désert	+	-	+	+
259.	Bras Coupé	+	-	+	-
260.	Lytton	+	-	+	-
261.	Pocknock	+	+	-	+
262.	Grand Cèdres	+	+	-	+
263.	Petit Cèdres	+	+	-	+
264.	Murray	+	+	+	+
265.	à l'Achigan	+	+	-	-
266.	Kensington	+	+	-	-
267.	Thirty-one Mile*	+	+	+	+
268.	Vert	-	-	+	+
269.	Bangall	-	-	+	-
270.	Pemichangan (A)	+	+	+	-
	(B)	+	+	+	+
271.	Heney*	+	+	+	-
272.	Bitobi	+	-	+	+
273.	Cameron	+	+	+	+
274.	Roddick*	+	+	+	+
275.	Blue Sea	+	+	+	+
276.	Paquin	+	+	+	-
277.	Profond	-	+	-	-
278.	Danford	+	+	-	-
279.	Gruice	+	+	-	-
280.	Thorne	-	+	-	-
281.	Johnston	-	+	-	-
282.	MacGregor	-	+	-	-
283.	McFee	-	-	+	-
284.	Dodds	-	-	+	-
285.	Echo	-	-	+	-
286.	Saint-Sixte	-	-	+	-
287.	Heart (GL)	-	-	+	-
288.	Britannique	-	-	+	-
289.	Lady	-	-	+	-
290.	Hawk	-	-	+	-

Table 1

No.	Locality	<i>Mysis relicta</i>	<i>Pontoporeia "affinis"</i>	<i>Limnocalanus macrurus</i>	<i>Senecella calanoides</i>
291.	Gull	-	-	+	-
292.	la Blanche	-	-	+	-
293.	Papineau	-	-	+	-
294.	Whitefish	-	-	+	-
295.	Louisa	+	+	-	-
296.	Barron	+	+	-	-
297.	Connelly	+	+	-	-
298.	l'Achigan	+	+	-	-
299.	des Piles	-	-	+	-
300.	Saint-Jean	-	-	+	-
301.	Saguenay estuary (GL)	-	-	-	-
302.	Temiscouata	-	-	+	-
303.	Saint-François	-	-	+	-
304.	Aylmer	-	-	+	-
305.	Grand Brompton	+	+	-	-
306.	Massawippi	+	+	+	-
307.	Orford	-	+	-	-
308.	Lovering	-	+	-	-
309.	Memphremagog	+	+	+	+
310.	Champlain	+	+	+	+
311.	Dunmore	+	-	-	-
312.	Sunset	-	+	-	-
313.	St. Catherine	+	+	-	-
314.	George	+	+	-	-
315.	Sylvia	+	+	-	-
316.	Lake of the Woods	+	+	-	-
317.	Sixberry	+	+	-	-
318.	Millsite	+	+	-	-
319.	Green†	+	-	-	-
320.	Skaneateles†	+	+	-	+
321.	Owasco†	+	+	-	+
322.	Cayuga†	+	+	-	+
323.	Seneca†	+	+	-	+
324.	Keuka	+	+	-	-
325.	Canadaigua	+	+	+	-
326.	Hemlock	+	-	-	-
327.	St. John estuary	-	-	+	-

* indicates *Myoxocephalus quadricornis* found in locality.

† indicates locality not sampled by the author, records from literature.

(GL) indicates *Gammaracanthus loricatus* found in locality.

Table 2

Efficiency of collecting methods: Species in the deepwater community found during the first and subsequent samplings of certain lakes

Numbers following the name of each lake are total species found in that lake on each visit. Numbers in brackets are previously unrecorded species found during the second or subsequent visit.

No.: Base-map lake number (see Map 20).

No.	Locality	1	2	Visit 3	4	5
3.	Albanel	2	2(0)	2(0)		
4.	Mistassini	4	4(0)			
14.	Obatogamau	2	2(0)			
15.	Malo	1	1(0)			
41.	Faillon	2	2(0)			
63.	Larder	1	1(0)			
82.	Obabika	1	3(2)			
90.	Temagami	3	3(0)			
98.	Kipawa	4	4(0)			
122.	Bernard	2	2(0)			
143.	Fairy	2	2(0)	2(0)	2(0)	2(0)
167.	Mazinaw	4	4(0)			
176.	Pike	0	2(2)			
179.	Wolfe	0	1(1)			
180.	Big Rideau	3	4(1)	4(0)		
182.	Canoe	3	3(0)			
189.	Devil	3	3(0)			
190.	Charleston	4	4(0)	4(0)	4(0)	4(0)
195.	Golden	2	2(0)			
196.	Round	3	3(0)	3(0)	3(0)	
242.	des Ecorces	1	1(0)			
259.	Bras Coupé	2	2(0)	2(0)	2(0)	
263.	Petit Cèdres	3	3(0)	3(0)		
270.	Pemichangan	4	4(0)			
271.	Heney	4	4(0)	4(0)		
274.	Roddick	5	5(0)			
275.	Blue Sea	4	4(0)			
276.	Paquin	2	3(1)	3(0)	3(0)	
278.	Danford	1	2(1)			
282.	MacGregor	1	1(0)	1(0)		
300.	Saint-Jean	1	1(0)			
302.	Temiscouata	1	1(0)			
304.	Aylmer	1	1(0)			
305.	Grand Brompton	2	2(0)			
306.	Massawippi	3	3(0)	3(0)		
308.	Lovering	0	1(1)			
309.	Memphremagog	3	4(1)			
373.	Nicabeau	0	0(0)			
376.	Argenson	0	0(0)			
416.	Gowganda	0	0(0)			

Table 2

No.	Locality	Visit				
		1	2	3	4	5
430.	Upper Twin	0	0(0)			
454.	Caughnawana	0	0(0)			
515.	Stevens	0	0(0)			
519.	aux Vers	0	0(0)			
536.	Caroline	0	0(0)			
570.	Meach	0	0(0)			
577.	Grand	0	0(0)			
621.	la Pêche	0	0(0)			
645.	Kenogami	0	0(0)			
667.	Chamberlain	0	0(0)			
669.	Megantic	0	0(0)			
674.	Bowker	0	0(0)			
675.	Stukely	0	0(0)			
676.	Lyster	0	0(0)			
714.	Western Brook	0	0(0)			

Number of lakes in which additional species of the deepwater community were found during the second or subsequent sampling trip: $8/37 = 21\%$
 Efficiency of sampling methods, that is, trips without additional species/total trips: $82/90 = 91\%$

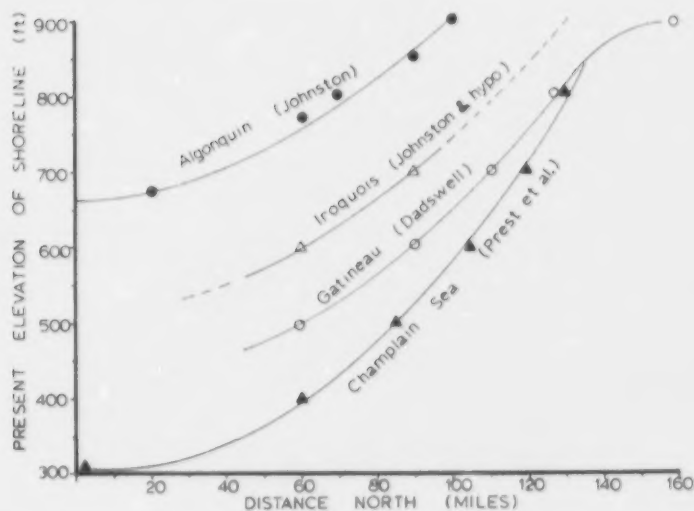


Figure 2
 Warped shorelines of various glacial waters. Champlain Sea is after Prest, Grant, and Rampton (1968). The Iroquois shoreline is extrapolated parallel to the Algonquin shoreline (after Johnston 1916). The Gatineau isobases of maximum elevation of the community occurrence do not conform to the normal shape of a rebound curve.

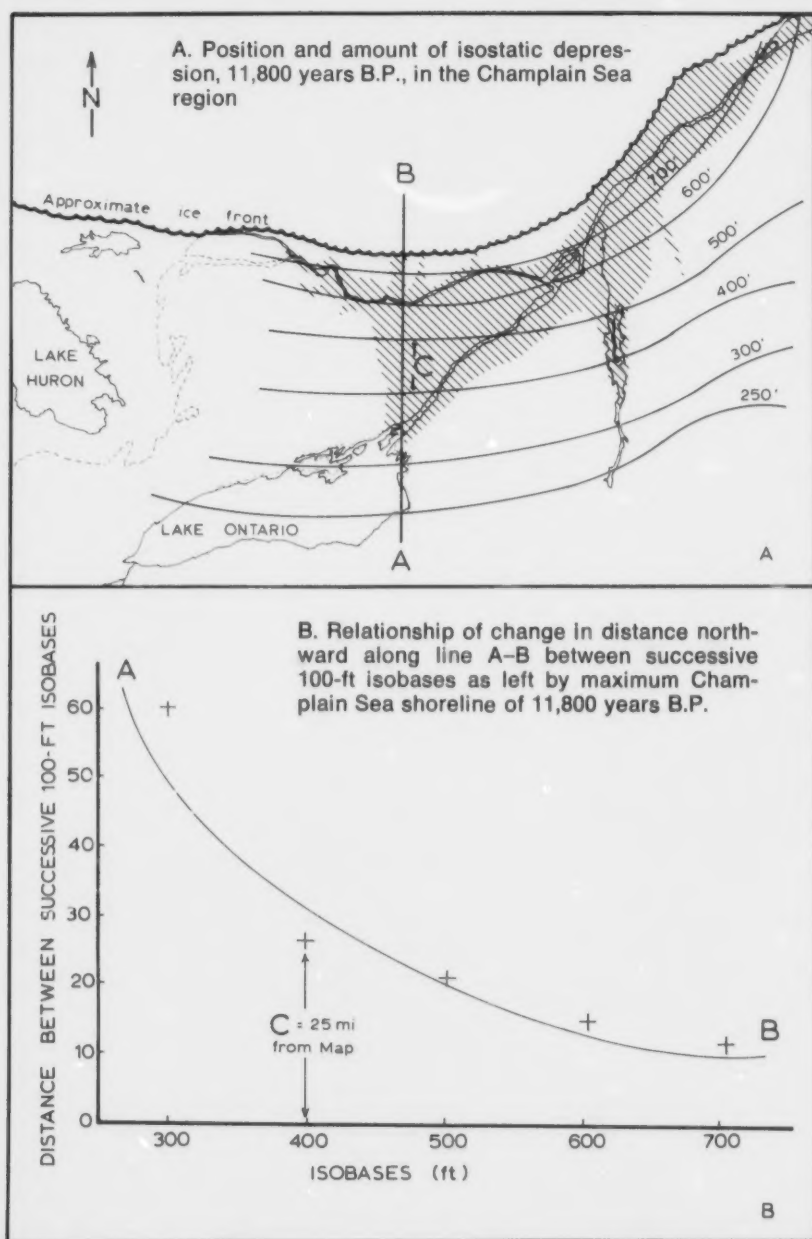


Figure 3
 A. Isobases of the Champlain Sea shoreline, 11,800 B.P. (after Prest, Grant, and Rampton 1966).
 B. Isobase-distance relationship (in miles) northward in the maximum stage of the Champlain Sea.

The Gatineau Valley Problem

The discovery of the deepwater community in the Gatineau Valley in western Quebec (Delisle and Van Vliet 1968) presented a special problem because the extent of glacial lakes in this region has not been mapped by geologists. Antevs (1928) reported glacial-lake sediments around Maniwaki, and various authors have mentioned that the Champlain Sea had penetrated into the valley (Mauffette 1948; Bickel 1970; Gadd 1971), but this is the extent of geological knowledge.

The distribution pattern of the community in lakes of the Gatineau Valley is consistent with dispersal through a glacially controlled standing water-body (Map 8). Isostatic depression of the region that formed this water body is indicated by the south-to-north rise in the maximum elevation of the occurrence of the community in lakes, but graphical analysis of the overall depression shows that the Gatineau curve does not fit the normal isostatic rebound curve form (Figure 2) (Andrews 1968; Broecker 1966). This means that the apparent maximum extent of the Gatineau water body was actually a combination of stages formed at different levels and times during ice retreat.

The maximum shoreline of the Champlain Sea, formed about 11,800 B.P.,* is quite well known (Prest, Grant, and Rampton 1968; Scott 1971) and can be used to estimate a rebound curve for the central portion of the study area (Figure 3A). When the south-to-north change in distance between successive 100-ft isobases of sea level at that time is graphed, a normal isostatic rebound curve is obtained (Figure 3B). Good evidence that this rebound curve is real for that moment in time is indicated by an inverse transformation of the values, resulting in a straight line: $Y = 0.02X - 4.19$ ($r = 0.99$) (Figure 4A), much like the one obtained by Lewis (1970) for uplift of Manitoulin Island. The radiocarbon dates of Lowden and Blake (1968, 1970), Mott (1968), and Romanelli (1972) were then used to construct a time-rebound curve for Kingsmere, Quebec (Figure 4B). This curve agrees closely with the Champlain Sea isobase curve. Therefore it is probable that the iso-

base-distance relationship (Figure 3B) for this area holds through time.

Assuming a continuous retreat of the ice sheet at an even rate and an equally smooth isostatic uplift of the region, we can predict the isobase positions at any one time provided we have one dated shoreline in the area (Andrews 1969). When the dated-shoreline levels of Kingsmere are used in conjunction with the isobase-distance relationship in Figure 3B, a series of hypothetical water-planes for various times can be projected up the Gatineau Valley (Map 9). These hypothetical water-planes show very good correlation to the distribution pattern of the deepwater community in the valley.

The *Limnocalanus* Problem

The eastern North American freshwater distribution pattern of *Limnocalanus macrurus* is slightly different from that of the other deepwater crustaceans dispersed by glacial lakes (Map 5B). It was consistently found in lakes at slightly higher elevations and farther eastward than the others. *Limnocalanus macrurus* appears to enter shallower waters habitually, and there is evidence that it may be more light-tolerant than the other crustaceans (MacKay 1924; Wilson 1929). This behaviour may have enabled it to gain access to lake basins formerly connected to the glacial lakes for only a short period by shallow, transient straits.

Perhaps *Limnocalanus* can disperse short distances by passive means. If this were the case, however, one would expect the distribution pattern to be somewhat random and *Limnocalanus* to be found in numerous lakes outside of, and at higher elevations than, the main distribution of the rest of the community. In fact, it is usually found only close to, or just slightly higher than, the known glacial-lake shoreline. Weslemkoon Lake (No. 161) and Lac Temiscouata (No. 302) are the two known exceptions. Both occurrences could be the result of human transport. Temiscouata, however, is the lowest lake in elevation in its region and the most likely route through which any glacial waters would have passed. If human and/or other means of passive dispersal are possible, why is Temiscouata the only one of many suitable lakes in the region that contains *Limnocalanus*.

Limnocalanus macrurus is known to survive in, and disperse through, brackish sur-

*Abbreviation for "Before Present". "Present" refers to A.D. 1950, and this abbreviation is to be understood with all datings that follow.

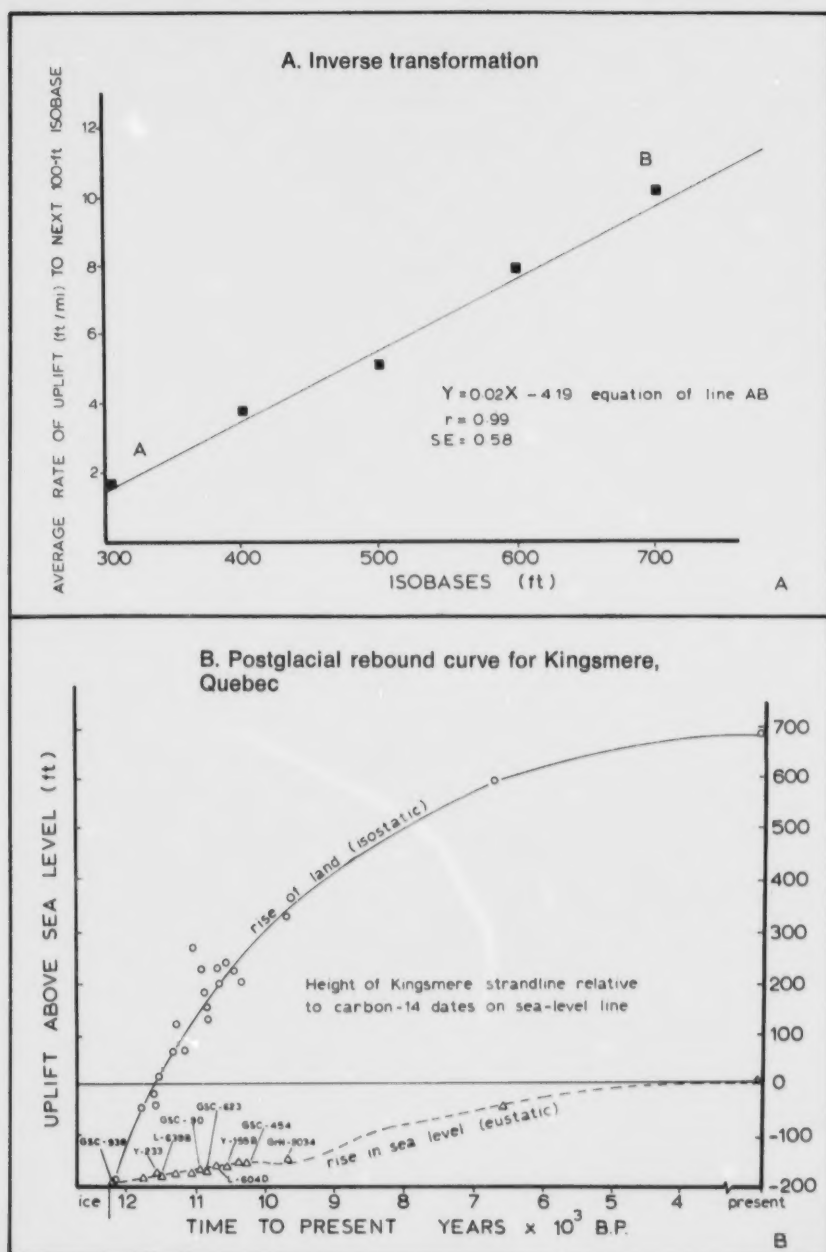


Figure 4
A. Inverse transformation of the isobase-distance relationship in Figure 3B.

B. Time-rebound curve for Kingsmere, Quebec (see Map 9). Eustatic rise in sea level is after Godwin, Suggate, and Willis (1958) and Kenney (1964).

face layers of arctic seas (Grainger 1962, 1965; Holmquist 1970) and to have dispersed widely in the Arctic Ocean (Bowman and Long 1968). This salinity tolerance, as well as its tolerance for light, apparently enabled it to disperse in the early stages of the Champlain Sea and may explain its presence, by itself, in a number of lakes inundated only by the early, highest levels of the sea (for example, Map 3, Lac des Piles, No. 299).

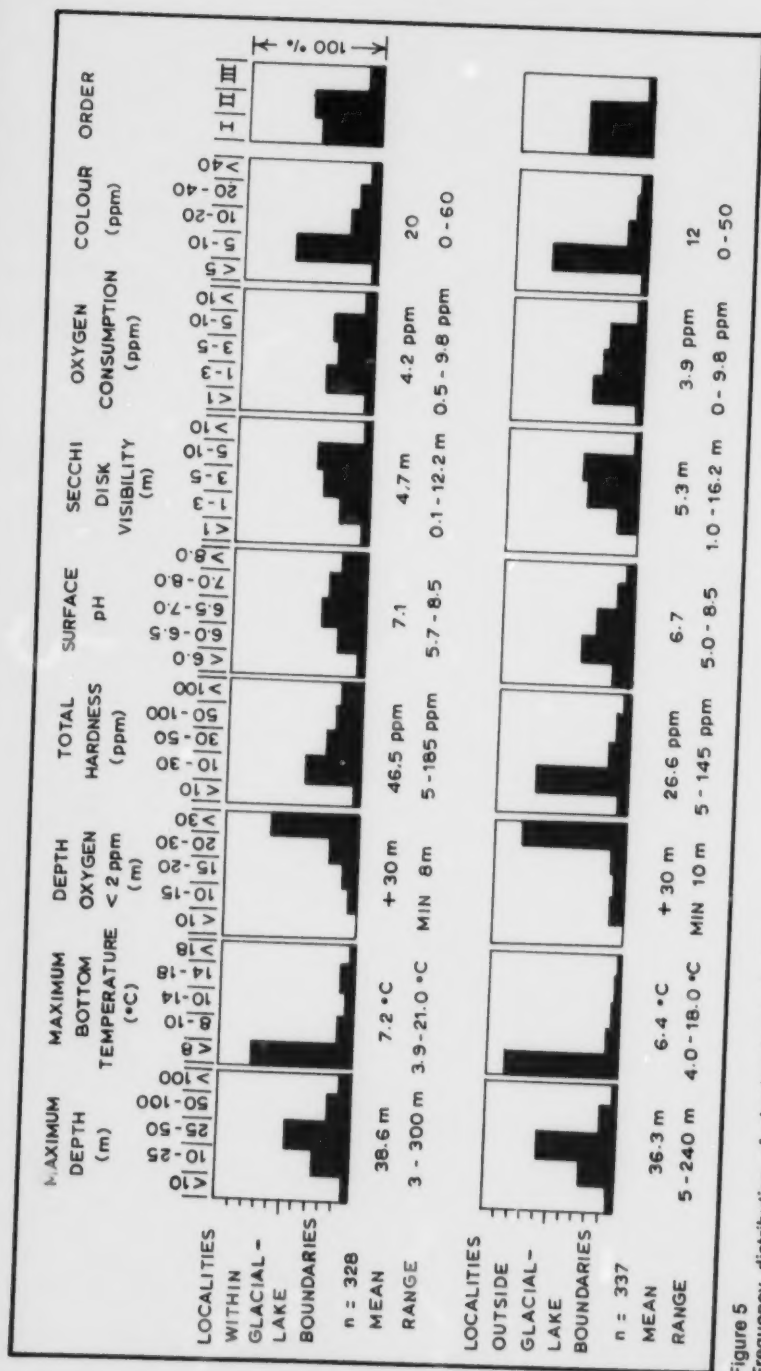


Figure 5

Frequency distribution of physicochemical parameters in the two groups of study lakes. Height of the graphs (that is, the boxes) corresponds to 100% of the lakes in a particular category. The height of the solid bars indicates the percentage of lakes in each category.

Introduction

There are two main reasons why an organism may not occur in a specific locality: either it has never dispersed into the area, or it cannot survive in the particular ecosystem found there. Survival is dictated by the organism's tolerance of its physical and biological environment. Accordingly, the distribution of the deepwater community depends not only on events in the past, but also on conditions of the present-day habitat.

Aquatic biologists generally associate "glaciomarine relicts" with oligotrophic lakes, emphasizing low temperatures and high levels of dissolved oxygen as necessary for their survival (Samter and Wettner 1904; Thienemann 1928; Larkin 1948). Holmquist (1959, 1966) points out that high temperatures and low levels of dissolved oxygen are not especially limiting to these organisms and that other unknown factors seem to limit their distribution, at least within lakes. Rawson (1960) observed that *Pontoporeia affinis* and *Limnocalanus macrurus* were absent from Cree Lake, Saskatchewan, for no apparent reason. Koshinsky (1965) could not explain why *P. affinis* was absent from depths greater than 10 m in one lake when it was abundant in less oxygenated, greater depths in nearby lakes.

It is apparent from numerous studies that other limnological factors can limit the occurrence of aquatic organisms. Tucker (1958) suggested calcium levels in combination with dissolved organic matter as a reason for the absence of leeches, crustaceans, and molluscs from certain lakes. Delorme (1965) found that pH limits the distribution of ostracods, and Carter (1971) obtained similar results from planktonic crustaceans. Green (1971) used a combination of physical and chemical factors to explain the differences between molluscan communities in some Manitoban lakes.

Physicochemical parameters of study lakes were determined for two reasons: (1) to ascertain whether lakes without glacial-lake inundation differed significantly from lakes that were inundated, and whether this difference has influenced the community's distribution; and (2) to determine the limiting factors of the individual crustacean species.

Holmquist (1966) seems to attribute the different distribution patterns of the individual "relict" species to their dispersal history. In

eastern North America it is more likely that dispersal of the deepwater community as a whole was similar (except for the slight variation for *Limnocalanus*), but that present-day limnological conditions have caused one or more species to disappear from certain lakes.

Methods

Physicochemical parameters were measured for each study lake. To check the accuracy of field determinations and obtain values for oxygen consumption and colour, one-liter samples of surface water were collected from 150 selected lakes, and a complete chemical analysis of them was made by the Inland Waters Directorate of Environment Canada. Physicochemical data on lakes not sampled were obtained from the literature (15 lakes), or from the records of regional fishery biologists (22 lakes). Pertinent physicochemical parameters for all lakes are listed in Appendix 1.

Vertical temperature profiles were taken with an Applied Research FT-3 hydrographic thermometer, accurate to 0.5°C. Dissolved oxygen was determined by the azide modification of the Winkler method. Hydrogen ion concentration (pH) was determined at the surface and just above the bottom of each lake, using a Hach comparator (5.5–8.5 Brom thymol blue or 4–10 wide-range indicator). Total hardness (TH) was measured at the surface using the EDTA titration method (Hach kit). Light penetration was measured with a Secchi disk, 20 cm in diameter. Oxygen consumption was determined by the potassium permanganate oxidation method for the 150 lakes analyzed by the Inland Waters Directorate, Environment Canada. Oxygen consumption and colour of these lakes were then plotted against the log of their Secchi visibilities in centimeters, and the oxygen consumption and colour of the remaining study lakes were estimated from these graphs, using the Secchi visibility of the lakes (after Schindler 1971).

Field determinations were in good agreement with laboratory determinations, but averaged 10% higher. This difference may have been due to the storage period of the water samples (up to six months), and for this reason field determinations (except for oxygen consumption) were used in the discussions that follow. Throughout the study, an effort was made to sample lakes suspected

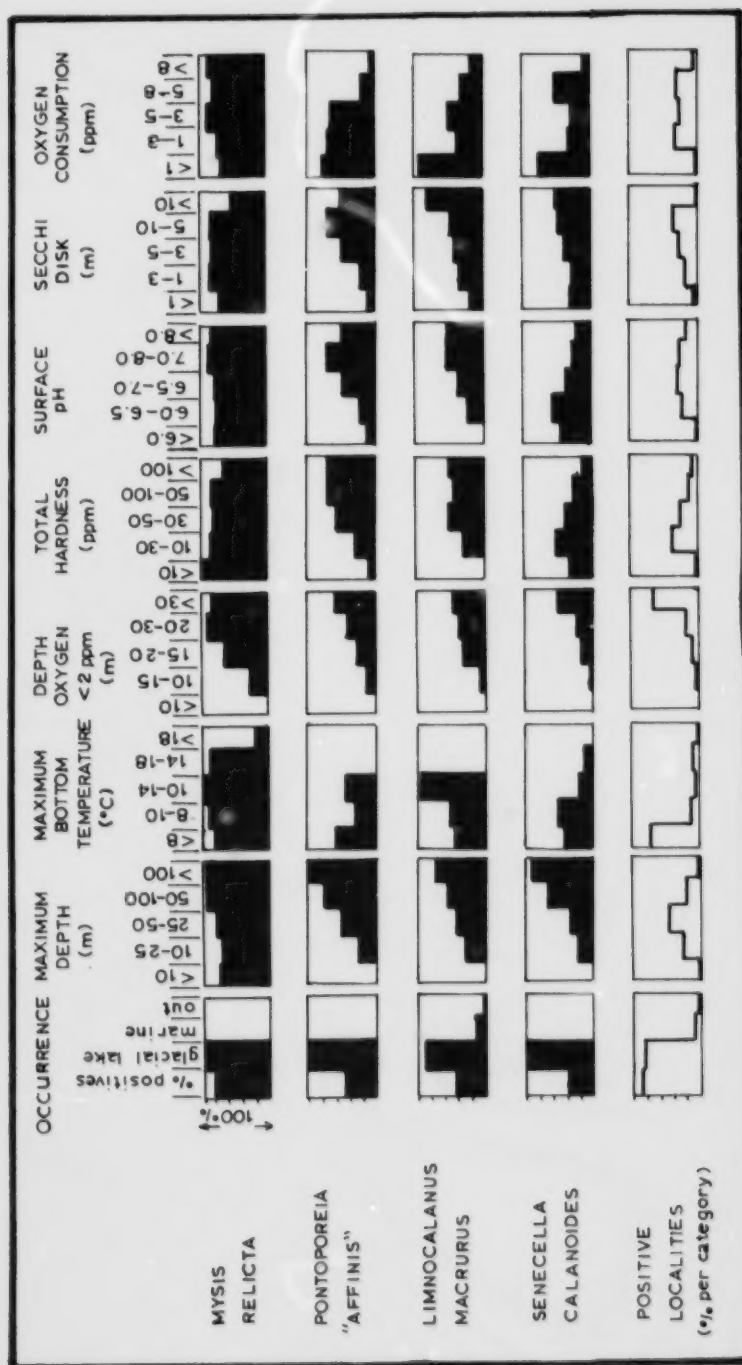


Figure 6
Frequency distribution of the occurrence of the crustaceans in the study lakes in relation to the physicochemical parameters examined. Height of the graphs (that is, the boxes) corresponds to the 100% of the lakes in a particular category. The height of the solid bars indicates the percentage of lakes in each category in which the species were found.

of having low levels of dissolved oxygen when oxygen levels were lowest, in autumn.

Nine lakes were selected on the basis of depth, total hardness, and oxygen consumption, and were sampled for positive occurrence, population density, and depth distribution of *Pontoporeia*. In each lake two or three Ekman grabs were taken at each of 10% increments of hypolimnion depth (that is, 9–10 stations per lake or 20–30 samples per lake). Temperature and pH were determined at each depth. *Pontoporeia* were separated from the mud by sieving in a wash-bucket (11 meshes per cm). Counts from each depth were averaged.

Comparison between "Inside" and "Outside" Lakes

The lakes sampled during this study can be divided into three groups: lakes inside the glacial-lake or marine boundaries that are positive for the community; lakes inside the boundaries that are negative for the community because of unsuitable ecological conditions; and lakes outside the glacial-lake or marine boundaries that are negative for zoogeographical reasons. For the purpose of discussing physicochemical (environmental) limiting factors, the first two groups will be combined and called "inside" lakes, and the third group will be known as "outside" lakes. Physicochemical data were obtained for 328 inside lakes, 313 of which I examined personally. The outside group includes 337 lakes, 317 of which I sampled personally.

Means and ranges of physicochemical conditions for inside and outside lakes were similar (Figure 5). Outside lakes were characterized by lower mean total hardness, lower pH, and clearer water than inside lakes. Inside lakes had slightly higher amounts of dissolved organic matter (oxygen consumption 4.2 ppm vs. 3.9 ppm), which probably caused the lower than expected mean pH for this group. Average surface pH of inside lakes was only slightly higher than that of outside lakes (7.1 vs. 6.7), although average total hardness of inside lakes was significantly higher than for outside lakes ($P < 0.01$). Christman and Ghassemi (1966) found that high organic levels reduced the pH of water.

The higher elevation of the outside lakes is probably the main reason for these differences. Headwater lakes receive less influx of dissolved solids and organics because of

smaller drainage basins (Schindler and Nighswander 1970), and they lack former glacial-lake coverage, the presence of which has been shown to contribute dissolved solids to a lake basin (Ryder 1964). Interestingly enough, outside lakes would probably be categorized as more oligotrophic (Hutchinson 1957), and therefore as better habitat for the deepwater community.

The community as a whole was found to tolerate nearly the complete range of physicochemical conditions existing at present in the inside lakes (Figure 6, Table 3). Although the means of physicochemical factors of inside and outside lakes differed (sometimes significantly), the ranges of outside lake conditions were within the ranges tolerated by the community in the inside lakes (Table 3). Consequently, physicochemical factors cannot be regarded as contributing to the absence of the community from the outside lakes.

Major Environmental Limiting Factors

My findings, however, indicate that environmental conditions can affect the species composition of the crustacean community in a given lake. In this section I will be referring only to the four crustaceans dispersed by glacial lakes (that is, those shown in Plate 1). *Myoxocephalus* and *Gammaracanthus* are not included because they were captured so few times. Also, only lakes with the potential to contain all four crustaceans have been considered. Positive lakes in areas reached only by *Limnocalanus* — for example, the Lièvre River Valley — have been excluded.

It is probable that when each present-day lake basin separated from the glacial lake, conditions in the new lake were oligotrophic, and it contained populations of all four crustaceans. But lakes change through time from oligotrophic to either eutrophic or dystrophic, and this causes a corresponding change in their profundal community (Brundin 1958). Consequently, depending on the original depth of the basin and on whatever chemical changes have occurred in the intervening years, one or more of the crustaceans may have become extinct there. Obviously this is the case in our area, since only 33 of 291 possible lakes were found to contain all four crustaceans (Table 1).

The niche of a species is theoretically defined as an "N-dimensional hypervolume" in

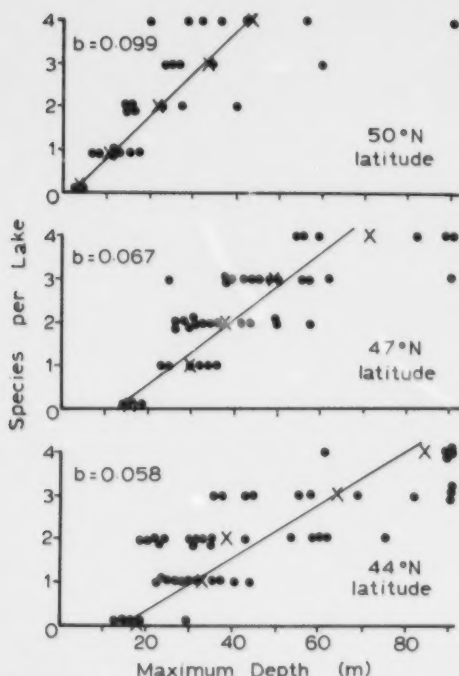


Figure 7
Relationship between maximum depth, latitude, and number of deepwater crustacean species occurring in one lake. Regressions are drawn through mean of depths (X). Slopes of the regressions (b) are significantly different ($P < 0.01$) from each other.

which fitness of individuals is positive (Green 1971). This definition makes it impossible to delimit the hypervolume of a species, and in practice "N" is reduced to a reasonable number of measurable parameters. In this study, seven parameters were chosen, based on the criteria set out by Green (1971).

1 *Maximum depth (z_m)* has been emphasized by various authors as important to the survival of these crustaceans (Ricker 1959; Segerstråle 1957). Martin and Chapman (1965) give 24 m as the minimum depth of the lake in which these crustaceans were found in Algonquin Park, Ontario.

Maximum depth is not an independent parameter within a lake; rather, its effect is to influence maximum bottom temperatures and levels of incipient light. It may also represent a spatial separation factor. Depth, how-

ever, is an easily measurable, independent parameter for each lake and can be used in comparison between lakes.

The occurrence of *Pontoporeia*, *Limnocalanus*, and *Senecella* was positively correlated with deep lakes (Figure 6, Table 4). *Pontoporeia* was found in 100% of lakes deeper than 100 m and in none of the lakes shallower than 10 m. *Mysis* was found as often in shallow lakes (78%) as it was in deeper ones (80–96%).

Maximum depth was the main factor determining the total number of crustacean species in a lake (Figure 7). Shallow lakes or shallow basins of one lake usually contain fewer species of the community than deeper lakes or basins. For example, the deepest basin of Charleston Lake, Running Bay (No. 190), contains four members of the community, the shallower basin, Big Water, only three (Table 1). The minimum depth necessary for the survival of the total crustacean community, and/or the individual species, decreases with an increase in latitude. Johnson (1962) found these crustaceans on Victoria Island, Northwest Territories (70°N), in lakes as shallow as 5 m. In the study area, lakes at the latitude of Chibougamau, Quebec (50°N), usually contain all four species if they are deeper than 20 m, and *Mysis* survives in lakes as shallow as 5 m. In southern Ontario (44°N), only lakes over 60 m deep contained all four species, and the shallowest lake found to contain any of the community was 18 m in depth.

2 *Maximum bottom temperature (T_m)*, above 14°C, has been considered one of the most important factors limiting the occurrence of the deepwater community (Thienemann 1928; Ricker 1959), but my findings indicate this is not wholly true. *Mysis* was found as often in lakes with T_m 's between 14 and 18°C as it was in colder lakes (Figure 6), and its occurrence showed little correlation to temperature (Table 4). But bottom temperatures are limiting once they exceed the upper lethal limit of the individual species.

The warmest bottom temperature at which a *Mysis* population was found surviving was 18°C (Grand Lac Victoria, No. 46). This temperature is considerably higher than values quoted in the literature (Thienemann 1928; Ricker 1959), although Holmquist (1966) gave 16°C as the upper limit known for *Mysis* in Swedish lakes. In my study area, lakes with

T_m 's up to 16°C contained large, healthy populations of *Mysis* (2,000 animals per trawl). *Pontoporeia* and *Limnocalanus* were never found in lakes with bottom temperatures above 14°C (Figure 6). *Senecella* was found a number of times in lakes with a T_m of 15°C. These last three species showed strong negative correlation to warmer temperatures (Table 4).

3 Depth at which dissolved oxygen was less than 2 ppm ($z_{0.2}$) was chosen as a parameter because the depth at which minimum tolerable oxygen levels occur is critical, depending on the transparency of the lake water. Many authors stress low oxygen levels as a factor limiting the occurrence of these crustaceans (Thienemann 1928; Samter and Wettner 1904).

Low levels of hypolimnion oxygen is the only condition that severely limits the occurrence of *Mysis* (correlation: 0.44, Table 4, Figure 6). *Limnocalanus* and *Senecella* were absent from lakes with sharp oxygen stratification, that is, those with no trace of oxygen at the bottom. *Pontoporeia* is very tolerant of

low levels of oxygen (correlation insignificant, Table 4) and was often the only species found in eutrophic lakes, for example, Wolfe Lake, No. 179.

Minimum oxygen values at which these species were found to occur in the study lakes are as follows: *Mysis*, 2 ppm (Red Horse Lake, No. 191, 12 Oct. 1970, 20 m); *Pontoporeia*, 1 ppm (Devil Lake, No. 189, 7 Oct. 1969, 30 m); *Limnocalanus* and *Senecella*, 0.6 ppm (Murray Lake, No. 264, 5 Oct. 1970, 20–23 m). Except for *Senecella*, whose oxygen tolerance is unreported, these values agree closely with those given by other authors (Juday and Birge 1927; Strøm 1946; Lasenby 1971).

4 Total hardness (TH) is the measure of the total content of calcium and magnesium ions in the water (Hutchinson 1957). Calcium is necessary to crustaceans for building exoskeletons. Green (1971) and Tucker (1958) found that low levels of calcium limited the occurrence of aquatic organisms.

Mysis was present less often in lakes with high TH (Figure 6). This, however, may be an artifact, since lakes with high TH commonly

Table 3

Total range of physicochemical parameters found in the study lakes and ranges tolerated by the crustaceans. Additional parameters are from the complete chemical analysis of 150 study lakes.

Abbreviations

TH	Total hardness	HCO ₃	Bicarbonate
SDV	Secchi disk visibility	CO ₂	Dissolved carbon dioxide
O ₂ c	Oxygen consumption		

Parameter	Outside lakes	Inside lakes	<i>Mysis relicta</i>	<i>Pontoporeia "affinis"</i>	<i>Limnocalanus macrurus</i>	<i>Senecella calanoides</i>
Depth (m)	5-240	3-300	5-300	18-300	14-300	14-240
Temperature (°C)	4.0-18.0	3.9-21.0	3.9-18.0	3.9-14.0	3.9-14.0	3.9-15.0
Oxygen (ppm)	12-0	12-0	12-2	12-1	12-0.6	12-0.6
TH (ppm)	5-145	6-185	5-185	8-185	10-185	8-130
Surface pH	5.0-8.5	5.7-8.5	5.7-8.5	6.1-8.5	6.1-8.5	5.7-8.5
SDV (m)	1.0-16.2	0.1-12.2	0.1-10	1.1-10	0.8-12.2	0.8-12.2
O ₂ c (ppm)	0.0-9.8	0.5-9.8	1.0-9.8	1.0-6.7	0.5-8.8	0.5-9.5
Alkalinity (ppm)	0.0-95.9	0.0-118	0.0-118	0.0-118	1.3-96	0.0-96
Calcium (ppm)	1.3-39.7	1.4-38.2	1.4-38.2	2.0-38.2	3.6-32.9	1.4-32.9
Magnesium (ppm)	0.01-2.8	0.08-9.6	0.08-9.6	0.08-9.6	0.1-7.5	0.08-7.5
Potassium (ppm)	0.1-0.9	0.1-2.0	0.1-1.5	0.1-1.3	0.1-2.0	0.1-1.5
Sodium (ppm)	0.2-2.9	0.2-3.2	0.2-3.2	0.4-3.2	0.3-2.9	0.2-3.2
HCO ₃ (ppm)	0.0-117	0.0-144	0.0-144	0.0-144	1.6-118	0.0-118
Chlorine (ppm)	0.0-14	0.0-7.2	0.0-7.2	0.0-7.2	0.0-6.7	0.0-7.2
Sulphate (ppm)	2.6-20.6	3.1-24.3	3.1-24.3	3.6-18.5	3.1-18.5	3.4-16.6
CO ₂ (ppm)	0.1-30.4	0.8-9.2	0.8-9.2	0.8-6.7	0.8-6.4	0.8-8.4

had low levels of hypolimnion oxygen. Occurrence of *Pontoporeia* and *Limnocalanus* was correlated with high TH (Table 4). *Pontoporeia* was found in only 27% of lakes with TH less than 30 ppm (43% of the positive study lakes). In eastern North America the distribution pattern of *Pontoporeia* is closely associated with the distribution of lakes having high TH within the boundaries of the former glacial lakes (Map 10). *Senecella* occurred most often in lakes with low TH (Figure 6, Table 4), but this may also be an artifact caused by competition between *Limnocalanus* and *Senecella* (see below).

5 *Hydrogen ion concentration* is a dominant factor in fresh water, as it affects the deposition of calcium and the availability of the bicarbonate ion (Hutchinson 1957). Delorme (1965) gave 6.1 as the lower limit of pH tolerance for ostracods. Beamish (1970) showed that low pH could inhibit fish reproduction. Surface pH, rather than hypolimnion pH was selected as a parameter because it varies less during the open-water season, allowing for comparison between lakes sampled at different times of the year.

The occurrence of *Mysis* was not restricted by pH within the range studied (Figure 6). The occurrence of *Pontoporeia* and *Limnocalanus* was correlated to high pH (Table 4). Neither of these species was found in lakes with surface pH less than 6.0 (Table 3). *Senecella*, on the other hand, was most common in lakes

with low pH, and was found in 60% of lakes between 6.0 and 6.5 (negative correlation, Table 4, Figure 6).

6 *Secchi disk visibility* (SDV) approximates the 15% level of percentage transmission of surface light intensity (Beeton 1958), and gives a rough measure of the amount of transmitted light reaching deepwater organisms. Light is known to affect the behaviour of these crustaceans (Beeton 1960; Brownell 1970), but it has not been suggested as an absolute limiting factor to occurrence. My findings indicate that strong light penetration, especially in combination with other factors, is very detrimental to the survival of these species.

Mysis was found least often in very clear lakes, particularly if there was an oxygen deficiency in the hypolimnion. *Senecella* and *Limnocalanus* were absent from a number of quite deep, well-oxygenated lakes with high SDVs (Table 1: Clear Lake, No. 186; Skeleton Lake, No. 139). All of these species were found to select deeper strata in clear lakes than in less transparent waters.

7 *Oxygen consumption* (O_2C) is one measure of the dissolved organic matter in water. This organic matter, or "humic acid", consists of acidic phenolic macromolecules originating from the decay of plant material (Christman and Ghassemi 1966). What effect high macromolecule concentrations have on aquatic or-

Table 4

Correlation between physicochemical parameters and the occurrence of the crustacean species. Calculated after logarithmic transformation with 300 degrees of freedom.

Abbreviations

z_m	Maximum depth	TH	Total hardness
T_m	Maximum bottom temperature	pH	Surface pH
zO_2	Depth of dissolved oxygen less than 2 ppm	SDV	Secchi disk visibility
		O_2C	Oxygen consumption

Species	Physicochemical Parameter						
	z_m	T_m	zO_2	TH	pH	SDV	O_2C
<i>Mysis relicta</i>	0.15*	-0.05	0.44*	-0.14*	-0.16*	-0.05	0.04
<i>Pontoporeia "affinis"</i>	0.34*	-0.37*	0.001	0.24*	0.24*	0.38*	0.45*
<i>Limnocalanus macrurus</i>	0.24*	-0.19*	0.11†	0.11†	-0.09†	0.07	0.11†
<i>Senecella calanoides</i>	0.27*	-0.22*	0.34*	-0.29*	-0.27*	0.06	-0.02
<i>Limnocalanus</i> and <i>Senecella</i>	0.22*	0.15*	-0.19*	-0.12†	-0.09†	0.02	0.05

* indicates significant correlation at the 1% level.

† indicates significant correlation at the 5% level.

ganisms is unknown (Shapiro 1957), but it may be possible that they affect the uptake of calcium ions. Tucker (1958) found that lakes with high O_2 contained fewer aquatic species, and that high levels of dissolved organic matter were lethal to flatworms.

Occurrence of *Mysis* shows no significant correlation to O_2 (Table 4). *Pontoporeia* and *Limnocalanus* occurrence was correlated to lakes with low O_2 (Table 4). *Pontoporeia* was seldom captured in lakes with O_2 greater than 5 ppm. *Senecella* occurrence correlated slightly, but not significantly, with lakes having heavily stained water (O_2 greater than 5 ppm) (Figure 6, Table 4).

Multiplication Effect of Limiting Factors

In reality no environmental limiting factor ever functions independently. Each of the preceding factors may place a certain amount of environmental stress on the crustaceans, but the effect can be alleviated or intensified by levels of one or more of the other six factors. Regression analysis indicates that certain major factors account for most of the differences in the distributions of each of the crustaceans (Table 4).

Factors Affecting *Mysis relicta*

Mysis is the most eurytopic of the crustaceans. It occurred in the most localities (263) and was found in nearly every type of lake sampled (Figure 6). *Mysis* was only absent from, or rare in, small clear-water lakes with sharp thermal stratification and low levels of dissolved oxygen in the hypolimnion (for example, Appendix 1: Thorne Lake, No. 280; Orford Lake, No. 307).

Carleton (1972) found that depth distribution of *M. relicta* was related to temperature and incipient light. I found that in a given lake, the bulk of the mysid population selected an optimum depth, balancing minimum incipient light, minimum temperature, and maximum dissolved oxygen. Substantial mysid populations still exist in many lakes with low levels of hypolimnion oxygen because the low level of transparency in their water allows the mysids to occupy shallower depths during the critical low-oxygen periods of the year. For example, in Murray Lake (No. 264, SDV 2.8 m), during October, the mysids can occupy depths of 8 to 12 m, the only part of the hypolimnion with temperatures below 18°C

and oxygen above 2 ppm. On the other hand, in Danford Lake (No. 278, SDV 6.3 m), during autumn, even though temperatures below 18°C were found as shallow as 8 m, no mysids were found in depths less than 18 m (presumably because of high incipient light) nor deeper than 20 m (because of low oxygen). Consequently its mysid population has been reduced almost to extinction.

Mysis relicta reaches its greatest abundance in three types of lakes: shallow unstratified lakes with T_m 's under 16°C and low SDVs (for example, Lac Gueguen, No. 43); moderately deep, stratified lakes with low SDVs and high TH (for example, Lac Bitobi, No. 272); and very deep lakes (for example, Trout Lake, No. 115). These lakes all have in common very low levels of transmitted light, occurring at some depth, in conjunction with optimum temperatures and oxygen.

Factors Affecting *Pontoporeia "affinis"*

Early in this study it became evident that factors other than temperature and oxygen were limiting the occurrence of *Pontoporeia*. Intensive sampling in a few lakes in which these two parameters were not a problem revealed that in fact the amphipod was either absent or very rare. For example, at Fairy Lake, No. 143, 30 Ekman grabs and 6 trawls between 10–70 m captured no *Pontoporeia*.

The relationship between maximum depth, TH, dissolved organic matter, and the occurrence of *Pontoporeia* is demonstrated in Figure 8. Shallow lakes with low TH and high O_2 almost always lack *Pontoporeia*; deep lakes with low O_2 , and high TH always contain it. The absence of *Pontoporeia* from Cree Lake, Saskatchewan (Rawson 1960), was probably due to the TH- O_2 relationship of that lake. Cree had the lowest TH and hypolimnion pH of all the lakes sampled by Rawson. In general, dystrophic lakes seem to provide poor habitat for *Pontoporeia*.

It was also noted that there was a relationship between the adult size of *Pontoporeia* and the favourableness of the lake. For example, the average size of penultimate animals in Mazinaw Lake, No. 167 (TH 36 ppm, O_2 5.3 ppm), was only 6.2 mm, whereas in nearby Canoe Lake, No. 182 (TH 90 ppm, O_2 1.4 ppm), the average size was 9.5 mm. Intermediate sizes between these extremes occurred in lakes with intermediate chemical conditions.

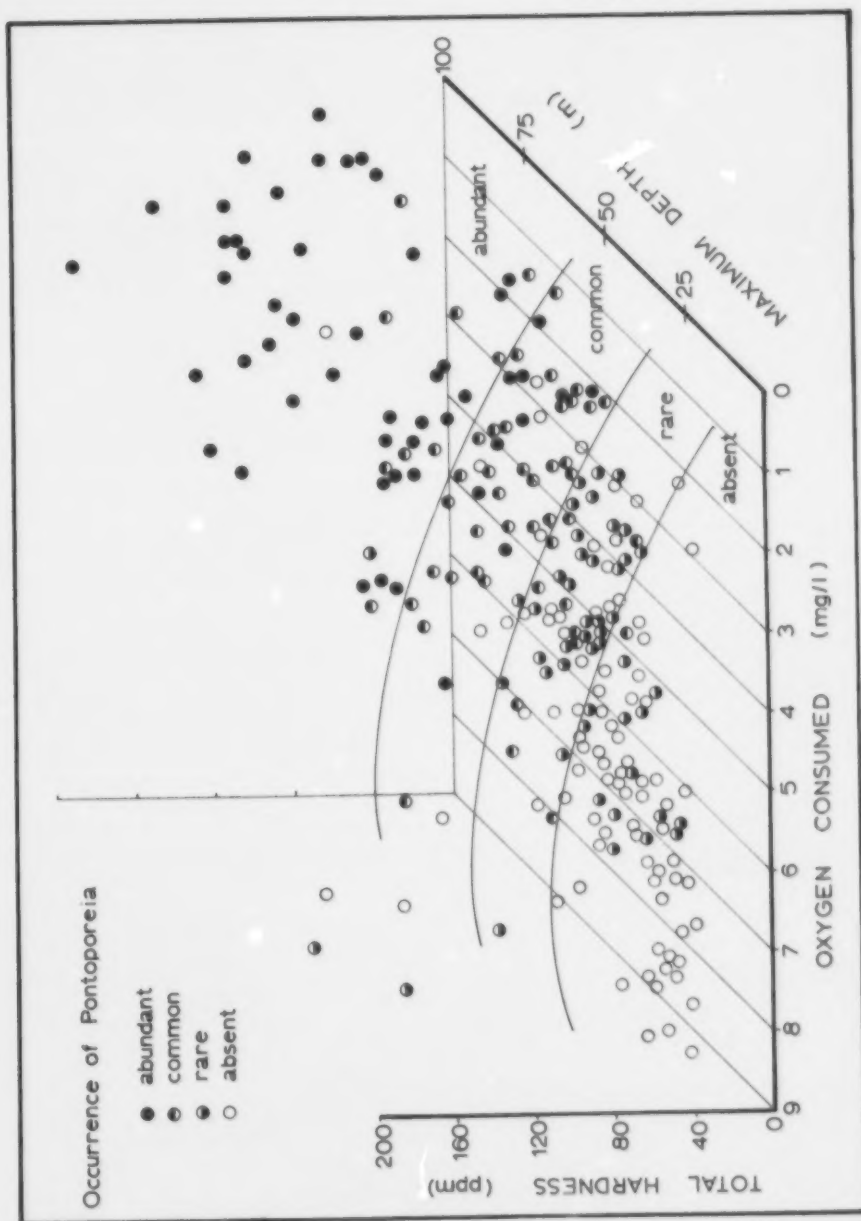


Figure 8
Relationship between maximum depth, total hardness, oxygen consumption, and occurrence of *Pontoporeia affinis* in the study lakes

Results of the Ekman sampling indicate that population size of *Pontoporeia* decreases with decreasing TH and increasing O_2C (Figure 9, Appendix 2). Abundance of *Pontoporeia* first increases and then decreases with depth (Figure 9), but this change does not seem to be precisely related to changes in pH (Figure 9, Appendix 2). Other factors, such as increase of CO_2 or decrease of food with depth, may influence the decline in the number of *Pontoporeia* at greater depths. Usually *Pontoporeia* was absent from lakes with hypolimnion pH below 5.5 or from deeper regions of hypolimnions, where pH is below 5.5 (for example, Lake Bernard, No. 122, Appendix 2). The absence of *Pontoporeia* below 10 m in Little Deer Lake, Saskatchewan (Koshinsky 1965), was probably due to the pH- O_2C situation in that lake. Koshinsky gave a colour of 26, which corresponds to about 6 ppm O_2C , and a hypolimnion pH of 6.5 for this lake. Both levels are within the critical range of these factors for the survival of *Pontoporeia* (Figure 8).

Obviously, low TH-pH and high levels of dissolved organic matter restrict the occurrence of *Pontoporeia* both between and within lakes. It is possible that this environmental problem is linked to the amphipod's osmoregulatory and calcium requirements. More research is needed to answer this question.

Factors Affecting *Limnocalanus* and *Senecella*

These two copepods are considered together, since it appears that interspecific competitive exclusion between the two is taking place in certain lakes, limiting their individual occurrences. Both species can survive the full range of physicochemical conditions found in the study lakes (Figure 6, Table 3), but they occur together in only 63 (33%) of the 191 localities containing one or the other.

Figure 10A demonstrates the relationship between surface pH, SDV, z_m , and the occurrence of the copepods. Hypervolumes of the four possible community combinations—that is, both occurring, *Limnocalanus* only, *Senecella* only, and neither occurring—are outlined in Figure 10B. Unlike the results of previous workers (Patalas 1971), lake surface area was not found to be a limiting factor in the distribution of these copepods. They were often found in very small lakes (for example, Lac Leamy, No. 251, 18 ha).

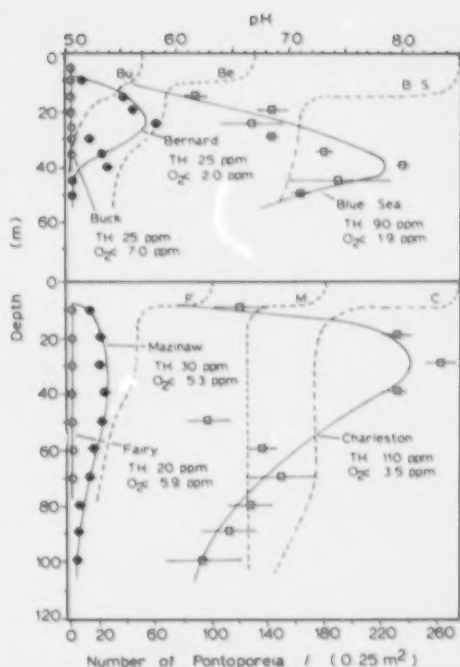


Figure 9
Abundance and depth distribution (solid lines and symbols) of *Pontoporeia affinis* in relation to water chemistry in six lakes. Broken lines represent pH profiles of the lakes.

Limnocalanus usually dominates in lakes with pH over 7.0, *Senecella* in lakes under 7.0. The species composition of other crustacean plankton communities is known to be affected by pH (Carter 1971).

The amount of light penetration, however, seems to be the dominant factor determining the coexistence of these two species in a lake, although their occurrence is not correlated to Secchi disk visibility (Table 4). Lakes that were either very deep or had low light penetration often contained both copepods. Perhaps, in these situations, light-tolerance levels of the copepods are not exceeded in most of the hypolimnion, allowing room for populations of the two species to segregate themselves vertically in the water column. Vertical separation of copepod species, usually to avoid interspecific competition for food and space, is a necessary condition in most planktonic communities (Rigler and

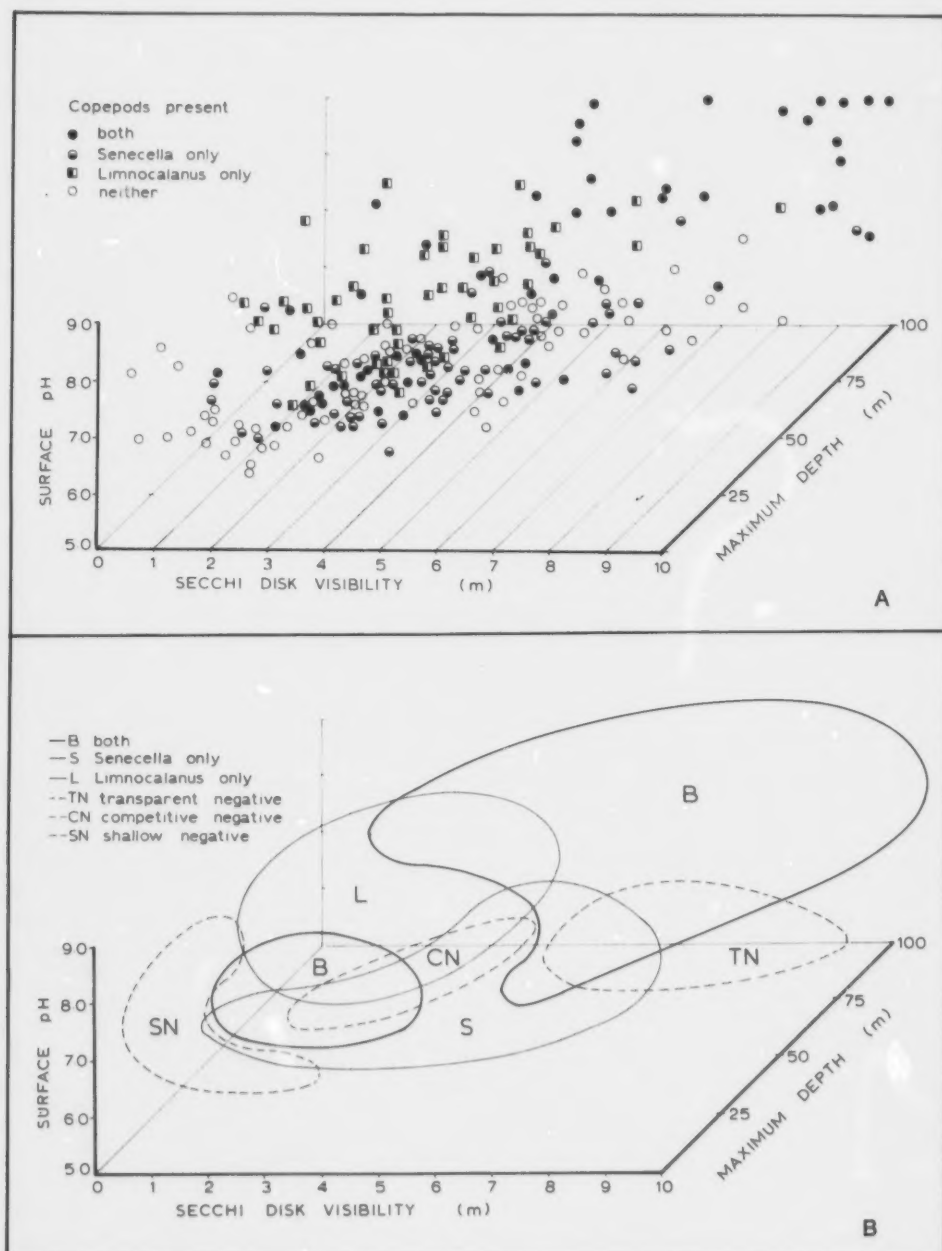


Figure 10

A. Occurrence of *Limnocalanus* and *Senecella* in the study lakes in relation to maximum depth, surface pH, and Secchi visibility.

B. Hypervolumes of the deepwater copepod communities.

Langford 1967). Carter (1969) found that *Limnocalanus* and *Senecella* vertically segregated themselves in Parry Sound.

Three lake types lack both of the copepods (Figure 10B). Very shallow lakes were probably unsuitable because of high water temperature and high levels of incipient light. In moderately deep lakes with extremely transparent water (SDV over 8 m), light-tolerance levels of the copepods may be exceeded even in the deepest water, or perhaps fish predation is increased, creating intense selective pressure against these large copepods. Brooks (1968) and Wells (1970) have documented cases in which planktivorous fish have eliminated the larger planktonic forms from certain lakes. Higgins (1966) found that cisco, *Coregonus artedii* (the dominant planktivorous fish in the study area), feed selectively on larger species of zooplankton. The "competitive negative" lakes have physico-chemical conditions that place them in the overlap region of the single-species community hypervolumes (Figure 10B). Possibly, if neither copepod has an adaptive advantage under these conditions, competition with each other may reduce both populations to extinction, or to such low numbers as to be virtually unobtainable by normal collecting.

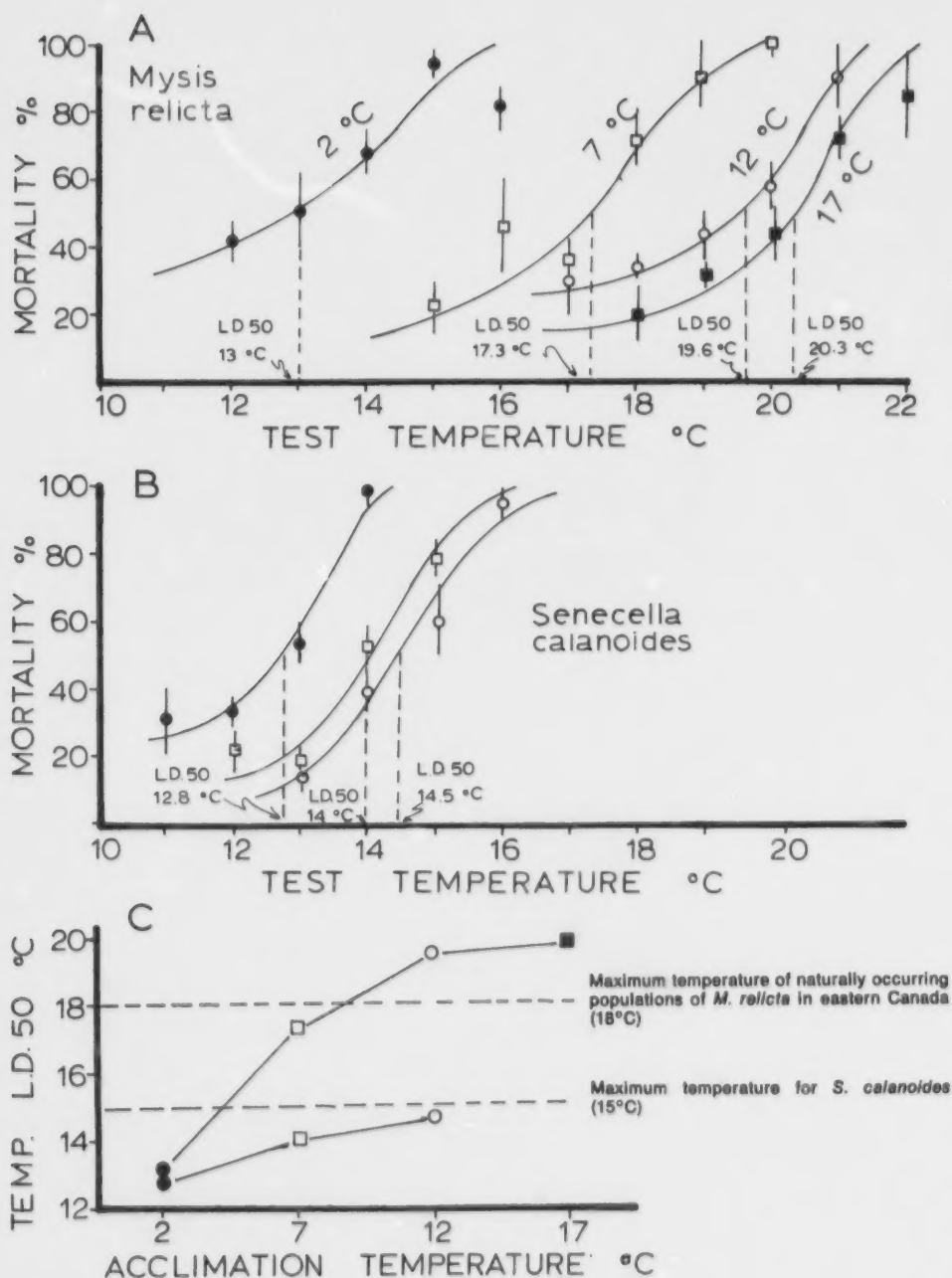


Figure 11
A,B. Mortality at various test temperatures after acclimation to 2, 7, 12, and (*Mysis* only) 17°C. C. Change of the thermal LD50 in relation to the level of thermal acclimation.

Introduction

Early in this study it was found that ecologically acceptable lakes east of Ottawa, which had been part of the Champlain Sea, contained few or none of the "glaciomarine relicts". Records from the Arctic Ocean and the Baltic Sea, however, indicate that most of these species are holeuryhaline, and that they are found in, or can disperse through, fairly saline water (Holmquist 1959, 1963, 1970; Zenkevitch 1963; Grainger 1965). What factors then prevented widespread dispersal of the community through the Champlain Sea?

Experiments on the thermal and saline tolerances of these species are numerous. Holmquist (1959), Belyaev (1949), Ricker (1959), and Smith (1970) report on *Mysis relicta*; Green (1965) and Smith (1972) on *Pontoporeia affinis*; Westin (1968) on *Myoxocephalus quadricornis*; and Lockwood and Groghan (1957) on *Saduria entomon*. Their results, however, are conflicting, and seem to have been influenced by the physiological adaptations of the test population or the length of acclimation and test periods — that is, if actually no more than one true species is involved in each case. *Senecella* has never been studied in the laboratory. Nor have previous workers (except Holmquist) considered the combined effects of temperature, salinity, and light, all of which probably acted together to limit dispersal in the Champlain Sea.

Mysis and *Senecella* were chosen to work with because they were obtainable in large numbers, were easy to keep, and seemed likely to have the greatest differences in tolerance to the experimental parameters. For example, *M. relicta* is known from water of 29‰; *Senecella* has not yet been found in salt water.

Methods

Specimens of *Mysis* and *Senecella* were collected from Charleston Lake (Map 3, No. 190). Collections were made in 80–90 m of water, with a large, conical plankton net (1-m mouth, 11 meshes per cm) hauled vertically by hand. The catch was stored in large plastic bags one-third filled with lake water, inflated with oxygen, and placed in cardboard boxes for transportation. Surface water temperatures during the December–March collecting pe-

riod ranged from 8° to 0°C. Collecting was done on cool, overcast days, and temperatures in the transportation chambers were kept below 6°C. Few animals died during collection and transportation.

Mysis and *Senecella* were kept separately in 12-liter plastic tanks, 200–300 animals per tank, in a dark constant-temperature room. Laboratory well water and lake water were mixed 50:50 during the first 24 hours of captivity to reduce osmotic shock (Fürst 1965), and were then replaced by well water. Water in the holding tanks was changed weekly, and dead animals were removed daily. The water in the tanks was not aerated, as this has been shown to be detrimental to survival (Smith 1972), but because of its small volume in relation to surface area, the water maintained adequate oxygen (8 ppm). Lake-bottom mud was placed in the tanks to provide food, and was supplemented with a daily feeding of a commercial dried egg-yolk preparation.

Experimental duration was 10,000 minutes (approximately 7 days), and animals were fed during the test period. Salt tests were done in 6-liter battery jars, thermal tests in 3-liter beakers. Thermal acclimation was at 2, 7, 12, and 17°C for *Mysis*, and 2, 7, and 12°C for *Senecella*. Salinity acclimation at each temperature was at 0, 5, 10, 15, and 20‰ for *Mysis*, and 0, 5, and 10‰ for *Senecella*. Acclimation was gradual — that is, temperature was raised to the required level by 0.5°C per day and salinity by 5‰ increments per week. Since earlier investigators (Dormaar 1970; Sturgeon 1970) have found that these animals acclimate slowly, they were kept at the acclimation level for at least two weeks before tests were conducted.

Test temperatures were maintained using a constant-temperature chamber. Solutions for salinity tests were made by diluting a 32‰ stock solution made from "Instant Ocean Sea Salts". Salinity levels were determined by the silver nitrate–potassium chromate low-precision titration method (Strickland and Parsons 1970), standardized at 7°C. Saline solutions were made up to 6,000 ml, and evaporation losses during the test period were replaced by the addition of distilled water. Halfway through each test period, salinity was redetermined and adjusted (if necessary) to within 0.1‰ of the desired level. All tests were done in total darkness.

Depending on the availability of animals, up to 10 but never fewer than 6 mysids, and

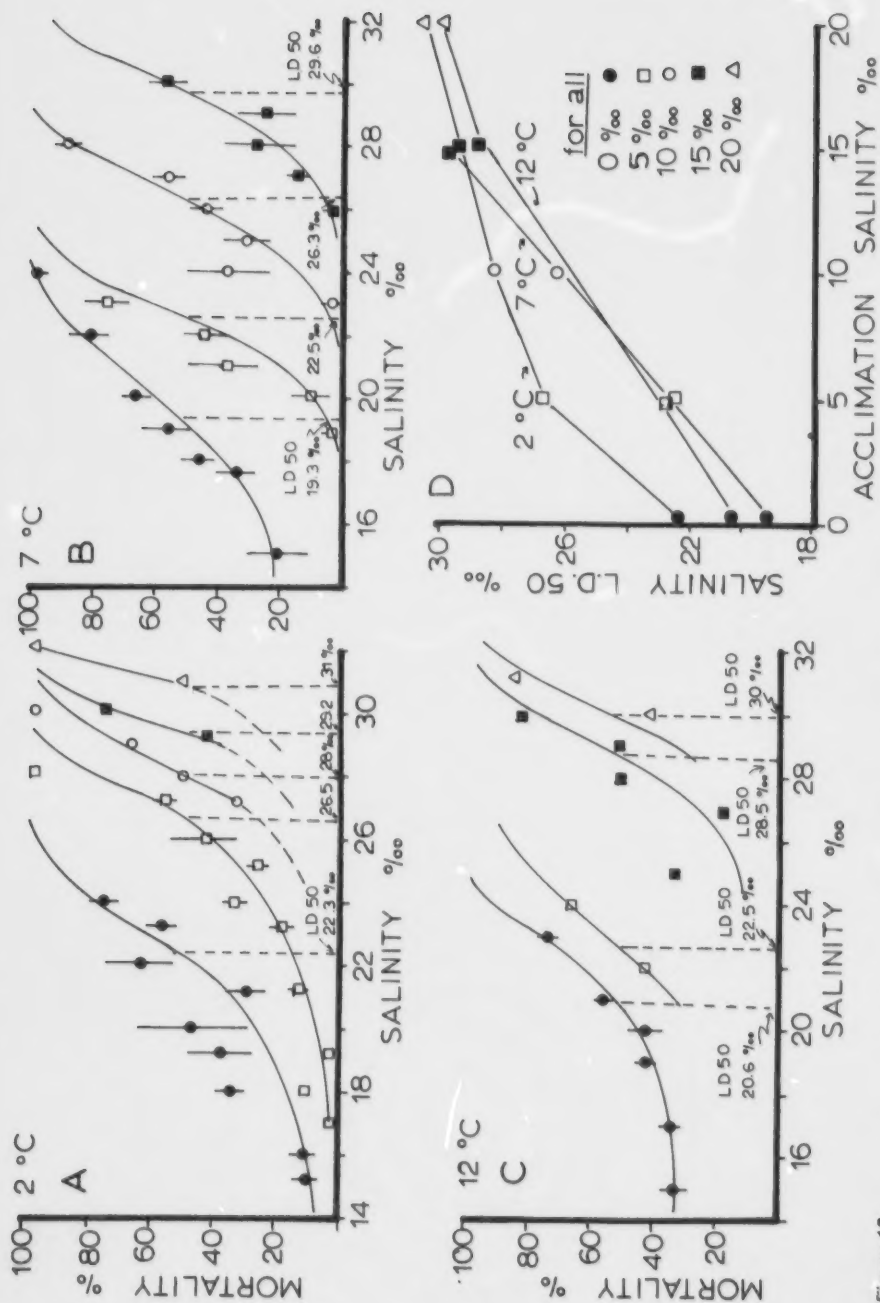


Figure 12
A, B, C. Mortality of *Mysis relicta* at various test salinities after acclimation at 2, 7, 12 °C and 0, 5, 10, 15, and 20 ‰ salt.
D. Effect of acclimation salinity on the salinity LD50 at various temperatures.

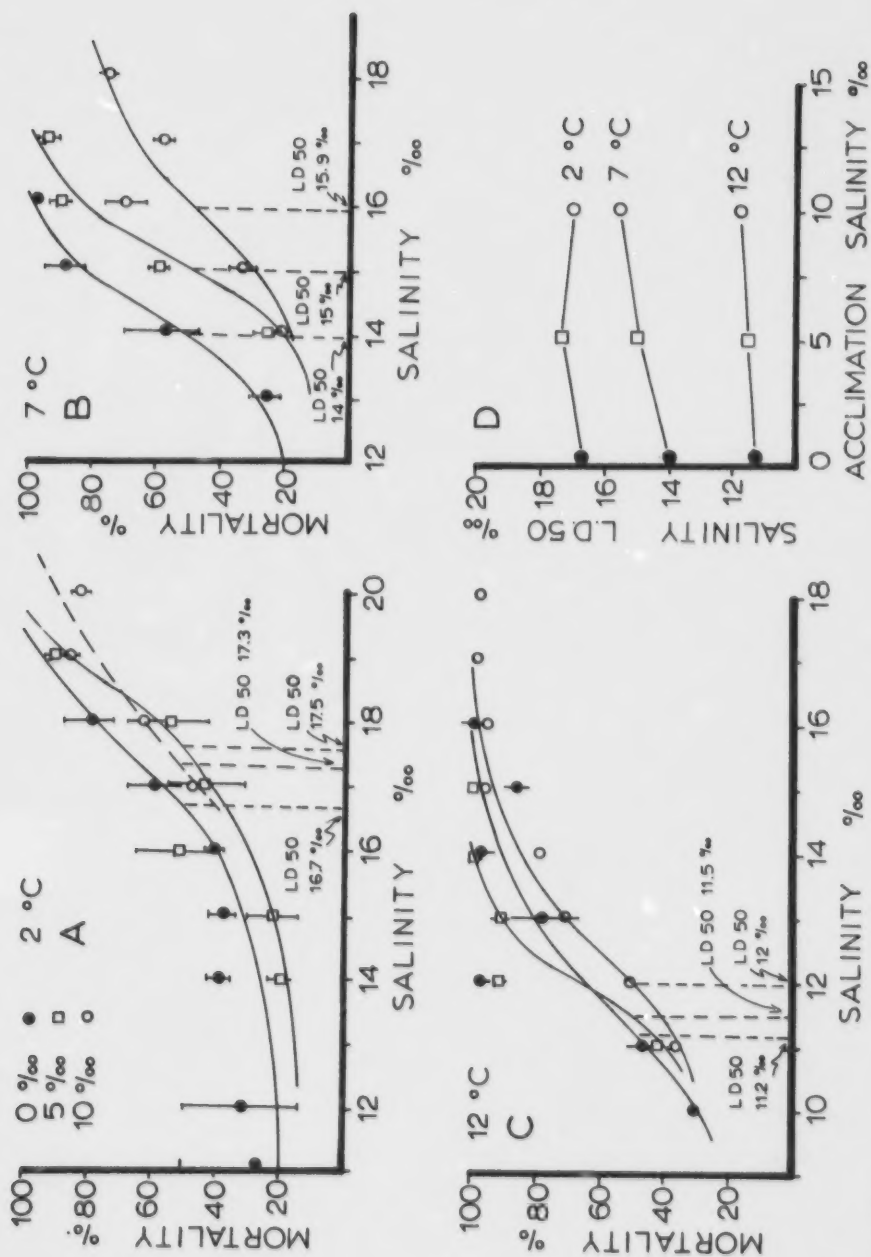


Figure 13
A,B,C. Mortality of *Sonocella calanoides* at various test salinities after acclimation at 2, 7, 12°C and 0, 5, 10‰ salt.
D. Effect of acclimation salinity on the salinity LD50 at various temperatures.

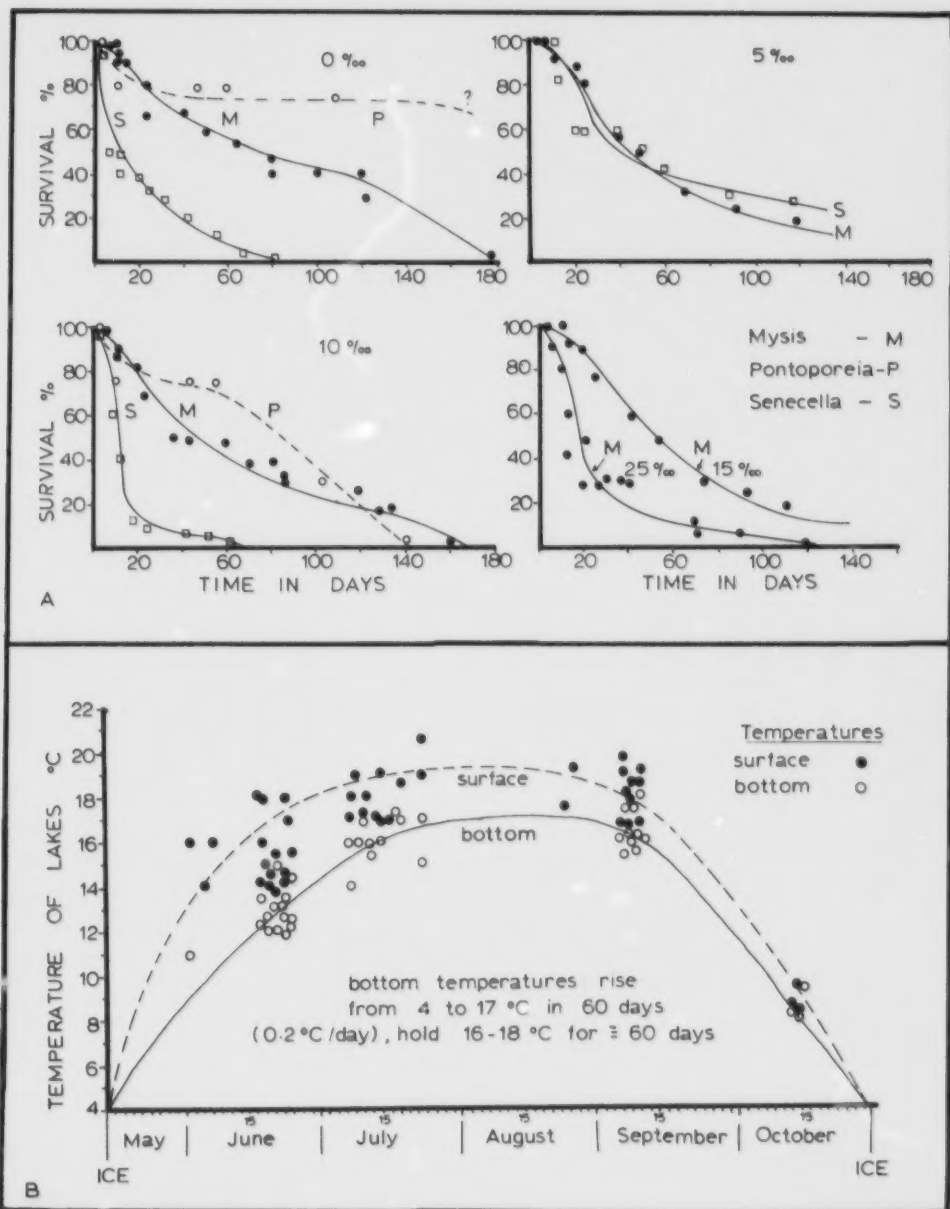


Figure 15
A. Long-term survival of *Mysis*, *Pontoporeia*, and *Senecella* at various salinity levels.

B. Thermograph of the shallow lakes (III Order) in the Abitibi region that contain populations of *Mysis relicta*. Lakes were sampled over a period of three summers.

up to 50 but never fewer than 10 *Senecella*, were used at each test level. Testing was done by plunging animals directly into the test level from the acclimation level. Test tanks were observed twice daily to record mortality. *Mysis* was determined as dead when probing with a fine needle produced no response, *Senecella* when the animal became opaque, which is about 2 hours after death. Percent mortality was recorded after 10,000 minutes. All tests were duplicated.

Controls containing 10 animals at the original acclimation level were run concurrently with tests. Invariably, mortality was less than 10%, and was not considered when calculating the LD50s.

Long-term survival experiments on *Mysis*, *Pontoporeia*, and *Senecella* in 0, 5, 10, 15, and 25‰ were run for a period of six months. These were kept at 7°C, in 6,000-ml tanks provided with bottom mud and food. Tanks were examined at weekly intervals and mortality was recorded.

Results and Discussion

Upper lethal temperatures of *Mysis* and *Senecella* were 20.3°C and 14.5°C respectively. Both values agree more closely with the observed upper thermal limit of these species in nature (Figure 11) than values obtained by previous workers. Ricker (1959, quoting Larkin's unpublished data) gave the upper thermal limit of *M. relicta* from Waterton Lake, Alberta, as 22°C. Smith (1970) disagreed, and obtained a value of only 16.5°C for animals from Lake Superior. In the first case, test times were apparently too short; in the second, acclimation periods (only one week) were not long enough.

Mysis was able to acclimate and survive over a wide thermal range (2–17°C), and its thermal tolerance rose significantly with acclimation to higher temperatures (Figure 11). *Senecella* had only a narrow thermal range. A 10-degree increase in acclimation temperature only raised its upper lethal limit by 2°C (Figure 11).

Salinity experiments yielded much the same results. *Mysis* survived up to almost full seawater concentrations (30‰), and its salinity LD50 rose with an elevation of acclimation level (Figure 12). Holmquist (1959) gave a range of 3–11‰ for occurrence of *M. relicta* in the Baltic Sea, and reported on a population living at 29‰ in an Alaskan la-

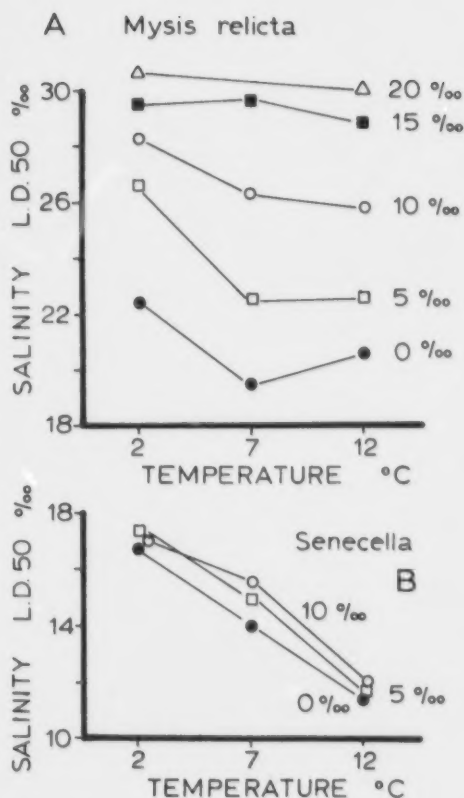


Figure 14
Salinity LD50 of *Mysis* and *Senecella* at 2, 7, and 12°C after acclimation to various salinities.

goon (Holmquist 1963). *Senecella*, on the other hand, showed virtually no acclimation to rising salinities, and had an upper lethal limit of only 17.5‰ (Figure 13). But the salinity tolerance of *Senecella* is still better than most oligohaline freshwater invertebrates, which usually have no tolerance above 10‰ (Kinne 1963), and possibly reflects a moderately recent marine ancestry.

The combined lethal effect of high temperature and high salinity is even more striking. Salinity tolerance of *Mysis* is reduced by high temperatures, particularly at low salinities (Figure 14A). At higher salinity acclimation levels this effect was negligible. With *Senecella*, rising temperatures lower the salinity resistance, but unlike results with *Mysis*, prior acclimation to higher salinities does not raise the tolerance level (Figure 14B).

Long-term experiments (6 months) disclosed that freshwater populations of *Mysis*, *Pontoporeia*, and *Senecella* are capable of extended survival in saline water, but that survival was best at low salinities (Figure 15A). Dormaar (1970) stated that *M. relicta* from Lake Ontario survived more than a month at 30‰. The major loss of animals during the long-term experiments was due to the natural death of adults; some immature mysids were kept as long as 16 months. The natural acclimation rate of *Mysis* to high temperatures in the shallow lakes of the Abitibi district is shown in Figure 15B. The average rate of temperature rise (0.2°C per day) is quite rapid. The mysids survive up to 2 months at 16–18°C in these lakes.

Smith (1970) found that a laboratory population of *M. relicta* underwent continuous die-off when subjected to normal laboratory lighting over extended periods. This confirmed similar reports by Larkin (1948) and Holmquist (1959). I was unable to study light-induced mortality of *Mysis*, but one observation was made. When an attempt was made to test the salinity resistance of freshwater 2°C-acclimated *Mysis* in a lighted room, 100% mortality occurred after only 24 hours at salinities as low as 18‰. The LD50 at 2°C when tested in the dark for a seven-day period was 22.3‰ (Figure 12). Merker (1940) observed that light increased the permeability of crustacean integuments, and in some cases caused osmotic failure. Perhaps this is what occurs in *M. relicta*.

One source of error that should be mentioned is the different survival rates of immature and mature *Mysis* under environmental stress. A shortage of animals made it impossible to consider this factor in my experiments and has introduced some error in the results. General observations in the field and during experiments indicate that immature mysids are more tolerant than adults to high temperatures but less tolerant to high salinities.

In conclusion, the experiments indicate that *Mysis relicta* is a eurythermic, holeuryhaline animal, but that *Senecella calanoides* is only slightly eurythermic and is oligohaline.

Origin and Early Dispersal

During the last 100 years, zoogeographers have developed a number of hypotheses to explain the time and place of origin, and subsequent Holarctic dispersal, of the "glaciomarine relicts". Three major views are held: a widespread, multiple, independent origin in fresh water after isolation from the sea; a single freshwater origin from salt water and a later dispersal in both fresh and salt water; and a single marine origin followed by a spread to fresh water. Whichever view is true, as Ricker (1959: 876) states, "The problem of the origin of North American relicts is inseparable from that of the origin of the corresponding relicts in Eurasia."

The multiple-origin hypothesis holds that "relict" species evolved independently in each lake from marine ancestors after separation of the lake from a postglacial marine inundation. This was the original argument put forward by Lovén in 1861 to explain the morphological differences between the marine and freshwater "species". Since then, in some of the species, forms identical to the "freshwater" species have been found in marine situations (Holmquist 1959, 1970), and it seems that they, and not their marine relatives, gave rise to the freshwater populations. Moreover, in view of the genetic drift usual in small founding populations, it seems unlikely that independent evolution could lead to such phenotypically similar populations of certain species in Eurasia and North America (Holmquist 1959; Lindquist 1961).

Segerstråle (1962) supports the view of a recent single origin in fresh water from salt-water forms. He proposed that the "glaciomarine relicts" evolved their form and freshwater tolerance in a large Siberian ice-dammed lake that resulted from the trapping of a shallow sea in that area by glaciers during the Riss glaciation, and that they then dispersed east and west mainly by freshwater routes. Segerstråle (1966) later admits that there is no good geological evidence in support of the existence of this Siberian ice-dammed lake. Holmquist (1959) holds a similar view, but believes the time of origin was earlier, perhaps during the Miocene, when the seas that had covered central Eurasia regressed, leaving populations of their ancestors isolated in lakes, and these evolved into the freshwater species.

Lomakina (1952) and a number of other Russian authors (such as Gurjanova and Pirozhnikov, cited in Ricker 1959) believe the

"glaciomarine relicts" evolved from marine ancestors in brackish portions of the Arctic Ocean during the Pleistocene, and then spread to fresh water. Most new evidence supports this latter view. Some of the "freshwater" species are known to form a major portion of the fauna in arctic, brackish situations (Zenkevitch 1963; Holmquist 1970). Holmquist (1963) found two closely related species of *Mysis*, the "marine" *litoralis* and the "freshwater" *relicta*, living in an Alaskan lagoon without introgression. Other genera (*Gammaracanthus*, *Myoxocephalus*, *Limnocalanus*) apparently form morphological clines along salinity gradients, with what is essentially the "freshwater" form occupying very brackish water, and the "marine" form the open sea. Dormaar (1970) found that the "freshwater" *M. relicta* still osmoregulates as an estuarine animal, and Lockwood and Groghan (1957) showed that chloride regulation in *Saduria entomon* living in fresh water was characteristic of a marine organism. All these facts suggest that the "glaciomarine relicts" evolved recently from marine ancestors, or that they are true marine animals capable of survival in fresh water.

Senecella calanoides may have a different history, since it seems to be primarily a freshwater organism. It is, however, the only paracalanid in fresh water (Marsh 1933), and all of its relatives are marine. It may have entered the fresh waters of North America earlier than the other species.

It seems plausible to me that the evolution of the "relict" species now occurring in fresh water originally took place in brackish portions of the Arctic Ocean during the late Pliocene-early Pleistocene. During Pleistocene glacial periods, sea levels were lowered by approximately 100 m (Flint 1957), and the Arctic Ocean was cut off from saltwater influx except through the Greenland Strait (Map 11). This decrease in area and volume, coupled with the probable tremendous inflow of fresh water from the Mackenzie and the other large rivers, probably created a very brackish Arctic Ocean (Bowman and Long 1968). A stable salinity gradient could have existed in the western part of the ocean for long periods during each glacial maximum, and habitat selection according to salinity could have resulted in speciation (Smith 1966) and in the ultimate development of pre-adapted freshwater animals. I believe that a slow, recurring situation like this would have

been necessary to develop freshwater tolerance in these animals. Stable salinity gradients of the type envisaged occur in the Baltic and Kara seas today, and, depending on salinity, one or the other of a number of marine-brackish sister species or subspecies dominates (Lomakina 1952; Segerstråle 1957; Zenkevitch 1963; Van der Land 1970).

Habitat stability usually results in species diversity (Valentine 1971). Organisms adapted to living between 0 and 4°C should respond genetically to stability at this temperature as organisms adapted to, and evolving in, tropical situations do. For example, the stability of the hypolimnion of Lake Baikal, USSR, over the last 60 million years has resulted in the evolution of over 30 endemic genera of amphipods inhabiting this part of the lake (Hutchinson 1967). It seems reasonable that at least one species pair could have evolved in a cold, moderately stable environment in the last 3 to 4 million years.

Although post- and inter-glacial marine inundations have undoubtedly dispersed these animals slightly inland around the Arctic Ocean (Holmquist 1963; Johnson 1964), dispersal farther inland to localities not inundated by marine waters during the Quaternary requires another mechanism. Marsh (1893) suggested avian transportation, but this seems unlikely for reasons discussed earlier (see pp. 3-5). Gurney (1923) proposed upstream migration, but this is improbable in view of the planktonic nature of some of the community. Högbom (1917) proposed an alternate means of dispersal, by proglacial* lakes, a hypothesis that has gained wide support from subsequent authors (Ricker 1959; Segerstråle 1962).

Högbom's "sluicing-up" theory proposed that the "relicts" existed in brackish seas in front of the advancing ice and were trapped and carried inland in the ice-frontal proglacial waters formed by isostatic depression (Walcott 1970). Then, from refugia situated at the maximum extents of glaciation, the animals redispersed through the glacial lakes formed during ice retreat. Maps 12 and 13 show that this kind of distribution pattern occurs in both Europe and North America.

I agree with Ricker (1959), in that the likely source of "glaciomarine relicts" for lakes in eastern North America is Hudson and James

bays. Two of the species, *Myoxocephalus quadricornis* and *Gammaracanthus loricatus*, are known to occur there (McAllister 1964; Johnson 1964). Early introduction of the organisms into fresh water around Hudson Bay could have occurred during previous post-glacial marine maxima, and with the advance of Labrador ice from the east (Flint 1957; Prest 1970) they were "sluiced-up", either from salt or fresh water, and carried inland to the south and west in ice-dammed proglacial lakes.

Senecella may have been in the fresh waters of North America before the Pleistocene. Perhaps it was left behind in arctic, coastal ponds after earlier marine regressions. Species of the genus *Eurytemora*, which is essentially a marine copepod, seem to be entering fresh water in this manner at the present time (Holmquist 1970). With the formation of the proglacial waters, *Senecella* could have joined the rest of the "relicts" and been dispersed inland.

Ricker (1959) thought it necessary for there to have been only one ice centre (that is, in northern Quebec) for the present distribution pattern of the community to have been formed, and he reasoned that ice at a Keewatin centre would have blocked dispersal. Even with two centres, however, dispersal inland would have been possible if the ice advance had followed a course similar to the known ice retreat (Prest 1969). Keewatin ice, advancing from the north and west, and Labrador ice from the north and east, could have coalesced over central Hudson Bay and trapped a proglacial water-body in the southwestern region of the bay. It is also unnecessary to postulate, as Ricker (1959) did, that the animals must have occurred in the St. Lawrence estuary in order for them to have reached eastern lakes. The glacial-lake systems and their connections, formed during the advance of the last ice sheet, are believed to be much the same as those formed during ice retreat (Mörner 1971). This means that even if the community were carried inland at only one point, perhaps in southern Hudson Bay, once it had gained access to the glacial-lake systems it could have easily dispersed to the eastern and western limits of the glacial lakes during ice advance and have been in position for transport to refugia around the southern ice margin during maximum glaciation.

*A "proglacial", or glacial, lake is a lake that has one shore consisting of glacial ice.

This argument does not necessarily imply that these animals invaded North American fresh waters only as recently as the last ice advance. There were three earlier major glaciations and other lesser ones (Prest 1970). Consequently, numerous opportunities have occurred for the "glaciomarine relicts" to disperse inland, and the mechanism of inland transport (that is, proglacial waters) was probably the same, no matter during which glaciation these animals gained access to central North America. However, evidence of these earlier possibilities has been masked by the last glaciation, of which the present distribution pattern of these organisms in eastern North America is the result. For that matter, the entire Holarctic distribution pattern of this group can be explained in terms of the last glaciation alone.

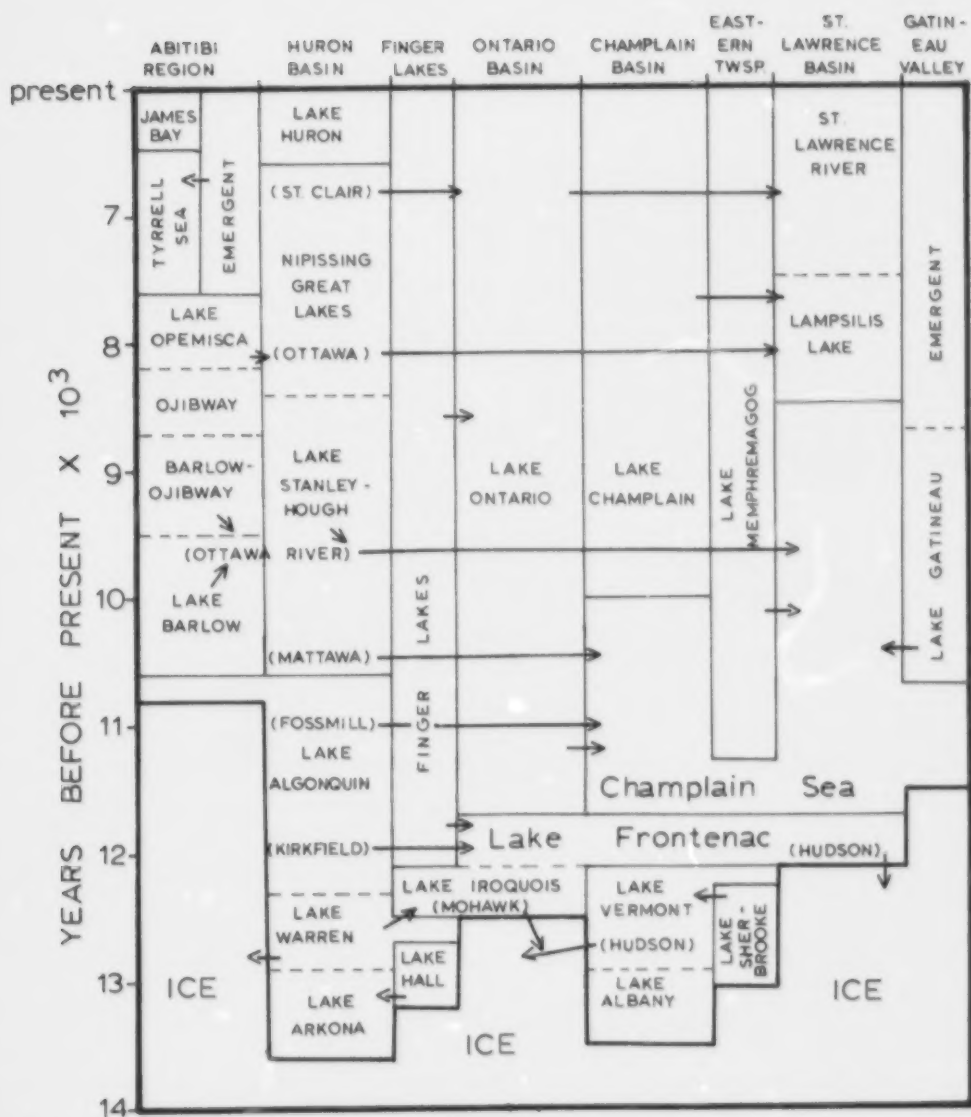


Figure 16
Sequence and time relationships between the glacial lakes, their spillways, and marine inundations formed during the retreat of the last ice sheet in eastern North America. Arrows indicate the direction of flow and the destination of the spillways. Correlation is based on Prest (1970).

The distribution of the deepwater community and the accompanying deepwater fishes (Dadswell 1972) in eastern North America suggests that their major dispersal routes during deglaciation were the interconnected standing bodies of glacial waters and their outlets. The occurrence of these animals in lakes inundated only briefly by the maximum levels of glacial waters indicates that dispersal closely followed ice retreat.

Names and sequences of glacial waters in the following discussion are taken mostly from Prest (1970). Since North American topographic maps and most geological papers give elevations and distances in the English measurement system, I have used it throughout this section to avoid confusion.

Refugia

Current evidence suggests that during the last maximum glaciation in eastern North America most aquatic organisms survived in refugia south of the ice margin (Frey 1965; Ross 1965; McPhail and Lindsey 1970). Most authors allude to only two refugia, generally in the Mississippi and Atlantic regions (Radforth 1944; Nelson 1968a; Khan and Qadri 1971). There are good indications that the deepwater community survived in a Mississippian refugium, since it is known to occur in Green and Geneva lakes, Wisconsin (Marsh 1893). Both these lakes are situated next to the driftless area, where proglacial lakes are known to have existed during the Wisconsin maximum (Frye, Willman, and Black 1965) (Map 14A). Geological evidence, however, indicates that small proglacial lakes existed in eastern Ohio (Goldthwait et al. 1965), the upper Genesee River Valley (Salamanca reentrant) (MacClintock and Apfel 1944), and the lower Hudson River Valley (Connally 1972) (Map 14A). All of these areas probably served as refugia for the deepwater community.

The absence of the community from lakes in the marine-inundated areas of Maine, New Brunswick, and Newfoundland, as well as from the valleys of the Connecticut and Merrimack rivers, where glacial lakes existed during ice retreat (Brooks and Deevey 1966) (Maps 3, 14A), suggests that no refugia existed east or north of the Hudson Valley. In New England, the advancing ice would have overtopped the Appalachians, and the steep downslope gradient would have caused any

frontal proglacial lakes to drain. Another possibility is that an ice cap already existed on the Appalachians (Prest, personal communication), and with the meeting of the two ice sheets the proglacial lakes were destroyed. In many places, such as the Gulf of Maine, the ice terminus fronted on marine situations that were probably too saline (Kenney 1964) for survival of the community (Map 14A). Nor does it seem that these animals were able to disperse north from the Hudson Valley into the sea along the Atlantic Coast, possibly because of high salinity or temperature.

When ice retreat began, between 17,000 and 14,500 B.P., the community followed in the small proglacial lakes formed along the ice front. As the ice retreated into the Lake Erie basin, these small lakes coalesced to form glacial Lake Maumee (Goldthwait et al. 1965), and with continued ice retreat the community spread into the Lake Huron basin during the glacial Lake Arkona phase around 13,500 B.P. (Figure 16).

Finger Lakes Region

Two lines of reasoning substantiate the possibility that a glacial lake in the Genesee River Valley served as the source of the community for the Finger Lakes. First, the community must have been in small regional glacial lakes before the Finger Lakes became connected to glacial Lakes Warren or Iroquois (Fairchild 1932; Goldthwait et al. 1965) because during these later phases, water levels were too low for the community to have gained access to higher lakes, such as Hemlock (No. 326, 905 ft) or Skaneateles (No. 320, 867 ft). The community could not have been present in the earlier Finger Lakes, for example, Newbury phase (Fairchild 1932), or it would have gained access to Canadice Lake (No. 708, 1,012 ft), which was part of the glacial lake at that time. The community probably dispersed from the Genesee Valley glacial lakes into the rest of the Finger Lake basins when these two lake systems joined during the glacial Lake Hall phase about 13,000 B.P. (Map 14B) (Prest 1970).

Lake Champlain Region

Whether the community survived in a Hudson Valley refugium is uncertain. Lakes sampled around New York City (Map 3) yielded none of the species, but all the sampled lakes were

above 300 ft in elevation, and Connally (1972) stated that the maximum glacial-lake level in this region was only 200 ft. Nevertheless, the community must have been in glacial Lake Albany during the Quaker Springs phase, since it is present in lakes inundated only by this stage (for example, St. Catherine, No. 313, Map 14B). Connally (1972) stated that the Quaker Springs phase existed about 13,000 B.P., but Prest (1970) indicated that the Mohawk Outlet from the Finger Lakes was not operating until 12,800 B.P. This means that the community probably survived in a Hudson refugium (possibly glacial Lake Hackensack), unless there was some earlier eastward flow from the Finger Lakes region.

The distribution pattern of the community in the Eastern Townships of Quebec is best related to the outline of the Sherbrooke phase of glacial Lake Memphremagog, as given by McDonald (1968). Existing lakes in Vermont that were inundated by earlier, higher levels of glacial Lake Memphremagog lack the community (Map 15A inset, Map 3). Eligo Pond (Map 20, No. 683), through which the earlier phases of glacial Lake Memphremagog discharged (Hitchcock 1906), also lacks the community.

The community probably gained access to glacial Lake Memphremagog from glacial Lake Vermont, but McDonald (1968) did not think that these two lakes were connected until the late Fort Ann phase of glacial Lake Vermont, by which time the water-plane was too low to have introduced the community into the higher lakes in the region where they occur (for example, Lovering, No. 308, 803 ft). There is a good possibility, however, that glacial Lake Vermont penetrated up the Missisquoi River and joined with glacial Lake Memphremagog just prior to the Sherbrooke phase. If Chapman's (1937) isobases of the late Coleville phase of glacial Lake Vermont are extended eastward, the level of glacial Lake Vermont is higher than that of the Lake Nick Outlet (Map 15A). McDonald (1968) noted no evidence of standing water over Lake Nick, but the connection was probably short-lived, and any evidence may have been removed by the later flow of water through this channel (McDonald 1968). The presence of the community in Orford Lake (No. 307, 905 ft) at the Missisquoi headwaters and the existence of a large reentrant (calving embayment?) in the ice front when it was in the Missisquoi Valley (McDonald 1968) indicate

that a glacial lake probably existed in the valley.

If the isobases of McDonald's Sherbrooke phase are extended up the Saint-François River, the probability arises that glacial Lake Memphremagog penetrated at least to the level of Lac Aylmer (No. 304, 820 ft, Map 15A) and allowed the community to reach there. The presence of only *Limnocalanus* and *Cotus ricei* in Lac Saint-François (No. 303, 900 ft) indicates that only a shallow, transient connection existed with the source area.

Lake Huron-Lake Ontario Region

As the ice front retreated, glacial Lakes Algonquin and Iroquois expanded northward, carrying the community with them (Map 15A). The precise relationship of the present localities of the community to the former extent of glacial Lake Algonquin (Maps 15A, B) was established by Martin and Chapman (1965). The occurrence of the community around Watertown, New York, east of Lake Ontario, is restricted to lakes below 700 ft, the highest known level of glacial Lake Iroquois in that area (Coleman 1932).

The occurrence of the community in two of three lakes north of Peterborough, Ontario, presents a problem. Crystal Lake (No. 158, 933 ft) contains most of the community; Mississagua Lake (No. 159, 966 ft) contains only *Limnocalanus*; and Jack Lake (No. 494, only 903 ft) contains none of the species. For this distribution pattern to have formed, ice must have covered Jack Lake, while at the same time it blocked the Kirkfield Outlet and held glacial-lake waters at a high enough level to inundate the other two lakes. The community could have entered the Kirkfield Outlet from glacial Lake Algonquin when the outlet first opened, about 12,500 B.P. (Prest 1970), and then dispersed into the lakes during the Dummer retreat (Map 15B). It appears possible that the two lakes containing the community were ice-free while Jack Lake was still ice-covered, because the Dummer ice margin was sharply angled northwest to southeast (Chapman and Putnam 1966) (Map 15B). When the glacier retreated from the sill at Stony Lake (No. 160, 796 ft) (Johnston 1916a), water levels dropped, becoming too low to penetrate into Jack Lake.

Maximum levels of the northern shoreline of glacial Lake Iroquois have not been mapped toward its northeastern end. To over-

come this problem I constructed a hypothetical Iroquois water-plane, using the known northwestern Iroquois beaches formed at the time of the Kirkfield Outlet (Johnston 1916a; Coleman 1932), and extrapolated a rebound curve northward, parallel to the known warped water-plane of nearby glacial Lake Algonquin (Figure 2) (Johnston 1916a). When the isobases of this water-plane are established on a topographic map of the area, the hypothetical outline of late glacial Lake Iroquois appears as in Map 15B. The occurrences of the community are almost 100% within this hypothetical lake-boundary. Only Weslemkoon Lake (No. 161, 1,030 ft) is outside.

Henderson (1971) did not believe that glacial Lake Iroquois extended into the Mazinaw Lake area (No. 167), and he attributed the lacustrine deposits there to a small glacial lake held up by a sill at Marble Lake, just south of Mazinaw Lake. However, an examination of 1:50,000 topographic maps of the area revealed that it was impossible for this small lake to have existed at the level Henderson gave for the lacustrine deposits; a topographic low at Swamp Creek would have drained the lake below this level. The presence of an outwash boulder-train south of Mazinaw Lake was cited by Henderson as evidence that glacial Lake Iroquois regressed when the ice front still covered Mazinaw Lake and the lakes to the north and east. Prest (personal communication) has suggested that the outwash may have been laid down following a small ice readvance into the deep basin of Mazinaw Lake only, after Iroquois had regressed, allowing the community to survive in the adjoining lakes at elevations up to 900 ft. After this ice retreated, the community reinvaded Mazinaw from the nearby higher lakes. The occurrence of *Limnocalanus* in Weslemkoon Lake remains unexplained; perhaps it was introduced there unknowingly by man.

Events in the St. Lawrence Lowland

Around 12,000 B.P. a number of events took place in rapid succession. The ice retreated from Covey Hill, Quebec, and lowered the level of glacial Lake Iroquois, uniting it with glacial Lake Vermont to form glacial Lake Belleville-Fort Ann (Map 16A) (Prest 1970). This lake was maintained in the St. Lawrence Valley by ice blockage of the valley at a point

between Quebec City and Rivière-du-Loup. Ice-calving caused the ice front to retreat rapidly to the Ottawa River and the highlands immediately north of Montreal. The north shore of Lake Belleville-Fort Ann is unknown, but if Chapman's (1937) isobases for the Fort Ann phase of glacial Lake Vermont are extended northward with the same isobase separation distance as that of the Champlain Sea (Figure 3B), the 900-ft isobase occurs just north of Montreal (Map 16A). The presence of the community in four lakes north of Montreal, but only up to 900 ft in elevation, is probably attributable to this lake phase (for example, Lac Louisa, No. 295, 890 ft, Map 16B).

The occurrence of *Limnocalanus* in Lac Temiscouata (No. 302, Map 16A) is problematic and cannot be readily explained on the basis of known geological evidence. This again may be a chance occurrence (human transport?), or it may be possible that the copepod reached the area from the west through a series of ice-marginal lakes formed along the Highland Front morainic system. Gadd's statement (1964: 1253) that "the ice front impinged against the highland at elevations of 700 ft", together with his map of the moraine and the hypothetical isobase situation at this time, makes it possible that a long arm of glacial Lake Vermont may have penetrated to the Rivière-du-Loup area. Lee (1962, 1963) indicated that when the ice front was at Rivière-du-Loup there was glacial drainage south to the St. John River, and regional topography makes it possible for this drainage to have gone through Temiscouata. Radio-carbon dates, however, indicate that the sea had penetrated almost to Quebec City by 12,700 B.P. (Prest, personal communication). If this date is right, it would be impossible for the glacial-lake situation I have outlined in Map 16A to have existed at 12,000 B.P. The possibility that *Limnocalanus* reached Lac Temiscouata via the flowage from the Chaudière Valley to the upper St. John River (Gadd 1964) is ruled out by the copepod's absence from lakes in northern Maine and New Brunswick, all of which were part of a glacial lake that existed in the upper St. John Valley at the time of the connection (Prest, Grant, and Rampton 1968). Once *Limnocalanus* had gained access to Temiscouata, dispersal down the St. John River was simple. *Limnocalanus* is found in the mainstream drift of this river in spring, and there is a permanent

population in Belleisle Bay in the St. John estuary.

The Champlain Sea Episode

At approximately 11,900 B.P., when the ice retreated from the St. Lawrence Valley south of Quebec City, Lake Belleville-Fort Ann drained to sea level, and the ocean flooded the valley to form the Champlain Sea (Prest 1970). By this time the ice front stood north of the St. Lawrence River along the Saint-Faustin-Saint-Narcisse Moraine (Parry and MacPherson 1964; Beland 1953), and the northern part of the sea flooded the land up to the present elevation of 700 ft (Johnston 1916b; Gadd 1971; Romanelli 1972) (Map 16B).

Palaeontological studies have revealed that conditions during the initial stages of the Champlain Sea were high arctic, with summer water temperatures below 5°C and surface salinities above 25‰ (Elson 1969; Harington and Sergeant 1972). Because of the proximity of the ice front at this time (Map 16B), there was probably a thin surface-layer of brackish water along the northern shore. The occurrence of landlocked arctic char in certain Quebec lakes (Map 16B) — the remnants of former anadromous populations — is also indicative of arctic saline conditions. Present-day anadromous char populations occur only where summer sea-temperatures are below 10°C (Map 17).

Some of the "glaciomarine relicts" cannot survive highly saline conditions (see experiments, pp. 43–44; Segerstråle 1957), and, in essence, the early phases of the Champlain Sea acted as a barrier to their dispersal. Nevertheless, *Limnocalanus* and some of the fishes (Dadswell 1972) continued to disperse in the early phase of the Champlain Sea, probably by brackish surface-layers, and *Gammaracanthus* apparently entered the sea during its earliest phase, probably from the Atlantic Ocean. *Gammaracanthus* would have had no trouble dispersing at this time, as it is commonly found in salinities as high as 25‰ (Lomakina 1952; Drainville 1970). Heart Lake (No. 287), the only freshwater locality in which *Gammaracanthus* was found, also contains *Limnocalanus* and *Pungitius*, both of which are known to be quite salt-tolerant (Holmquist 1970; Nelson 1968b). The difficulty of collecting *Gammaracanthus* may be the reason for its apparent absence from

other marine-inundated lakes. Even in lakes where this species is well known, it is not abundant (Grimas 1969).

Limnocalanus probably dispersed widely in the early phase of the Champlain Sea, but populations survived only locally — in areas where maximum dilutions would be expected to have occurred (for example, river mouths) — and were isolated in lake basins in these areas by falling water-levels (for example, Lac Papineau, No. 293). Inundated lakes that had broad connections to the sea at this time (for example, Lac Simon, No. 602, Map 20) (Faessler 1948 a,b) are all negative for *Limnocalanus*. The salinity in the region of these basins during the period when they were separating from the sea was probably too high for its survival.

Limnocalanus and *Myoxocephalus* — also quite salt-tolerant (McAllister 1964) — are the only two species of the community known from lakes in the Lièvre Valley. These species and the fishes *Cottus ricei* and *Pungitius pungitius* probably penetrated into the valley before sea level dropped below the 600-ft sill at High Falls, Quebec (Map 18B). Once in the valley, they were able to continue dispersing in the glacial lake formed north of this sill (Map 18C).

During the early phases of the Champlain Sea, a brackish-water embayment apparently continued to exist in the upper Ottawa Valley (Goldring 1922; Wagner 1970), and the other crustaceans probably survived there (Map 18C, for example, the Golden Lake area). As the Champlain Sea shoaled, the water freshened (Elson 1969), allowing these species to begin dispersal, the initiation of which seems to have been controlled by more than just the salinity tolerance of each species. It seems that dispersal, when it occurred, was nearly simultaneous for all the species, since to reach the Gatineau Valley through standing water they would have had to disperse through the Champlain Sea, and although *Mysis relicta* is more salt-tolerant than either *Senecella* (see experiments, pp. 43–44) or the fish *Percopsis* (Dadswell 1972), their respective distribution patterns in the Gatineau Valley lakes are nearly identical.

Dispersal probably took place at a time when conditions were tolerable for *Senecella* (deep salinity less than 15‰). Observations indicate that *M. relicta* is unable to osmoregulate in sublethal salinities if exposed to light (see experiments, pp. 43–44).

It is possible that as the ice terminus retreated from the northern shore of the sea, areas of murky, brackish water were isolated in river mouths and were separated by areas of clear, saline water owing to the rapid precipitation of clay by ions in the seawater (Terasmae 1959). A clear-water area such as this may have existed along the Luskville escarpment and acted as a barrier between the upper Ottawa and the Gatineau valleys until deepwater salinities declined.

Pontoporeia "affinis" may have begun dispersal slightly before either *Mysis* or *Sonecella*, since it occurs in three lakes (Map 3: Thorne, No. 280; Johnston, No. 281; MacGregor, No. 282) near the Gatineau River at higher elevations than those of lakes inhabited by the other two species. Green (1965) obtained an LD50 of 21.9‰ at 7°C for *Pontoporeia* from Cayuga Lake (No. 322). At 7°C, the freshwater-acclimated *M. relicta* has an LD50 of only 19.3‰ (Figure 12B). Also, *P. "affinis"* in the Baltic Sea is more salt-tolerant than *M. relicta* (Segerstråle 1957).

Since the greatest influx of fresh water was probably from the north side of the Champlain Sea (where the glacier was situated), and since dispersal across the sea from southern lakes was probably prevented by high salinities, the probable dispersal route into the Gatineau Valley was across the northern reaches of the Champlain Sea from the upper Ottawa Valley. The hypothetical isobase situation for this period (Map 18B), the constricted neck of the Gatineau Valley, a probable low tidal-amplitude (Karrow 1961), and the presence of a 450-ft sill at Low, Quebec, blocking deep salt-wedge penetration into the valley (Gadd 1971), all point to the probability that a nearly freshwater standing connection existed between glacial Lake Gatineau and the Champlain Sea. The community probably reached the upper Ottawa River via the Fossmill Outlet (Martin and Chapman 1965), and dispersal of the less salt-tolerant species in the Champlain Sea began when water levels were about 600 ft (for example, Thorne Lake, No. 280, Map 16B). They seem to have reached the Gatineau Valley about 11,200 B.P., when sea level was about 500 ft present-day elevation (Map 18B).

Further evidence for this reasoning is found in the present distribution of smelt, *Osmerus mordax*, in lakes of the Ottawa and St. Lawrence valleys (Map 18C). Their pres-

ent marine distribution pattern suggests that anadromous populations of smelt require summer sea temperatures above 10°C (Map 17), and this fish was probably unable to invade the Champlain Sea until about 11,000 B.P., when conditions apparently approached those of the present-day Baltic Sea (Elson 1969; Wagner 1970). By 11,000 B.P. the sea level in the Ottawa area had dropped to the present-day elevation of 450 ft (Mott 1968) (Map 18C), and smelt, along with the community, were able to gain access only to lakes in the central Gatineau Valley. Lakes above 450 ft in the southern portion of the Gatineau Valley and in the upper Ottawa Valley contain the community but no smelt.

As isostatic readjustment continued, the Pagan Falls sill rose above the sea level, and separated glacial Lake Gatineau from the Champlain Sea (Map 19B). Glacial Lake Gatineau expanded northward as the ice retreated, reaching a possible present-day elevation of 900 ft at its upper end, and carried the community to the basins it inundated. Continuing isostatic rebound finally caused the glacial lake to drain, leaving the community in the existing lakes (Map 19B).

During its history, the Champlain Sea apparently alternated between a dispersal barrier and a dispersal route for various species, depending on their individual environmental tolerances. In its earlier phases especially, it exercised a strong influence over the dispersal of aquatic organisms in eastern North America. After 10,500 B.P., the sea level in the St. Lawrence Valley dropped below the elevation of any suitable existing lakes, and there is no evidence for further eastward dispersal of the less salt-tolerant members of the community.

Late Glacial Lake Algonquin and Glacial Lake Barlow-Ojibway Events

While the above events were taking place in the St. Lawrence Valley, glacial Lake Algonquin continued to expand northward, and about 11,900 B.P. the first of its northern outlets opened at South River, Ontario (Map 16B inset) (Harrison 1971).

Martin and Chapman (1965) attribute the majority of the community's occurrences in the northern portion of Algonquin Park to the Fossmill Outlet stage, but they explain its presence in a number of lakes above this outlet level (1,075 ft) by "sluicing-up" in a

small proglacial lake (to 1,250 ft) during a short readvance of the ice front. There is no evidence to indicate that a readvance occurred at this time (Harrison 1971), but there is a good possibility that a glacial lake existed in the Fossmill area, with maximum levels of 1,250 ft, during the operation of the South River Outlet. This glacial lake — early glacial Lake Amable du Fond (Harrison 1971) — could have been held up along the ice front by a sill, at 1,250 ft, just south of White Partridge Lake (No. 204, Map 16B inset). This outlet would have led down the Bonnechere River.

As the ice front retreated, lower outlets opened, dropping the level of glacial Lake Algonquin (Harrison 1971). By the time the present northern shore of Lake Huron became ice-free, water levels stood at a present-day elevation of 1,100 ft (Boissoneau 1968), and when the ice front was at the Whiskey Lake Moraine (Boissoneau 1968), the community could have dispersed into lakes in the Sudbury region (Map 3: Windy Lake, No. 105, 1,093 ft). With further ice retreat, even lower outlets were opened, and a large calving embayment opened in the ice front north and west of the Fossmill Outlet (Harrison 1971). When the ice front stood at the Cartier-McConnell Moraine (Map 18A), water levels were only 900–1,000 ft north and west of North Bay (Boissoneau 1968). But this was sufficient to allow the community access to the Temagami region (Map 18A).

As the ice front continued to retreat in the Temagami area, strong ice flow from Quebec blocked the Ottawa Valley at Deux Rivières (Harrison 1971), and a standing body of water could have extended up the Ottawa River (Harrison, personal communication) (Map 18C inset). If hypothetical isobases are drawn for this water body, two facts are revealed: it would have inundated the lakes in Quebec next to the Ottawa River south of Lake Temiskaming that contain the community (Map 18C inset: Lac Beauchêne, No. 96, 840 ft); and a standing-water link would have existed between the last phase of glacial Lake Algonquin and the first phase of glacial Lake Barlow, allowing the community to disperse into the Barlow basin.

When the ice withdrew from Deux Rivières, glacial Lake Algonquin drained, and glacial Lake Barlow became separate (Map 19A). Glacial Lake Barlow extended northward from a sill of morainic material that

plugged the Temiskaming trench (Boissoneau 1966), and although its shoreline has not been accurately mapped, it can be approximated. If the elevation of the morainic sill in the Temiskaming trench is assumed to have been at 850 ft (Boissoneau 1968 assumes 800 ft), and the standard isobase separation distances are plotted (from Figure 3B) northward to the ice front as depicted by Prest (1969) for 10,500 B.P., the resulting water-plane is as in Map 19A. The fit of the known community occurrences to this hypothetical water-plane is excellent, especially on the Quebec side of the Ottawa River. The argument is further strengthened by Hughes's (1959) observations of terraces (normally below a maximum lake level) at 985 ft along the Montreal River (at the hypothetical 1,000-ft isobase) and at 1,025 ft around New Liskeard (near the hypothetical 1,100-ft isobase). Vincent (1971) found the upper limit of lacustrine erosion at Ville-Marie, Quebec, to be 1,000 ft (just north of my 1,000-ft isobase).

Between 10,500 and 7,500 B.P., glacial Lake Barlow-Ojibway spread northward and segmented into a series of proglacial lakes (Norman 1938; Prest 1970). The community dispersed northward via this lake series. Its present distribution pattern agrees well with the overall maximum glacial-lake coverage given by Prest, Grant, and Rampton (1968). As a result of isostatic depression of the region during the existence of the glacial lakes, the occurrences of the community undergo a consistent northeastward rise in elevation, reaching lakes as high as 1,350 ft near Chibougamau, Quebec (Map 19C).

When the glacier receded from James Bay, these glacial lakes drained, except for a small water body around Lac Mistassini, No. 4 (Prest 1970). This small glacial lake spread northward, carrying the community until it reached elevations of 1,350 ft north of Lac Albanel, No. 3 (Ignatius 1953), and further expansion was halted by the rapid rise of the Otish Highlands (Map 19C: Lac Bethoulat, No. 2, 1,330 ft). This ended the known eastward glacial-lake dispersal of the community in North America.

Further dispersal to the south, however, apparently occurred after this. It seems that the upper level of glacial Lake Ojibway barely penetrated into two lakes around 1,300 ft in elevation south of Chibougamau (Map 3: Malo, No. 15; Rohault, No. 16), and *Limnoca-*

lanus and some of the fishes (Dadswell 1972) gained access to them. At first these lakes probably drained northward to James Bay, but isostatic readjustment has diverted their drainage southward down the Chamouchane River. *Limnocalanus* and the fishes have spread down this river into Lac Saint-Jean (No. 300, Map 19C).

The alternate dispersal route for *Limnocalanus* into Lac Saint-Jean (that is, down the St. Lawrence and up the Saguenay rivers) seems unlikely. Not only are water currents against this route (Drainville 1970), but lakes around Lac Saint-Jean that were also inundated by the sea do not contain *Limnocalanus* (Map 19C). The presence of *Percopsis* in Lac Saint-Jean indicates a freshwater dispersal route was in operation.

The presence of the community in lakes along the parallel Macho and Megiscane river systems also seems attributable to postglacial rebound (Map 19C). The drainage divide in this region is virtually nonexistent, and the Bell and Waswanipi river headwaters are strongly interdigitated. It is quite probable that lakes at the present headwaters of these rivers were once within the glacial-lake boundaries and drained northward, but isostatic rebound has diverted their drainage southward, or conceivably some headwater capture has taken place.

Gammaracanthus loricatus occurs in the Saguenay estuary (Drainville 1970). As mentioned earlier, this species, unlike the other members of the deepwater community, probably reached eastern North America by dispersing from the Arctic along the east coast of Labrador. At present, it is known from two brackish-water localities along the Labrador coast (Dunbar 1954; Carter 1966) and the St. Lawrence estuary (Bousfield and Laubitz 1972). The absence of this species from the Great Lakes and from lake basins I studied that were inundated only by glacial lakes seems to be more than just an artifact of collecting. The numerous trawls that I carried out, as well as the many fish-food studies (Dryer, Erkkila, and Tetzloff 1965) and extensive bottom sampling with otter trawls and dredges in the Great Lakes (Teter 1960; Dryer 1966; Henson 1966; Brownell 1970), have failed to find *Gammaracanthus*. I think that it was not present in the refugia south of the ice margin during maximum glaciation, and that it did not disperse in the main glacial-lake systems.

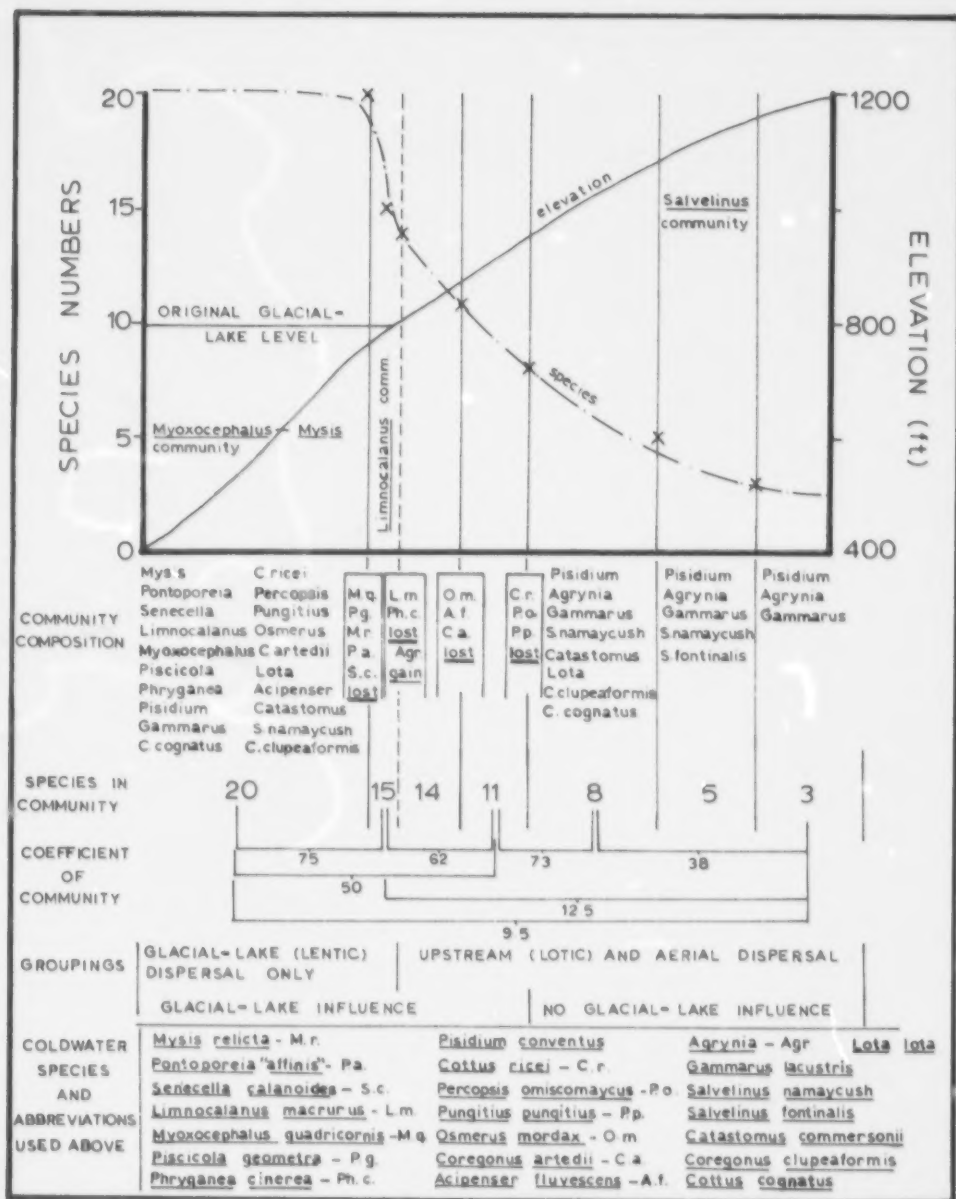


Figure 17
Change in the species composition of the deep-water community in relation to the elevation of the lake basin and the former glacial-lake level across a hypothetical transect of lakes in the Gatineau

Valley, Quebec. Note the sharp drop in the number of species at the uppermost former glacial-lake level.

Conclusions

Zoogeographers, for justifiable reasons, usually consider the Pleistocene glaciations as a period of destruction that disrupted habitats and caused many extinctions. During the glacial periods, however, unprecedented dispersal opportunities were opened for aquatic organisms, and an abundance of lakes not present before the Pleistocene were left behind. In fact, the "glaciomarine relicts" may have evolved their freshwater tolerance in response to conditions present during the Pleistocene, and they appear to have flourished during glacial periods and to be dependent on glaciations for inland dispersal.

In eastern North America most of the community's dispersal took place via the glacial-lake systems during ice retreat. The importance of glacial lakes for dispersal of deepwater or cold-adapted organisms is evident in Figure 17. Elevation of a lake basin in relation to former glacial-lake occurrence is the major factor determining the makeup of the lake's deepwater community. There is a sharp decline in the number of deepwater species as one crosses the former glacial-lake shoreline.

Even more striking is the strong similarity between the deepwater communities of lakes in widely separated geographical locations that have been inundated by the main glacial lakes. Although they are separated by 1,800 miles, the deepwater communities of Great Slave Lake (Rawson 1951, 1953, 1956) and Thirty-one Mile Lake (Map 3, No. 267) have a community coefficient of similarity of 80%. Sam Lake (Map 20, No. 573) is only 12 miles from Thirty-one Mile Lake and 340 ft higher in elevation, but the deepwater community coefficient of similarity is only 14%. Sam Lake was never inundated by glacial waters.

The "glaciomarine relicts" do not form a homogeneous group. *Mysis relicta*, *Pontoporeia "affinis"*, *Limnocalanus macrurus*, and *Myoxocephalus quadricornis* have much the same distribution patterns in brackish and fresh water throughout the Holarctic (Map 2). *Senecella calanoides*, however, is restricted to the fresh waters of North America and may have been in fresh water for a longer period than the other species. *Gammaracanthus loricatus* is primarily a marine form, with a short history of freshwater adaptation, and is restricted to lakes in areas of former marine submergence. *Limnocalanus macrurus* has dispersed farther east in fresh water than any of the other species in North America, and

may be able to disperse by means other than standing-water connections. In short, they are a group of animals held together mostly by their lack of highly evolved dispersal mechanisms, whose distribution pattern is a result of past geological events and present ecological conditions.

I agree with Holmquist (1966) that these species are not relicts. When their distributions are completely established, they will probably be known from lakes and brackish water in an unbroken chain westward from Quebec through the Holarctic region to Ireland. This type of distribution pattern is completely unsuitable to Darlington's (1957) definition of a relict. Also, rather than having been *left behind* by glacial lakes and marine inundations (therefore a "relict" as defined by Ekman and discussed by Holmquist 1959), one could interpret their distribution another way: that is, these animals have gained access to fresh water using glacial water-systems.

It may be that these are marine organisms that, because of competition from more advanced marine relatives, were becoming relicts in the marine situation, and to avoid organic competition had evolved a more advanced physiological system (holeuryhalinity) to live in very brackish estuarine situations. This resulted in a certain amount of "pre-adaptation" and enabled the animals to survive when introduced to fresh water by glaciation. In fresh water they have found a virtually unoccupied niche (deep water) and have taken advantage of it. They can no more be considered relicts than other ecologically restricted freshwater organisms, such as lake trout, which formerly enjoyed a wider range, but which have become restricted because of ecological conditions. Probably the post-glacial range of this group is wider than it has ever been in the past. A better term for this heterogeneous group would perhaps be either "glacial opportunists" or "glacial immigrants".



Appendix 1

Physicochemical characteristics of 716 lakes in the study area, eastern North America (see Map 20)

Lake numbers 1-327 are positive for the community; lake numbers 328-67 are negative lakes inside the glacial-lake boundaries; lake numbers 358-716 are negative lakes outside the glacial-lake boundaries. All locality names are from Canada and United States Topographic Map Series. Unless otherwise specified, all parameters were measured by the author.

Abbreviations

No.	Base-map lake number (Map 20)
Elev.	Elevation of lake a.s.l. (± feet)
A _s	Surface area (hectares)
z _s	Maximum depth (meters)
T _m	Maximum bottom temperature (°C)
zO ₂	Depth of dissolved oxygen less than 2 ppm (meters)

TH	Total hardness (ppm)
pH E-H	pH epilimnion-hypolimnion
SDV	Secchi disk visibility (meters)
O ₂ C	Oxygen consumption (ppm)
Co.	Colour (ppm)

No.	Locality	Elev.	A _s	z _s	T _m	zO ₂	TH	E	pH	H	SDV	O ₂ C	Co.	Order
<i>Positive lakes within glacial-lake boundaries</i>														
1.	Baudeau	1220	1827	27	9.5	-*	8	6.1	6.0		3.9	9.5	25	II
2.	Bethoulat	1330	1540	24	9.4	-	10	6.2	6.0		3.0	8.5†	30	II
3.	Albanet	1279	33500	48	6.0	-	17	6.6	6.3		3.6	5.3	20	II
4.	Mistassini	1229	100000	184	4.0	-	26	7.0	6.6		9.0	4.7	30	I
5.	Opataca	1180	5148	13	13.5	-	25	6.8	6.4		3.9	6.0†	20	II
6.	Waconichi	1250	6774	34	9.5	-	42	7.8	7.2		5.7	3.8	5	II
7.	Lymburner	1340	86	24	5.5	-	45	7.4	7.1		6.3	1.0†	5	I
8.	Bourbeau	1358	459	40	4.0	-	76	7.8	7.4		6.6	2.9	5	I
9.	Gilman	1270	134	25	6.0	-	60	7.8	6.7		3.9	4.2	5	II
10.	Chibougamau	1246	17647	56	9.0	-	36	7.2	6.6		5.4	6.7	10	II
11.	Armitage	1310	287	10	13.5	-	20	6.5	6.1		2.7	7.3†	40	II
12.	Antoinette	1250	115	27	6.0	-	50	7.0	6.0		4.8	2.0†	5	II
13.	Caché	1270	220	23	7.2	-	35	7.5	6.4		4.1	4.0†	5	II
14.	Obatogamau	1218	6067	27	7.5	-	20	6.7	5.8		2.8	8.4	30	II
15.	Malo	1320	268	15	9.3	-	15	6.8	6.2		3.0	7.0†	25	II
16.	Rohault	1270	4631	21	10.0	-	15	6.7	6.3		4.1	3.3	10	II
17.	Gabriel	1290	1215	15	13.0	-	10	6.4	6.0		2.9	2.6	30	II

No.	Locality	Elev.	A _s	Z _s	T _s	zO ₂	TH	E	pH	SDV	O ₂ c	Co.	Order
18.	Presqu'île	1185	968	14	11.3	-	20	6.9	6.7	3.0	7.0	30	II
19.	Caopatrina	1198	4086	12	17.0	-	10	6.4	7.0†	2.6	7.0†	30	III
20.	de la Surprise	1223	4966	27	9.0	-	12	6.4	6.3	2.8	6.7	40	II
21.	Sylvie	1290	134	18	8.0	-	12	6.3	5.5	2.1	9.0†	50	II
22.	Hébert	1278	2794	15	16.1	-	10	6.2	4.7	4.8	4.7	10	III
23.	Lacroix	1270	909	12	17.5	-	12	6.5	6.6	2.1	8.9	50	III
24.	Doda	1109	11225	18	12.5	-	15	6.6	6.6	3.3	6.2	30	III
25.	Opemiska	1176	6010	14	11.5	-	20	7.2	7.1	3.0	1.7	30	II
26.	la Trêve	1120	4823	15	12.5	-	15	6.6	6.2	3.3	5.5†	20	II
27.	Renault	990	373	25	6.0	-	14	6.2	5.6	2.4	8.0†	50	II
28.	Goeland	820	29667	11	16.0	-	18	6.6	1.2	6.5†	20	20	III
29.	Mattagami	796	19618	12	16.0	-	25	6.8	1.1	6.0†	20	20	III
30.	Waswanipi	830	17704	11	16.0	-	15	6.5	1.5	7.0	30	30	III
31.	Bachelor	960	928	15	7.5	-	25	6.7	6.1	1.5	6.5†	20	II
32.	Pustimica	925	5435	27	7.0	-	15	6.0	5.9	2.3	5.0†	25	II
33.	Madeleine	1020	1224	12	16.0	-	40	7.2	3.3	3.3	5.5†	15	III
34.	Quevillon	940	4105	13	16.0	-	20	6.6	2.1	7.0†	30	30	III
35.	Cuvillier	1230	952	21	9.3	-	5	5.7	5.5	3.2	7.5†	30	II
36.	Wettnagami	1205	1732	60	6.0	-	8	6.1	5.6	3.4	5.2	30	II
37.	Maseres	1265	1847	27	8.0	-	11	6.5	5.7	3.4	6.4	40	II
38.	Megiscan	1260	2871	18	15.5	-	10	5.7	5.5	2.2	7.5†	40	II
39.	Valmy	1230	698	39	5.5	-	12	6.1	5.6	3.0	9.0†	50	I
40.	Berthelot	1225	527	18	9.0	-	10	5.9	5.6	2.8	8.0†	45	II
41.	Faillon	1164	2344	82	4.5	-	10	5.7	5.2	2.8	6.5†	30	I
42.	Matchi-Manitou	1120	3550	100	4.0	-	17	6.1	5.6	3.9	5.0†	15	I
43.	Gueguen	1060	3971	14	16.4	-	12	6.5	6.0	1.3	6.9	30	II
44.	Villebon	1070	4058	14	15.0	-	12	6.5	6.2	1.9	7.2	40	II
45.	Granet	1050	3378	17	14.8	15	12	6.2	5.5	2.5	8.4†	30	II
46.	Grand Victoria	1065	8268	11	18.0	-	10	6.1	5.8	2.4	5.8	30	III
47.	Parent	990	7943	22	15.0	-	12	6.0	5.8	1.9	7.0†	40	II
48.	Fiedmont	990	823	28	6.0	-	25	6.9	6.2	0.9	4.0†	10	II
49.	Mourier-Lemoine	980	3205	12	14.0	-	24	6.5	1.2	1.2	6.0†	20	III
50.	Priessac	958	8690	15	17.0	-	25	6.6	1.5	1.5	5.0†	20	III

Appendix 1

No.	Locality	Elev.	A _s	Z _s	T _s	zO ₂	TH	E	pH	SDV	O ₂ C	Co.	Order
51.	Loia	990	1043	6	15.0	-	35	6.7		1.0	7.0†	20	III
52.	Chicobi	980	1732	6	15.0	-	42	6.7		0.2	8.0†	30	III
53.	Turgeon	950	2000	10	14.0	-	45	7.0		0.7	6.5†	30	III
54.	Macamic	927	4125	11	15.0	-	50	7.5		1.0	7.0†	20	III
55.	Abitibi	868	85760	5	17.0	-	64	8.4		0.1	3.5	10	III
56.	James Bay ‡	0		100	?	?	?	?		?	?	?	
57.	Remi	746	2414	9	15.8	-	120	8.5		1.6	6.0†	10	III
58.	Papakomelka	1005	180	45	5.0	-	70	7.8	6.8	3.2	7.5†	35	I
59.	Clarice	990	297	19	9.0	-	42	7.2		3.4	4.5†	20	II
60.	Duparquet	896	3617	10	15.0	-	50	7.1	6.7	1.6	6.5†	30	II
61.	Caron	880	1942	67	4.5	-	50	7.4	6.0	3.2	5.9†	20	I
62.	Opasatica	866	4220	42	6.0	-	43	7.6	6.5	1.8	3.5†	10	II
63.	Larder	950	3435	37	6.0	-	80	8.4	7.1	2.1	5.5†	10	II
64.	Mistnikon	1030	803	26	7.0	-	40	7.3	6.1	3.3	6.5†	30	II
65.	Round	886	1148	28	5.8	-	85	8.5	6.6	1.0	6.0	20	II
66.	Raveni	950	507	48	4.2	-	35	6.9	6.6	3.2	5.0†	20	I
67.	St. Anthony	990	483	24	6.5	-	26	7.0	5.8	4.2	4.0†	10	II
68.	Kinogami	820	734	36	5.0	-	85	8.3	6.9	2.7	8.3†	40	I
69.	Wendigo	750	388	27	5.8	-	45	7.1	6.2	4.9	4.2†	20	II
70.	Rémigny	880	2613	27	6.0	-	32	7.0	6.5	1.1	6.4	30	II
71.	Mountain	930	476	45	8.0	-	40	7.5	6.3	2.7	7.0†	40	II
72.	Mendelssohn	925	433	33	5.5	-	20	6.4	6.0	8.1	3.5†	5	I
73.	Hammond	915	100	24	5.5	18	42	7.5	6.3	6.6	2.0†	5	I
74.	Sinard	863	14355	47	7.5	-	15	6.3	5.6	1.2	6.0†	20	II
75.	des Quinze	850	11149	30	8.3	-	15	6.3	6.3	0.8	5.1	20	II
76.	Temiskaming	585	100000+	140	4.0	-	39	7.0	7.2	0.8	2.6†	5	I
77.	Gilles	950	70	28	5.5	-	55	8.0	6.6	6.5	2.3†	5	II
78.	Bay	903	1063	24	6.7	-	24	6.9	6.4	3.3	5.5†	10	II
79.	Kitt	935	80	30	6.0	-	25	6.6	5.8	5.1	5.5†	20	II
80.	Barriar	1020	45	30	5.0	-	20	6.1	5.8	5.4	3.0†	5	I
81.	Lady Evelyn	930	5611	31	7.0	-	20	6.4	5.6	5.7	3.3†	10	II
82.	Obabika (upper)	932	1000	36	6.0	-	24	6.5	5.9	7.5	2.5†	5	II
	(lower)		1376	27	7.0	-	24	6.5	6.1	6.0	2.4	5	II
83.	Kokoko	975	496	42	6.5	-	38	6.6	5.8	5.1	4.4†	15	II

No.	Locality	Elev.	A.	Z.	T.	pO ₂	TH	E	pH	SDV	O ₂	Co.	Order
84.	Kanichee	990	224	32	4.5	-	30	6.9	6.1	3.9	6.0†	25	I
85.	Hait	980	714	37	5.0	-	35	6.9	6.3	4.2	5.9	20	I
86.	Rib	1025	578	47	7.0	-	35	6.9	6.5	6.6	2.1	5	II
87.	Rabbit	938	2000	25	9.0	-	35	6.9	6.5	4.8	5.3†	15	II
88.	Herridge	1043	194	30	4.5	-	27	6.7	6.1	3.6	6.1†	20	I
89.	Jumping Caribou	1028	404	34	7.5	-	27	6.8	6.0	4.7	4.5†	10	II
90.	Tenagami	962	20000	61	6.0	-	26	6.8	6.6	7.5	6.3	10	II
91.	Manitou	872	326	42	5.5	-	28	6.6	6.5	7.5	1.8	5	I
92.	Waswakash	827	415	24	5.0	-	34	6.8	6.7	5.3	2.3	5	I
93.	Cucumber	925	44	24	5.0	-	34	6.8	6.7	5.7	2.3	5	I
94.	Red Cedar	914	1868	35	8.5	-	38	6.9	6.4	3.3	7.5†	25	II
95.	Marten	941	862	49	5.5	-	25	6.4	5.9	3.6	6.5†	20	II
96.	Beauchêne	840	3205	33	6.5	-	13	6.4	5.7	4.6	4.7	10	II
97.	Tee	855	411	55	5.0	-	14	6.2	5.5	5.8	1.5†	5	I
98.	Kipawa	885	30000	89	5.2	-	15	6.1	5.4	7.5	2.5	5	I
99.	Ostabonique	930	2354	41	5.5	-	15	5.9	5.5	4.2	3.6	10	I
100.	Cooks	940	145	36	4.9	-	12	6.1	5.5	5.4	2.6	5	I
101.	Pomeroy	930	1005	57	5.0	-	18	6.1	5.5	7.9	2.2	5	I
102.	Saint-Amand	940	86	45	5.5	-	20	6.6	5.9	3.9	3.5	5	I
103.	Argentier	920	153	25	6.5	-	20	6.6	5.6	5.1	3.0†	5	II
104.	Wanapitei	876	12519	109	4.8	-	30	6.8	6.7	6.3	3.0†	10	I
105.	Windy	1093	1200	55	4.3	-	18	6.5	5.9	5.0	1.0†	5	I
106.	Fairbank	985	653	44	4.2	-	28	7.0	6.3	5.4	1.8†	5	I
107.	Panache	733	7025	30	8.2	-	26	6.6	6.0	6.9	2.8†	5	II
108.	Trout	680	800	41	5.5	-	26	6.3	5.5	3.9	5.3†	20	I
109.	Tomiko	800	1542	24	12.8	22	16	6.1	5.5	3.3	7.0†	20	II
110.	Tilden	936	396	25	8.9	20	12	5.7	5.4	4.2	6.5†	20	II
111.	Marin	740	363	42	5.0	-	15	6.5	5.6	3.7	3.0†	5	I
112.	Talon ‡	636	1293	58	4.5	-	30	6.8	6.5	4.2	3.0†	5	I
113.	Smith ‡	700	183	28									
114.	Turtle ‡	663	210	50	4.5	-	30	6.9	6.2	4.0	3.0†	5	I
115.	Trout	663	1850	74	5.0	-	30	6.9	6.3	3.9	3.0†	10	I
116.	Nipissing ‡	644	83820	51	8.3	40	76	7.4	6.0	4.5	5.5†	10	II
117.	Restoule	691	1223	24	6.5	-	25	6.3	5.6	3.6	6.0†	20	II

No.	Locality	Elev.	A _s	Z _s	T _s	zO ₂	TH	E	pH	SDV	O ₂ C	Co.	Order
118.	Waukimakog	600	920	29	9.0	-	25	6.7	5.6	2.7	6.5†	30	II
119.	Fowke	981	241	51	5.0	-	35	7.3	6.4	5.1	3.1†	5	I
120.	Wawashkesh	880	1500	39	6.0	-	20	6.7	5.6	3.3	4.5†	10	II
121.	Eagle	1166	901	21	6.5	-	17	6.5	5.6	5.1	6.0†	30	II
122.	Bernard	1082	2074	45	5.0	-	26	6.7	5.8	3.9	4.5	5	I
123.	Sand †	1110	565	38	6.0	-	20	6.7	5.9	4.0	3.8†	5	II
124.	Pickrel	1050	528	33	6.5	-	26	6.5	5.5	3.0	5.7†	20	II
125.	Bethune †	1100	111	25	5.0	20	25	6.4	5.5	2.5	7.2†	25	II
126.	Ebe	956	902	24	6.0	-	25	6.6	5.7	3.0	5.9†	20	II
127.	Ahmik †	917	1800	27	7.5	-	24	6.6	5.6	3.2	5.0†	10	II
128.	Manitouweabing	700	769	34	6.5	-	25	6.7	5.5	2.7	6.6†	20	II
129.	Mill	650	611	38	7.0	-	25	6.3	5.6	3.0	6.2†	25	II
130.	Blackstone	655	510	57	5.5	-	25	6.6	5.6	6.3	2.4†	5	I
131.	Kashe	796	478	21	7.5	20	15	6.7	5.5	3.0	8.0†	30	II
132.	Simcoe	718	76800	43	9.5	35	150	8.5	8.3	6.6	1.0	5	II
133.	Ontario †	248	1925120	300	4.0	-	185	8.5	7.5	6.0	2.3	5	I
134.	Erie †	572	2542000	57	6.0	30	180	8.5	7.0	4.8	3.0	10	II
135.	Huron †	573	6000000	240	4.0	-	110	8.0	6.8	8.0	1.5	5	I
136.	Muskoka	739	5193	60	5.5	-	25	6.5	5.8	5.4	3.2†	5	I
137.	Rousseau	735	5186	60	4.0	-	24	6.7	5.9	6.6	2.0†	5	I
138.	Joseph	735	4268	90	5.0	-	20	6.7	5.7	9.6	1.3†	5	I
139.	Skeleton	920	1858	62	5.0	-	26	6.5	5.6	9.6	1.0†	5	I
140.	Buck	955	210	24	6.0	-	25	5.7	5.0	1.5	9.8	50	II
141.	Vernon	931	1426	32	6.0	-	25	6.1	6.0	3.3	6.8†	30	II
142.	Mary	921	800	54	5.5	-	25	6.6	5.7	3.3	5.5†	20	I
143.	Fairy	931	667	74	5.0	-	22	6.5	5.2	3.6	5.9	20	I
144.	Peninsula	931	650	33	5.5	-	20	6.8	5.6	5.2	4.0†	10	I
145.	Lake of Bays	1034	6104	53	4.0	-	25	6.5	6.1	4.2	4.3†	10	I
146.	Halls	1069	547	69	5.0	-	25	6.3	6.0	7.2	1.8†	5	I
147.	Maple †	1032	331	31	6.5	-	32	6.8	6.1	6.3	3.8†	5	II
148.	Beech	1013	133	28	7.0	-	34	6.9	6.1	5.1	4.3†	5	II
149.	Boshung	1013	690	60	4.5	-	32	7.2	6.1	6.3	2.9†	5	I
150.	Twelvemile †	1009	345	25	8.0	-	34	7.1	6.2	6.0	2.8†	5	I

No.	Locality	Elev.	A.	Z _m	T _m	zO ₂	TH	E	pH	H	SDV	O ₂ c	Co.	Order
151.	Mountain ‡	1005	309	30	6.5	-	35	7.2	6.1	5.5	5.5	3.3†	5	II
152.	Horseshoe	1002	282	21	7.5	20	34	6.7	5.8	6.9	6.9	2.9†	5	II
153.	Minden	1000	204	31	7.0	-	35	7.1	6.0	5.5	5.5	3.2†	5	II
154.	Bob	924	210	56	5.0	-	34	6.7	5.8	6.9	6.9	1.7†	5	I
155.	Big Trout ‡	1010	190	26	6.5	-	30	6.8	5.5	5.0	5.0	4.4†	10	II
156.	Gull	885	996	39	6.5	-	30	7.1	6.3	5.4	5.4	2.6†	5	II
157.	Shadow	850	303	22	8.5	15	38	7.1	6.1	5.7	5.7	4.7†	10	II
158.	Crystal	933	468	34	5.5	25	102	8.5	6.7	4.8	4.8	5.2†	10	I
159.	Mississagua	966	581	34	6.0	-	35	6.9	6.1	4.2	4.2	5.5†	20	II
160.	Stony ‡	769	2000	30	7.5	15	110	8.5	7.0	3.0	3.0	5.4†	10	II
161.	Weslemkoon	1030	1933	55	6.0	-	18	6.7	6.1	5.8	5.8	3.2†	5	II
162.	Mackie	825	156	22	7.0	-	50	8.0	6.9	6.5	6.5	1.5†	5	II
163.	Wensley	860	411	57	5.0	-	60	8.2	7.2	8.9	8.9	1.0†	5	I
164.	Big Ohlman	900	38	39	4.5	12	55	8.4	7.1	6.0	6.0	3.8†	5	I
165.	Palmerston	890	382	53	5.0	-	103	7.5	6.9	5.7	5.7	3.2†	5	I
166.	MacKavoy	885	19	21	6.0	20	60	8.3	6.3	3.9	3.9	4.4†	15	II
167.	Mazinaw	870	1572	135	4.0	-	36	7.3	6.6	6.0	6.0	5.3	20	I
168.	Shabomeka	895	168	31	5.0	-	44	7.6	6.5	5.4	5.4	4.7	10	I
169.	Plevna	895	175	18	7.0	15	95	7.4	6.0	5.5	5.5	2.3†	5	II
170.	Cross	787	1540	27	8.0	18	44	6.9	6.2	3.9	3.9	4.3	5	II
171.	Mississagagon	878	378	26	5.5	21	102	8.3	7.1	6.6	6.6	2.8	5	I
172.	Kaswakamak	854	1043	24	6.7	-	55	8.0	6.7	6.3	6.3	3.3†	5	II
173.	Clarendon	831	2336	21	10.5	15	35	7.5	6.5	4.1	4.1	4.8†	10	II
174.	Silver	570	243	25	7.0	18	154	8.2	7.2	4.2	4.2	3.8†	5	II
175.	Sharbot	638	1438	34	6.5	32	125	7.2	6.8	4.1	4.1	5.7†	10	II
176.	Pike	476	312	42	6.5	16	75	7.3	6.3	4.1	4.1	4.1†	10	II
177.	Crowe	550	476	33	8.0	21	70	8.0	6.8	4.5	4.5	3.4	5	II
178.	Bobs	532	2420	25	6.0	18	110	8.5	6.7	5.1	5.1	2.1†	5	II
179.	Wolfe	447	885	28	7.5	14	130	8.5	7.0	3.9	3.9	4.2†	10	II
180.	Big Rideau	406	3568	108	4.5	-	100	8.5	7.0	4.5	4.5	3.3	5	I
181.	Otter	408	595	34	8.8	12	110	8.5	7.0	3.3	3.3	4.0†	15	II
182.	Canoe	454	230	42	6.1	-	90	8.5	7.0	6.0	6.0	1.4	5	II
183.	Desert	444	378	68	5.0	-	110	7.4	7.3	5.1	5.1	2.5†	5	I
184.	Knowlton	430	156	33	3.9	20	130	8.3	7.0	4.3	4.3	3.8	5	I
185.	Birch	450	193	36	5.9	33	85	8.3	6.9	5.5	5.5	1.6†	5	II

Appendix 1

No.	Locality	Elev.	A _v	z _m	T _m	zO ₂	TH	E	pH	SDV	O ₂ c	Co.	Order
186.	Clear	520	143	57	4.0	—	35	7.1	6.3	8.4	2.0†	5	I
187.	Buck	434	658	39	6.0	27	90	7.4	6.8	5.1	3.0†	5	II
188.	Indian	268	189	28	8.9	18	110	8.3	7.2	4.3	3.1†	5	II
189.	Devil	430	997	40	5.0	30	103	8.0	7.0	5.4	2.1	5	I
190.	Charleston (BW) (RB)	278	1200	44	11.2	—	130	8.5	7.8	3.7	4.0	10	II
			500	92	4.5	—	130	8.5	7.5	5.7	3.5	5	I
191.	Red Horse	280	224	39	4.3	16	135	8.5	7.1	4.2	4.3†	15	I
192.	Loughborough	410	1782	34	9.5	30	155	8.3	7.2	4.8	2.8†	10	II
193.	Sydenham	470	398	33	5.5	25	140	7.9	7.0	3.1	4.0†	10	I
194.	Muskat	404	1169	56	7.0	45	100	8.4	7.1	1.5	5.5†	20	II
195.	Golden	556	3531	25	9.9	20	45	8.0	6.9	4.2	6.1	20	II
196.	Round	560	2944	44	8.0	—	34	6.8	6.0	4.5	4.0	20	II
197.	Grand †	730	717	39	6.0	—	34	7.5	7.0	4.5	5.0†	10	II
198.	St. Andrews †	730	89	24	5.0	—	26	7.0	6.5	4.0	6.0†	15	I
199.	Guthrie †	895	45	33									
200.	Little Carcajou †	780	11	48									
201.	Carcajou †	829	82	43									
202.	Greenleaf †	819	61	72									
203.	Eustache †	900	40	100									
204.	White Partridge †	1155	563	45									
205.	Cartier †	590	260	21	5.0	—	34	7.5	7.0	3.0	7.0†	30	I
206.	Ottawa River	400		150	4.5	—	35	6.7	6.3	4.3	4.5†	20	I
207.	McSourley †	650	28	33									
208.	Waterloo †	907	178	28	5.0	—	20	7.0	6.0	3.9	5.8†	20	I
209.	Wendigo †	1040	160	28	6.0	—	20	7.0	6.0	4.2	5.0†	10	II
210.	Papineau †	995	230	24									
211.	Lauder †	1043	200	35									
212.	Guilmette †	1073	64	33									
213.	Radiant †	914	627	36									
214.	Cedar †	1007	2560	58	8.0	—	34	6.7	6.7	4.0	5.2†	5	II
215.	Hogan †	1250	1230	29									
216.	Gilmour †	1022	162	25									
217.	Laurie †	1026	63	28									
218.	Gouinlock †	1125	69	46									

No.	Locality	Elev.	A.	Z.	T.	zO ₂	TH	E	pH	SDV	O ₂ c	Co.	Order
219.	Cauchon ‡	1066	271	43									
220.	Whitebirch ‡	1080	62	28									
221.	Mink ‡	1034											
222.	Kioshkokwi ‡	993	222	43									
223.	Wilkes ‡	1129	1024	39									
224.	Threemile ‡	1250	1434	32									
225.	Biggar ‡	1222	402	37									
			366	31									
226.	Waskigomog ‡	1215	1331	32									
227.	Big Gibson ‡	500	48	38									
228.	Nottawissi	795	1254	36	5.5		20	6.4	5.6	4.2	5.3†	20	I
229.	Kettle	810	60	57	5.0	—	10	6.0	5.6	6.9	3.3†	5	I
230.	Crevier	800	409	30	5.0	25	35	7.3	6.4	4.5	5.2†	10	I
231.	Marguerite	810	557	48	5.0	—	15	6.3	5.9	7.5	2.0†	5	I
232.	Cobble	800	71	21	5.0	15	15	6.2	5.6	3.5	8.0†	30	I
233.	Petawaga	750	2233	44	6.0	—	10	6.3	5.8	5.1	2.0†	8	II
234.	Baskatong	732	20000+	82	5.1	—	10	6.5	6.0	3.6	5.5	20	I
235.	Chopin	830	485	30	7.6	—	11	6.3	5.4	2.9	5.1	20	II
236.	Polonais	810	1015	40	5.3	—	15	6.6	5.6	3.6	5.6	20	I
237.	Piscatosine	732	2005	50	5.5	—	20	6.6	5.9	4.2	6.0†	20	I
238.	Tapani	818	578	24	6.7	—	23	6.9	6.3	3.0	4.6	20	II
239.	Chinard	900	120	57	4.2	—	16	6.6	5.5	6.9	3.4	6	I
240.	Gravel	880	133	36	5.6	—	25	7.0	6.0	3.9	2.8	10	I
241.	Moreau	788	406	33	5.5	—	40	7.7	6.2	4.6	2.5†	10	I
242.	des Écorces	753	516	37	5.8	—	35	7.7	6.2	1.9	4.3†	20	II
243.	Pope	830	174	30	6.0	—	30	6.6	5.5	4.6	6.5	20	II
244.	Gatineau	850	54	30	5.0	25	38	6.9	6.1	6.9	2.5	5	I
245.	des Îles	660	1472	39	8.0	—	45	8.0	6.6	3.9	3.0	10	II
246.	du Cerf	680	1305	72	4.5	—	35	7.2	6.8	5.4	3.1†	5	I
247.	Corbeau	710	182	57	5.0	—	35	7.1	6.6	4.2	4.5†	10	I
248.	Serpent	690	187	60	4.0	—	45	7.4	6.7	5.7	4.0†	10	I
249.	O'Neil	650	80	37	5.5	—	40	7.3	6.5	5.7	2.5†	5	I
250.	Poisson Blanc	660	4420	75	5.0	—	30	6.9	6.3	7.8	2.0†	5	I
251.	Leamy	790	18	28	4.6	—	16	6.5	5.5	2.1	8.5†	40	I
252.	Serpent	840	41	24	4.6	—	17	6.5	5.5	3.3	7.1†	20	I

No.	Locality	Elev.	A.	Z.	T _m	±O ₂	TH	E	pH	H	SDV	O ₂ c	Co.	Order
253.	Quinn	785	49	28	4.2	25	15	6.7	5.5	5.5	3.8	6.8†	20	I
254.	de la Vieille	810	301	28	4.8	-	18	6.6	5.5	4.2	4.2	6.2†	20	I
255.	Tomasine	758	393	28	8.0	-	10	6.4	5.5	4.0	4.0	6.0†	20	II
256.	Savary	790	403	55	4.2	-	8	6.7	5.5	5.1	5.1	5.1†	10	I
257.	Rond	739	547	29	8.5	-	18	6.2	5.5	4.1	4.1	6.3†	15	II
258.	Désert	740	1554	42	7.0	-	20	6.5	5.9	5.4	5.4	4.5†	8	II
259.	Bras Coupé	755	675	48	5.0	-	30	6.8	6.4	5.7	5.7	4.6	5	I
260.	Lytton	745	221	30	7.5	-	25	6.6	5.5	6.5	6.5	3.8	5	II
261.	Pocknock	580	231	36	5.5	-	35	7.0	6.3	5.7	5.7	2.9†	10	I
262.	Grand Cèdres	560	740	37	8.0	-	70	7.3	7.1	6.0	6.0	2.4†	5	II
263.	Petit Cèdres	555	257	37	6.0	-	75	7.3	7.1	6.3	6.3	2.5†	5	II
264.	Murray	612	269	24	7.0	12	45	7.5	6.5	2.8	2.8	5.0†	20	II
265.	à l'Achigan	653	128	25	4.5	20	62	7.3	6.3	6.0	6.0	6.4	20	I
266.	Kensington	710	298	39	5.0	30	68	7.2	6.4	6.3	6.3	2.0†	5	I
267.	Thirty-one Mile	540	3867	33	5.2	-	72	8.3	6.7	6.7	6.7	1.0	5	I
268.	Vert	626	150	96	4.0	80	95	8.5	7.4	12.2	12.2	0.5	5	I
269.	Bangall	625	109	43	5.0	40	78	8.5	6.7	7.8	7.8	3.2	5	I
270.	Pemichangan (A)	555	868	35	5.5	15	85	8.5	6.9	6.4	6.4	4.0	5	I
	(B)		1000	54	4.5	-	85	8.5	6.7	7.9	7.9	2.8	5	I
271.	Heney	470	1230	32	7.0	20	75	8.5	6.5	4.9	4.9	3.0†	10	II
272.	Bitobi	490	208	29	5.0	25	35	7.0	6.0	3.9	3.9	6.2†	10	I
273.	Cameron	480	149	24	5.0	20	45	7.1	6.3	3.8	3.8	5.2	10	I
274.	Roddick	483	501	45	5.0	40	80	8.5	6.5	7.3	7.3	2.2	5	I
275.	Blue Sea	541	1440	59	5.5	-	90	8.5	7.0	6.0	6.0	2.3	10	I
276.	Paquin	590	155	48	4.5	36	95	8.5	6.8	10.0	10.0	3.0	5	I
277.	Profond	580	94	33	4.4	25	55	8.0	6.0	7.8	7.8	2.5	5	I
278.	Danford	581	115	34	6.0	20	86	8.1	7.0	6.3	6.3	2.4†	5	II
279.	Gruice	542	150	37	5.0	-	55	8.3	6.6	5.6	5.6	2.0†	8	I
280.	Thorne	618	86	29	5.4	25	60	7.6	6.5	6.0	6.0	1.8†	5	I
281.	Johnston	503	54	34	5.0	25	70	7.8	6.0	3.3	3.3	5.5†	20	I
282.	MacGregor	466	538	43	7.5	-	62	7.5	7.0	5.1	5.1	3.0†	8	II
283.	McFee	669	69	48	4.0	45	35	6.7	6.0	8.7	8.7	2.9†	5	I
284.	Dodds	633	53	42	4.0	38	26	6.8	6.4	8.1	8.1	1.5†	5	I
285.	Echo	774	514	60	4.5	-	35	7.0	6.3	9.6	9.6	1.0†	5	I

No.	Locality	Elev.	A _s	Z _s	T _m	zO ₂	TH	E	pH	SDV	O ₂ c	Co.	Order
286.	Saint-Sixte	740	141	45	5.0	-	35	6.8	5.5	6.0	4.3	10	I
287.	Heart	670	105	41	5.5	-	35	7.1	5.5	5.7	3.0†	5	I
288.	Britannique	715	154	60	4.5	-	45	7.4	6.3	6.6	1.5†	5	I
289.	Lady	760	95	45	4.5	-	35	6.8	6.0	9.3	0.8†	5	I
290.	Hawk	745	151	39	4.5	-	35	6.9	6.1	8.7	1.4†	5	I
291.	Gull	730	115	39	4.5	33	51	7.4	6.5	9.6	2.4	10	I
292.	la Blanche	590	711	14	10.0	12	35	6.8	6.0	2.7	5.0†	10	II
293.	Papineau	525	963	81	4.5	-	25	6.8	6.0	10.7	3.2	5	I
294.	Whitefish	490	49	16	5.0	-	34	6.9	6.2	3.6	5.0†	20	I
295.	Louisa	890	364	33	5.4	20	20	7.0	6.0	6.0	4.2	5	I
296.	Barron	910	87	31	5.5	-	16	6.5	5.6	3.0	6.3†	40	I
297.	Connelly	690	104	19	6.0	-	24	6.8	5.6	3.9	4.5†	20	II
298.	l'Achigan	690	665	21	6.0	-	25	6.7	6.2	4.8	2.8†	10	II
299.	des Piles	543	503	66	4.5	-	14	6.5	6.2	10.2	1.0†	5	I
300.	Saint-Jean	312	20000 +	68	4.5	-	15	6.2	5.9	2.7	7.8	30	I
301.	Saguenay estuary			60	1.0	-	20 ppt			3.0	1.0†	5	I
302.	Temiscouata	490	5502	66	4.0	-	55	7.6	7.4	2.9	2.9	5	I
303.	Saint-François	900	3456	37	4.0	-	43	6.8	6.5	2.1	8.6	40	I
304.	Aylmer	820	2528	30	4.9	-	36	6.9	6.0	2.0	8.8†	40	I
305.	Grand Brompton	780	729	44	7.0	-	30	6.5	6.0	3.9	6.3	10	II
306.	Massawippi	570	1572	82	4.0	-	102	8.0	7.3	4.5	3.4	5	I
307.	Orford	905	95	47	4.0	30	55	7.0	6.7	8.1	1.7	5	I
308.	Lovering	803	426	25	9.5	-	38	6.6	5.5	3.6	6.7	10	II
309.	Memphremagog	683	7737	117	5.0	-	70	7.5	6.9	5.7	2.1	5	I
310.	Champlain	62	120000	120	4.0	-	45	8.0	7.0	3.9	3.0	5	I
311.	Dunmore	571	386	30	6.5	-	40	7.8	6.5	3.9	4.0†	10	II
312.	Sussex	498	80	36	5.5	-	35	7.0	6.2	9.0	1.5†	5	I
313.	St. Catherine	480	500	21	7.0	15	55	8.0	6.0	5.4	3.0†	10	II
314.	George	322	11280	56	5.0	-	42	6.9	6.0	9.0	1.0	5	I
315.	Sylvia	670	113	42	5.0	-	130	8.2	7.1	8.4	1.0†	5	I
316.	Lake of the Woods	300	71	25	6.0	20	45	7.4	6.6	4.2	4.0†	10	II
317.	Sixberry	345	42	27	5.5	20	40	7.6	6.4	2.8	5.2†	15	I
318.	Millsite	301	200	22	6.0	-	40	7.2	7.0	5.7	1.0†	5	II
319.	Green ‡	419	27	59	7.2	20	150	8.0	6.8	9.0	1.0†	5	II
320.	Skaneateles	867	3584	90	6.3	-	110	8.0	7.8	9.3	2.0	5	II

No.	Locality	Elev.	A _s	Z _m	T _m	ZO ₂	TH	E	pH	SDV	O ₂ c	Co.	Order
321.	Owasco ‡	710	2560	48	7.3	—	110	8.4	7.8	4.2	3.0	10	II
322.	Cayuga ‡	381	16800	120	4.1	—	104	8.5	8.0	6.3	2.0	5	I
323.	Seneca ‡	444	17100	180	4.0	—	110	8.3	7.7	9.0	2.5†	5	I
324.	Keuka	750	2000	48	4.8	—	110	8.4	7.6	4.5	2.5†	5	I
325.	Canadseigua ‡	686	4250	75	4.3	—	120	8.3	7.9	4.5	2.0†	5	I
326.	Hemlock	905	800	27	6.0	20	95	7.8	7.4	3.0	4.5†	10	II
327.	St. John estuary	0		27	13.0	—	35	7.1	6.5	2.5			
<i>Negative lakes within glacial-lake boundaries</i>													
328.	Nellie	900		24	6.0	15	35	7.3	6.4	3.7	4.3†	20	II
329.	Perry	1050	106	28	6.0	14	70	8.5	7.0	7.8	1.0†	5	II
330.	Nighthawk	890	8523	5	18.0	—	70	8.0		0.5	2.0†	5	III
331.	Kenogami	1030	1000	6	18.0	—	43	8.0		1.3	4.0†	10	III
332.	Castagnier	980		3	20.0	—	24	6.7		0.4	6.3†	30	III
333.	Malartic	960		25	17.0	—	33	6.7		1.0	6.0†	30	III
334.	Descelles	830		30	9.2	20	12	5.7	5.5	2.0	9.8†	50	II
335.	Brisebois	830		18	8.5	10	20	6.7	5.5	3.0	8.6	40	II
336.	Wakimika	950		18	9.0	—	23	6.3	5.6	6.0	3.0†	5	II
337.	Lee	890	131	21	7.5	—	28	6.4	5.5	3.9	5.3†	20	II
338.	Capreol	990		28	5.0	—	20	5.5	5.0	6.9	1.3†	5	I
339.	Little Panache	812		24	5.0	21	45	8.3	6.0	4.5	3.5†	5	I
340.	Kawawaymog	1250		13	15.8	10	25	6.9	6.0	2.8	7.4†	30	II
341.	Otter	676	500	42	5.0	—	17	6.6	5.6	3.9	5.8†	20	I
342.	Clearwater	850	71	28	6.0	25	15	6.5	5.3	6.6	2.8†	10	II
343.	Round	555		10	20.0	—	110	8.5		2.4	6.3†	5	III
344.	Coxvale	850		16	7.0	10	40	6.8	6.0	4.0	4.7†	10	II
345.	Clear	400	170	33	8.5	8	120	8.5	6.3	4.8	2.2†	5	II
346.	Lower Beverly	300	638	24	7.5	10	145	8.3	7.3	2.1	4.8†	10	II
347.	Calabogie	505		36	19.5	—	50	7.0		3.1	5.5†	10	III
348.	Esturgeon	820		27	4.0	?	22	6.5	5.6	5.4	3.9†	10	I
349.	Bois Franc	540		20	4.0	10	80	8.3	6.7	5.4	7.2	20	I
350.	Green	480		22	6.0	10	70	8.0	6.5	4.2	2.3	5	II
351.	Gilmore	480		36	4.5	17	55	8.5	6.2	4.9	2.0	5	I
352.	Plumbago	425	56	27	4.2	13	60	7.9	6.5	2.4	4.4†	10	I

No.	Locality	Elev.	A _s	z	T _s	zO ₂	TH	E	pH	H	SDV	O ₂ c	Co.	Order
353.	Litchfield	588		24	4.0	16	35	7.8	6.3		3.0	5.9†	20	I
354.	Gilmour	600		21	5.5	12	65	8.0	6.6		3.3	8.4†	30	I
355.	Venosta	530		21	6.8	16	75	8.5	7.0		6.6	2.3†	5	II
356.	Manitou	520	40	24	4.5	18	140	8.5	7.6		4.6	2.8	10	I
357.	Saratoga	204	1765	28	6.0	15	85	7.2	6.9		4.8	5.4†	10	II
<i>Negative lakes outside glacial-lake boundaries</i>														
358.	Naocrane	1785		19	8.2	—	15	6.8	6.4		6.9	2.3†	5	II
359.	Vallard	1700		13	13.0	—	10	6.5			3.0	6.8†	30	III
360.	Base Camp	2300		13	11.0	—	15	6.5	6.0		2.7	7.5†	40	II
361.	Indicator	1700		22	9.0	—	8	6.1	5.6		2.4	8.3†	40	II
362.	Petit Temiscamie	1480		43	7.5	—	8	6.0	6.0		4.4	5.5	25	II
363.	Temiscamie	1380		31	9.5	—	12	6.2	6.2		3.0	5.4	25	II
364.	Kallio	1420		18	9.0	—	36	7.4	6.8		4.1	1.3	5	I
365.	Tournamine	1380		10	12.5	—	15	6.5	6.0		3.3	7.0†	30	II
366.	Coldwater	1530		35	6.0	—	10	6.0	6.0		3.3	5.0†	20	II
367.	File-axe	1480		13	12.0	—	15	6.6	6.6		3.6	8.0†	40	II
368.	Margonne	1425		15	10.0	—	25	6.7	6.0		4.2	5.8†	20	II
369.	Ida	1400		5	17.5	—	20	6.5	6.5		2.1	9.5†	50	III
370.	Vimont	1330		25	8.5	—	20	6.5	5.9		4.5	5.0†	15	II
371.	Dufresne	1270		6	14.5	—	17	6.8	6.1		2.1	9.0†	50	II
372.	Charron	1310		6	16.0	—	10	5.5			1.5	8.8†	45	III
373.	Nicabeau	1260		18	15.5	—	20	6.8	6.4		2.9	7.2†	30	II
374.	Aigremont	1320		14	11.9	—	15	6.5	6.0		3.3	6.5†	30	II
375.	d'Églis	1195		24	7.7	—	10	6.4	5.7		2.5	6.0†	30	II
376.	Argenson	1106		27	10.0	—	12	6.3	5.8		3.6	6.5	30	II
377.	Chamouchouane	1240		11	12.0	—	8	6.0	5.7		2.0	8.5†	40	II
378.	Poutrincourt	1280		10	16.5	—	10	6.3			3.0	6.2	40	III
379.	Robert	1302		20	11.0	—	8	6.0	5.5		2.0	7.5†	50	II
380.	Nemagousse	1450		35	6.5	—	12	6.3	6.3		4.8	4.6	20	II
381.	Baptiste	1425		13	8.0	—	10	6.1	6.1		3.5	6.0†	30	II
382.	Gouin	1310		20	10.0	—	8	6.0	6.0		3.8	5.5†	20	II
383.	Pascagama	1380		15	15.5	—	10	5.7	5.5		2.0	7.8†	40	II
384.	Mesplet	1279		5	18.0	—	12	6.5			1.8	6.5†	30	III
385.	Maudé	1290		16	12.0	—	8	5.5	5.0		1.8	9.7†	50	II

No.	Locality	Elev.	A _s	Z _m	T _m	ZO ₂	TH	E	pH	H	SDV	O ₂ c	Co.	Order
386.	Attic	1290		23	7.0	-	10	6.3	5.6	5.6	4.5	5.6†	30	II
387.	Hickey	1190		16	5.5	-	15	6.3	6.1	2.1	2.1	9.2†	50	I
388.	Castonguay	1290		29	6.5	-	8	5.5	5.0	5.0	2.0	6.6	40	II
389.	Desforges	1250		18	7.3	-	12	6.1	5.5	7.0	7.0	1.3†	5	II
390.	Augier	1280		27	5.4	-	8	6.0	5.5	5.5	5.5	2.9†	10	I
391.	Charrette	1270		55	5.0	-	8	5.9	5.6	6.0	6.0	2.3	5	I
392.	Valeis	1279		18	11.0	-	10	5.5	5.3	4.5	4.5	4.6†	20	II
393.	Martin	1480		19	15.5	-	5	5.5	5.0	6.3	6.3	1.5†	5	II
394.	Yser	1340		30	6.0	-	10	5.6	5.3	8.7	8.7	1.1†	5	II
395.	Denain	1300		31	6.0	-	10	5.6	5.3	7.5	7.5	1.2†	5	II
396.	Camachigama	1190		43	6.0	-	12	5.8	5.5	6.0	6.0	2.7†	5	II
397.	Dozois	1135		27	8.5	-	8	5.7	5.5	2.7	5.6	5.6	30	II
398.	Cabonga	1188		28	8.5	21	12	6.4	5.5	4.0	5.1†	5.1†	20	II
399.	O'Sullivan	1300		53	6.0	-	15	5.8	5.5	6.9	6.9	2.3†	5	II
400.	Lenôtre	1220		33	6.5	-	15	6.0	5.5	4.8	3.5†	3.5†	10	II
401.	Lecointre	1350		24	8.5	-	10	5.9	5.5	3.2	3.2	6.7†	30	II
402.	Mitchinamcus	1240		35	6.5	-	10	6.0	5.7	4.2	3.5	4.2	10	II
403.	Transparent	1080		36	5.0	-	10	6.1	5.5	11.0	11.0	0.5	3	I
404.	Byrd	1200		34	8.0	-	12	6.0	5.5	5.1	5.1	3.5†	10	II
405.	Dumoine	850		50	5.5	-	10	6.1	5.5	3.9	3.9	5.2†	10	II
406.	Rabbit	1070		18	12.0	-	10	5.5	5.0	3.3	3.3	4.0	10	II
407.	Nodier	1030		30	6.0	-	15	5.8	5.5	2.5	2.5	5.5	20	II
408.	Kenogemissi	1015	1600	26	10.0	20	40	7.2	6.0	3.0	3.0	7.2†	30	II
409.	Muskasenda	1050	624	42	5.0	-	30	7.0	6.0	3.2	3.2	6.0†	20	I
410.	Watabeeg	1080	2140	32	8.0	-	88	8.0	6.7	5.7	5.7	3.3†	5	II
411.	Mattagami	1080		50	5.7	-	35	7.5	6.9	2.7	2.7	7.0†	30	I
412.	Duncan	1100	1118	30	6.0	-	30	7.1	6.1	3.3	3.3	4.6†	20	II
413.	Longpoint	1030	372	20	7.0	13	35	7.3	6.1	3.6	3.6	5.8†	20	II
414.	Isabel	1375	197	33	5.0	-	24	6.7	5.7	9.0	9.0	1.2†	5	I
415.	Banks-Makoba	1190	3200	27	7.5	-	10	5.5	5.0	10.5	10.5	1.3†	3	II
416.	Gowganda	1110	1033	32	7.0	-	25	6.7	5.8	4.0	4.0	5.6†	20	II
417.	West Shining	1250		20	6.5	-	25	6.5	5.5	3.3	3.3	6.4†	30	II
418.	Onaping	1350		35	7.0	-	15	6.7	5.8	5.4	5.4	3.4†	10	II
419.	Smoothwater	1220	1300	85	4.0	-	15	5.9	5.8	16.2	16.2	0.1†	0	I
420.	Florence	1250		24	6.0	-	15	5.0	4.8	8.4	8.4	1.0†	5	II

No.	Locality	Elev.	A _s	Z _s	T _s	zO ₂	TH	E	pH	H	SDV	O ₂ c	Co.	Order
421.	Yorstan	1025		32	7.0	-	20	5.6	5.4	5.4	10.2	3.2†	10	II
422.	Emerald	1040	560	35	5.0	-	30	6.5	6.4	8.5	9.0	1.7	5	I
423.	Eaglerock	1040	120	30	5.5	-	28	6.4	6.1	9.0	2.6	5	5	I
424.	Gull	1055	1297	50	5.3	-	26	6.3	6.3	7.8	1.8	2.6	5	I
425.	Turtlesell	1030	150	33	5.5	-	20	6.3	5.8	7.5	2.4	2.4	5	I
426.	Turner	1070	50	24	7.0	-	12	5.5	5.0	9.0	2.3†	5	5	II
427.	Animal-Nipiasing	1070	1416	69	5.0	-	25	6.3	5.8	10.5	1.0	1.0	5	I
428.	Friday	1171	320	20	8.5	-	30	7.1	5.6	5.1	3.4†	5	5	II
429.	Lowell	1050	89	45	4.8	-	35	6.8	6.1	7.0	4.2	4.2	5	I
430.	Upper Twin	1050	178	45	4.8	-	35	7.1	6.6	6.3	3.5†	5	5	I
431.	Spring	1040	69	58	5.0	-	20	7.0	5.8	10.2	0.5†	3	3	I
432.	McConnell	1035	208	35	6.5	-	20	7.0	5.9	9.3	1.0†	5	5	II
433.	Gordon	913		25	5.2	-	20	5.8	5.4	5.1	3.5†	10	10	I
434.	White	895		42	5.8	-	15	6.4	5.5	6.5	3.4	3.4	10	II
435.	aux Sables	1010		11	11.5	-	14	6.5		2.7	9.0†	50	50	III
436.	Soufflot	1040		30	6.5	-	8	5.5	5.3	4.8	2.1	5	5	II
437.	la Truite	1122		33	7.4	-	12	5.5	5.5	6.0	1.1	5	5	II
438.	Lescoi	1042		35	5.5	-	12	5.7	5.5	4.8	3.9	5	5	I
439.	Sandeau	1015		27	5.6	-	15	5.7	5.0	4.5	3.5†	5	5	I
440.	Ogascan	1030		30	7.8	-	15	5.5	5.5	4.5	3.6	10	10	II
441.	des Loups	995		30	8.5	-	12	5.7	5.5	3.6	5.3	10	10	II
442.	Sasaginaga	1020		26	7.8	-	12	5.6	5.5	6.5	1.2	5	5	II
443.	Regenzie	945		33	7.0	-	15	6.1	5.6	5.5	2.3†	5	5	II
444.	Kikwesi	960		36	7.8	-	15	5.5	5.5	6.7	1.5	5	5	II
445.	Pants	890		24	8.9	-	12	5.8	5.3	4.2	2.7	5	5	II
446.	Booth	895		30	8.0	-	17	6.3	6.1	8.1	3.3	5	5	II
447.	Smith	890		75	4.5	-	15	6.6	6.0	9.0	1.0†	5	5	I
448.	Windy	930		30	5.1	-	15	6.5	5.5	8.1	1.4†	5	5	I
449.	Douglas	870		20	5.0	15	26	7.0	6.0	5.5	4.5†	10	10	I
450.	Petit Beauchêne	855		42	5.2	-	20	6.9	5.7	5.7	2.5	5	5	I
451.	Sairs	890		39	6.5	-	12	6.3	5.6	4.2	5.4†	20	20	II
452.	Sand	1080		25	8.0	-	17	6.5	5.6	4.2	4.0†	10	10	II
453.	Bleu	1075		57	6.5	-	15	6.1	6.0	7.5	1.0†	5	5	II
454.	Caughnawana	1075		24	7.0	-	15	6.1	6.0	7.8	1.0†	5	5	II
455.	Loon	1090		30	5.5	-	15	6.0	5.5	6.9	2.3†	5	5	I

Appendix 1

No.	Locality	Elev.	A _c	Z _c	T _m	zO ₂	TH	E	pH	SDV	O ₂ c	Co.	Order
456.	Memewith	840		36	7.0	-	15	6.5	5.5	5.1	2.8†	5	II
457.	Timber	1035	150	27	6.0	20	16	6.0	5.5	5.5	1.9†	5	II
458.	Transparent	975	93	36	4.4	-	18	6.5	5.8	8.7	1.5	5	I
459.	McCracken	980		40	6.0	-	15	6.0	5.5	6.0	2.8†	5	II
460.	Smyth	1325	46	23	5.0	20	20	6.2	5.5	5.0	4.2†	10	I
461.	Sweny	1220		35	5.5	-	26	6.5	5.8	6.9	1.5†	5	I
462.	Proudfoot ‡	1180	125	28	5.0	-	25	6.4	5.8	6.5	1.8†	5	I
463.	Butt ‡	1475											
464.	Burntroot ‡	1304		30	4.0	-	20	7.0	6.0	6.6	1.0†	5	I
465.	McCraney ‡	1431											
466.	Smoke ‡	1380		45	6.0	-	10	6.5	6.5	6.6	3.0†	5	II
467.	Canoe ‡	1379		26	6.0	-	10	7.2	6.6	4.8	3.5†	5	II
468.	Big Trout ‡	1322		50	6.0	-	12	6.5	6.0	4.0	3.5†	5	II
469.	Opeongo	1323	6000	45	7.0	-	15	6.5	6.0	3.6	3.0	10	II
470.	Two Rivers ‡	1289		33	5.0	-	20	6.3	5.8	3.2	6.0†	10	I
471.	Louisa ‡	1445											
472.	Bella	1160	337	35	6.5	-	25	6.8	5.5	3.3	3.5†	5	II
473.	Camp ‡	1370		45	5.0	-	25	6.7	5.6	3.8	3.0†	5	I
474.	Harp	1090	71	33	5.5	-	24	6.7	5.5	6.0	2.5†	5	I
475.	Oxtongue	1195	246	27	6.5	-	15	6.1	5.8	2.2	7.8†	30	II
476.	Livingstone ‡	1160		33	6.5	-	10	6.0	5.7	4.6	4.5†	10	II
477.	Kawagana	1165		70	5.0	-	13	6.5	6.5	8.4	1.5†	5	I
478.	Kennisia ‡	1212	1344	72	4.5	-	15	6.7	6.0	9.7	1.0†	5	I
479.	Redstone ‡	1195	1179	80	4.0	-	20	6.8	5.8	8.6	1.0†	5	I
480.	Little Hawk	1180	256	80	5.0	-	25	6.2	6.0	7.2	1.4†	5	I
481.	Raven ‡	1148		45	5.0	-	25	6.4	6.1	5.8	1.7†	5	I
482.	St. Nora	1092	841	35	6.5	-	26	6.6	5.6	6.6	1.8†	5	II
483.	White Pine	1100	76	22	6.0	-	25	6.5	5.5	4.5	4.0†	10	II
484.	Clear	1084	99	25	6.0	-	17	6.5	5.8	6.3	2.3†	5	II
485.	Kashagawigamog	1043	820	38	5.5	-	42	7.2	6.6	5.7	1.3†	5	I
486.	Koshlong	1137	396	35	5.5	-	15	6.4	5.5	6.3	2.3†	5	I
487.	Drag ‡	1161	900	55	4.5	-	15	6.8	5.5	8.0	2.0†	5	I
488.	Haliburton ‡	1183	998	52	4.5	-	30	7.0	6.0	5.0	3.0†	5	I
489.	Baptiste	1157	1952	31	8.0	-							
490.	Paudash ‡	1120	640	33	4.5	-	70	7.5	6.3	4.8	4.3†	10	I

No.	Locality	Elev.	A.	Z.	T _a	zO ₂	TH	E	pH	H	SDV	O ₂ C	Co.	Order
491.	Eels	1180	884	30	5.5	-	35	6.7	5.7	3.3	5.8†	20	I	
492.	Salmon	1030	161	32	5.5	20	90	8.2	6.7	8.1	1.2†	5	I	
493.	Chandos	1054	1360	42	5.5	-	55	7.3	6.3	4.5	2.5†	5	I	
494.	Jack	903	608	40	5.0	-	70	8.5	6.9	3.9	3.4†	10	I	
495.	Oak	853	650	24	7.0	12	60	8.0	6.6	3.6	3.5†	10	II	
496.	Dickey	1014	96	55	4.5	-	76	8.2	6.8	5.0	6.0	20	I	
497.	Wollaston	1050	348	30	4.9	-	80	8.5	7.0	3.5	3.0†	5	I	
498.	Limerick	1030	690	28	6.3	-	115	8.5	7.2	4.5	3.7	5	II	
499.	Ashby	1150	242	27	5.5	-	30	7.1	6.0	5.7	4.0	10	I	
500.	Aurden	1120	134	25	5.0	20	100	8.5	6.8	7.5	2.4	5	I	
501.	Buckshot	956	434	33	6.0	-	35	6.9	6.2	5.7	4.1	10	II	
502.	Mosque	1025	136	33	5.0	20	50	7.6	6.5	5.5	4.3†	10	I	
503.	Skootamatta	955	1271	30	6.5	-	25	6.8	5.7	4.0	6.0	20	II	
504.	Galeelny ‡	1278		24	5.0	-	30	6.8	5.9	4.2	6.0†	10	I	
505.	Louisa ‡	1300		60	4.0	-	34	8.0	6.5	7.5	1.0†	5	I	
506.	Papineau ‡	1039		60	5.0	-	20	6.5	6.0	7.0	1.0†	5	I	
507.	Clear ‡	755	2399	40	5.0	-	26	7.0	6.5	7.5	2.0†	5	I	
508.	Bark ‡	1027	6000	72	5.0	-	26	7.0	6.5	4.2	5.0†	10	II	
509.	Kameniskieg	928	2172	42	5.5	-	34	7.5	7.0	4.2	4.5†	10	I	
510.	Paugh	990	740	50	4.5	-	20	6.8	6.0	5.0	4.0†	10	I	
511.	Victoria	1270		45	4.5	-	25	7.0	6.5	7.5	2.0†	5	I	
512.	Aylen ‡	1170		70	5.5	-	29	7.0	6.0	4.2	5.0†	10	I	
513.	Lavielle ‡	1266		45	4.5	-	20	6.8	5.0	5.7	1.0†	5	II	
514.	Loat	1150		37	6.0	-	20	7.0	6.2	6.0	2.1†	5	II	
515.	Stevens	901		36	6.0	-	20	7.0	6.2	6.0	2.1†	5	II	
516.	Vert	895		30	6.0	25	20	6.8	6.0	7.2	1.0†	5	II	
517.	St. Patrick	870		60	6.5	-	10	6.5	6.0	9.5	1.2†	5	II	
518.	Lynch	890		75	6.5	-	12	6.6	6.0	9.0	1.0†	5	II	
519.	aux Verc	880		65	4.5	-	8	6.2	6.0	12.9	0.5†	3	I	
520.	Pappin	760		33	5.0	-	10	6.5	5.5	4.8	3.0†	10	II	
521.	Trout	750		28	6.0	-	20	6.5	6.0	5.7	2.8†	5	II	
522.	Jim	681		24	7.5	-	28	7.5	6.2	4.0	5.9†	30	II	
523.	Vert	550		20	4.5	15	50	7.6	6.4	6.0	2.0†	5	I	
524.	Bark	1183		37	6.0	-	12	6.4	5.7	5.0	4.5†	10	II	
525.	Roland	1225		20	8.0	15	16	6.3	6.0	4.6	5.0†	20	II	

No.	Locality	Elev.	A.	Z.	T.	xO ₂	TH	E	pH	H	SDV	O ₂	Co.	Order
526.	Embarrass	1157		45	4.1	-	8	6.2	5.5		4.5	5.3†	10	I
527.	du Rocher	1085		55	4.8	-	15	6.3	5.5		6.9	2.0†	5	I
528.	Buckshot	1020		37	4.2	-	12	6.5	5.5		4.9	4.0	5	I
529.	Ross	810		18	6.2	15	18	6.9	5.6		6.2	1.3†	5	II
530.	Corbeau	900		36	5.4	-	8	6.5	5.5		4.8	4.0†	10	I
531.	Gilmore	850		24	5.0	-	15	6.6	5.5		4.8	4.0†	15	I
532.	Gagamo	890		22	7.5	-	10	5.6	5.0		3.0	5.6†	20	II
533.	la Croche	852		24	6.0	-	15	5.7	5.4		4.3	5.2†	15	II
534.	Cole	800		22	6.0	12	12	6.0	5.5		3.6	6.7†	30	II
535.	Royal	535		27	6.0	-	15	6.0	5.5		5.0	3.4†	5	II
536.	Caroline	1025		36	5.3	-	12	5.6	5.0		6.0	1.5†	5	I
537.	Clearwater	780		25	4.5	-	16	6.4	5.6		6.0	2.3†	5	I
538.	Green	825		30	4.5	-	15	6.5	5.5		5.4	3.2†	5	I
539.	à la Tortue	730		20	7.0	15	20	6.7	5.5		5.1	4.5†	20	II
540.	Pythonga	686		78	5.6	-	15	6.7	6.0		6.3	4.9	10	I
541.	David	716		33	6.0	-	16	6.6	5.5		8.4	2.0†	5	II
542.	Abatis	899		27	6.4	-	16	6.6	5.8		4.5	1.9	10	II
543.	Urborne	796		26	4.3	-	20	6.1	6.0		3.5	6.9	10	I
544.	Conway	725		32	5.0	-	20	6.5	5.6		4.5	2.3†	5	I
545.	Mer Bleue	680		33	7.2	-	35	7.0	6.0		4.8	3.0†	10	II
546.	Latourrelle	640		18	5.5	15	70	8.5	6.5		5.4	6.0†	5	I
547.	des Îles	630		21	5.5	18	60	8.5	6.6		6.3	6.7	5	I
548.	Patterson	800		25	6.5	-	15	6.8	5.5		5.4	2.4	5	II
549.	Cayamant	640	680	57	7.0	-	35	7.2	6.5		4.8	3.0	10	II
550.	Hook	890		42	4.0	-	25	7.1	6.0		10.5	0.5†	3	I
551.	Lamers	640		32	4.2	15	17	7.0	5.5		7.5	2.3†	5	I
552.	Petit Cayamant	610		48	5.8	-	35	7.0	6.5		5.7	4.5†	20	II
553.	Laforêt	825		18	4.3	15	27	6.7	6.5		7.0	2.3†	5	I
554.	Dumont	851		55	4.3	-	27	6.6	6.5		10.5	1.2	5	I
555.	Squaw	803		42	4.0	-	35	6.8	6.5		6.6	3.2	5	I
556.	Hickey	738		55	4.0	-	30	7.5	6.6		6.3	2.5†	5	I
557.	Huddersfield	770		22	4.5	-	27	6.4	6.0		3.9	4.0†	5	I
558.	Lealie	658		33	4.2	-	25	7.4	6.7		3.4	4.5†	10	I
559.	MacCraig	647	80	30	4.2	18	35	7.6	6.6		6.0	1.5†	5	I
560.	Hughes	646		20	6.5	10	25	6.9	6.0		4.5	5.2†	20	II

No.	Locality	Elev.	A _s	Z _s	T _s	ZO ₂	TH	E	pH	H	SDV	O ₂ c	Co.	Order
561.	Otter	638		22	12.3	15	32	7.0	6.0	6.0	4.4	5.3†	20	II
562.	Johnson	645		34	4.0	18	55	7.6	6.3	6.3	5.1	2.3†	5	I
563.	Mechan	647		42	4.0	12	112	8.6	7.2	4.3	4.3	5.0†	5	I
564.	Charlette	1200		18	5.0	-	20	6.5	6.0	2.4	7.0†	3.0	30	I
565.	Harrington	560		21	7.0	10	45	7.0	6.6	6.3	6.3	3.0	5	II
566.	Sinclair	784		36	5.0	25	75	8.4	6.7	4.2	4.2	4.8	5	I
567.	Isabel	797		33	5.6	-	60	7.8	6.8	9.0	3.5†	5	5	I
568.	Cabaret	799		25	6.5	-	60	7.0	6.7	2.4	7.5†	30	30	II
569.	Bernard	564		20	6.2	13	76	8.1	6.7	5.4	3.5†	10	10	II
570.	Meach	555		23	6.0	14	46	7.0	6.6	5.4	4.0	5	5	II
571.	Little Trout	895		33	4.5	25	30	7.2	5.5	8.7	1.0†	5	5	I
572.	à la Truite	990		80	4.0	-	18	6.0	5.5	8.4	2.3†	5	5	I
573.	Sam	880		60	5.0	-	20	6.2	5.5	7.5	1.8†	5	5	I
574.	Normandeau	1150		40	5.0	-	15	6.4	5.5	6.9	1.8†	5	5	I
575.	Écluse	890	120	42	5.5	-	25	6.8	5.5	6.6	2.3†	5	5	I
576.	Wakefield	539	352	54	4.5	40	52	7.9	6.0	4.8	4.0†	5	5	I
577.	Grand	516	530	40	7.5	-	60	7.2	6.0	8.1	3.0†	5	5	II
578.	Long	790		33	5.0	12	45	7.0	5.5	6.3	4.5†	5	5	I
579.	Clay	667	440	40	7.5	-	50	7.5	7.0	2.0	6.3†	10	10	II
580.	Vert	825		42	4.5	24	55	6.7	6.0	8.4	1.5†	5	5	I
581.	Lethbury	990		35	4.5	-	33	6.3	5.5	6.9	3.0†	5	5	I
582.	O'Hara	695		45	4.5	-	95	8.0	6.8	6.9	1.5†	5	5	I
583.	Babiche	1015		32	6.0	-	25	6.8	5.5	5.5	4.5†	10	10	II
584.	Perras	980		53	5.0	-	25	6.8	5.8	7.5	2.0†	5	5	I
585.	Pimodon	1050		32	6.0	-	20	6.5	5.8	6.0	3.0†	5	5	II
586.	Quinn	740		40	5.0	-	27	6.9	6.0	6.9	2.5†	5	5	I
587.	Winding	990		36	6.0	-	12	6.3	5.5	5.7	3.0†	5	5	II
588.	Kiamuka	682		43	6.0	-	26	7.0	6.5	4.2	5.5†	10	10	II
589.	des Cornes	960		33	5.5	-	20	6.4	5.6	6.0	4.5†	5	5	I
590.	Major	925		36	5.4	-	17	6.7	6.0	4.6	5.3	10	10	I
591.	Laluge	1020		28	5.5	-	15	6.8	5.9	4.5	5.5†	10	10	I
592.	Iveteaux	915		27	5.0	-	12	6.4	5.4	3.3	3.6	5	5	I
593.	Dieppe	1240		30	5.5	-	15	6.0	5.5	3.6	6.3†	20	20	I
594.	Bondy	960		105	4.0	-	10	5.9	5.7	8.5	1.0†	5	5	I
595.	Douaire	1125		32	6.5	-	8	5.8	5.6	5.3	1.2†	5	5	II

No.	Locality	Elev.	A _s	z _m	T _m	zO ₂	TH	E	pH	SDV	O ₂ C	Co.	Order
596.	Nominique	812		35	4.5	-	26	6.6	6.6	6.3	4.0	5	I
597.	Chaud	850	640	33	5.4	-	26	5.9	5.9	4.2	5.6	10	I
598.	Petit Nominique	813		30	6.0	15	20	6.3	6.0	3.3	3.8	10	II
599.	Montjoie	875	1203	75	4.5	-	15	6.4	5.8	6.4	1.5†	5	I
600.	Labelle	812		60	4.0	-	26	6.6	6.4	5.4	3.7	10	I
601.	Tremblant	747		62	4.5	-	12	6.0	6.0	5.5	3.9	5	I
602.	Simon	652	3200	105	6.5	-	30	6.6	6.0	6.7	1.0†	5	I
603.	Désormeaux	722	269	45	7.0	-	35	7.2	6.0	4.8	3.5†	5	II
604.	Viceroi	694		27	8.0	-	35	7.4	6.0	6.3	2.4†	5	II
605.	Cross	690		30	4.5	-	30	5.7	5.5	4.8	4.5†	10	I
606.	Beavin	593	358	27	5.6	22	35	6.8	6.0	4.2	4.2	10	I
607.	Sixteen Island	930	358	54	5.0	-	25	6.6	6.0	5.7	3.0†	5	I
608.	Anne	1020	113	25	5.5	21	16	6.7	5.8	4.5	4.2†	10	I
609.	Bark	711	645	48	4.0	30	15	6.3	5.2	4.5	2.6	5	I
610.	Saint-Joseph	1199	143	32	6.0	-	17	6.6	5.5	2.8	6.5†	20	II
611.	Manitou	1273	394	23	5.5	-	18	6.7	5.6	3.9	5.0†	10	I
612.	Masson	1100		45	4.5	-	18	6.7	5.6	3.3	6.0†	20	I
613.	Westford	1192		24	5.5	-	19	6.8	5.9	4.2	4.5†	10	I
614.	Archambault	1261		40	4.5	-	12	5.9	5.9	5.7	3.0	5	I
615.	Chertsey	990	152	33	5.0	-	16	6.6	5.9	5.1	3.8†	5	I
616.	Cloutier	730		24	5.5	-	25	7.0	5.9	4.5	4.5†	10	I
617.	des Français	550		24	5.0	20	25	6.9	5.7	3.5	5.0†	10	I
618.	Maskinonge	467		27	7.0	-	15	6.1	5.5	3.3	4.3	10	II
619.	Long	560		30	4.5	-	20	6.7	5.5	4.5	4.0†	5	I
620.	Goulet	519		30	4.2	12	16	6.6	5.5	3.3	4.5†	5	I
621.	la Pêche	555	320	40	4.1	-	27	6.6	5.5	3.6	4.0	5	I
622.	Isle	540		24	6.5	-	15	6.5	6.0	5.0	4.0†	5	II
623.	Édouard	913	174	30	6.0	-	8	5.8	5.0	5.0	4.9	10	II
624.	Sacconie	914		65	5.5	-	10	6.3	5.5	14.5	0.5†	3	I
625.	à l'Eau Claire	815		72	5.5	-	12	6.5	5.5	6.9	2.0†	5	I
626.	Wapazigouke	725	564	27	6.0	-	10	6.4	5.7	6.6	1.8	5	II
627.	Dauphinais	825	200	30	5.0	-	10	6.1	5.0	4.5	2.0	5	I
628.	Atikagami	560	164	28	4.3	-	15	6.4	5.5	3.0	4.4	5	I
629.	Caribou	1300	387	35	5.7	-	10	6.3	5.3	9.3	2.0	5	I
630.	Taureau	1190		25	6.0	-	15	5.9	5.7	2.7	8.1†	30	II

No.	Locality	Elev.	A.	Z _m	T _m	zO ₂	TH	E	pH	H	SDV	O ₂ c	Co.	Order
631.	Clear	1350		50	5.0	—	15	6.6	6.6	6.3	6.6	4.0†	10	I
632.	Normand	1350	920	60	5.0	—	10	6.3	6.3	6.0	4.2	5.5†	10	I
633.	Kempt	1350		48	6.0	—	10	6.0	6.0	5.8	4.8	5.0†	10	II
634.	Harper	801		68	4.5	—	12	6.4	6.0	6.0	4.5	4.5†	5	I
635.	Wayagamac	887		35	5.0	—	10	6.2	6.2	6.0	6.3	3.5†	5	I
636.	à Beauce	705		26	6.0	—	25	6.4	6.3	6.3	4.5	5.0†	10	II
637.	Mekinac	525	2341	125	4.0	—	10	6.2	6.2	6.2	4.1	4.5†	5	I
638.	du Missionnaire	520		42	6.5	—	18	6.3	6.3	6.1	6.0	4.0	5	II
639.	du Jésuite	670		33	6.0	—	15	6.3	6.3	5.8	4.8	3.0†	5	II
640.	aux Sables	520		35	6.0	—	12	5.8	5.5	5.5	7.5	2.5	5	II
641.	Blanc	480		45	5.0	—	15	5.5	5.5	5.5	3.0	6.0	20	I
642.	Saint-Joseph	520		35	4.0	—	15	5.5	5.5	5.5	4.5	5.0†	20	I
643.	Jacques-Cartier	2500		100	4.9	—	8	5.6	5.6	5.4	3.9	6.3†	20	I
644.	des Commissaires	900		25	9.5	—	9	6.2	5.8	5.8	5.5	5.0†	20	II
645.	Kenogami	500		75	5.0	—	15	5.7	5.6	5.6	1.3	9.0†	50	I
646.	Jim	690	432	42	5.0	—	10	5.8	5.6	5.6	2.0	7.5†	30	I
647.	aux Rats	625		26	5.5	—	10	5.5	5.0	5.0	2.4	7.3†	30	I
648.	La Mothe	820		50	5.0	—	15	6.5	6.0	6.0	2.5	6.5†	20	I
649.	Labrecque	440		27	6.5	—	26	6.8	6.2	6.2	3.0	5.5†	10	II
650.	Matapedia	514		33	5.5	—	85	8.0	7.8	7.8	3.0	4.0†	5	I
651.	Humqui	700		37	4.2	—	85	8.0	7.4	7.4	4.2	2.8†	5	I
652.	Petit Macpes	500		33	5.8	—	85	8.4	7.6	7.6	4.5	3.0†	5	II
653.	Saint-Simon	390	572	20	9.9	—	42	7.2	7.1	7.1	2.7	5.0†	10	II
654.	Squatek	525		47	5.8	—	60	7.3	7.0	5.4	1.8†	1.8†	5	II
655.	Merumitcook	650		55	5.5	—	51	7.5	7.1	7.1	4.8	3.2	5	I
656.	Long	620	1664	22	8.6	20	36	7.4	7.0	7.0	3.6	5.5†	10	II
657.	Pohenegamook	650	856	35	4.0	—	38	6.5	6.4	6.4	2.4	4.0†	10	I
658.	de l'Est	1050		29	4.4	—	26	7.4	6.3	6.3	3.0	5.5†	20	I
659.	Long	620	2400	45	5.3	—	42	7.0	6.7	1.0	4.5†	1.5	15	I
660.	Eagle	574		40	4.0	—	35	6.7	6.5	6.5	2.4	6.0†	30	I
661.	St. Froid	581		30	4.5	—	25	6.6	6.4	6.4	2.7	6.0†	30	I
662.	Pushineer	1200		15	5.0	—	20	6.9	6.0	6.0	4.5	1.8†	5	I
663.	Grand	50	16240	30	7.0	—	30	6.8	6.0	6.0	2.4	5.5†	20	II
664.	Utopia	55		20	10.0	—	20	6.0	5.5	5.5	3.3	6.0†	30	II
665.	Tunk	200	804	60	5.0	—	30	7.0	6.0	6.0	10.8	1.0†	5	I

No.	Locality	Elev.	A _s	Z _m	T _m	zO ₂	TH	E	pH	SDV	O ₂ c	Co.	Order
666.	Green	150	1210	40	7.0	—	30	6.3	5.5	4.0	6.5†	20	II
667.	Chamberlain	945	4433	45	4.8	—	18	6.3	6.3	4.2	7.2	30	I
668.	Moosehead ‡	1050	33100	75	4.5	—	16	6.5	6.0	5.0	4.5†	10	I
669.	Meganic	1294	2650	45	4.0	—	30	6.5	6.3	3.6	3.0	15	I
670.	William	650		27	5.0	15	68	7.8	6.6	1.2	5.0†	5	I
671.	Elgin	880		33	4.0	—	36	6.8	6.5	3.6	5.0†	20	I
672.	Nicolet	1150		40	4.0	—	26	6.5	6.4	7.8	1.0†	5	I
673.	Montjoie	890		20	8.0	15	30	6.4	6.0	3.6	6.0†	30	II
674.	Bowker	1110		45	5.0	—	25	6.5	6.3	8.1	0.0	5	I
675.	Stukely	935		30	6.0	—	38	6.6	6.5	5.1	2.5	5	II
676.	Lyster	1550	170	45	5.9	—	37	7.0	6.7	5.1	2.1	5	II
677.	Big Averill	1684	339	30	5.5	—	15	6.1	5.8	6.3	4.5†	5	I
678.	Salem	963		21	7.0	15	50	8.0	7.2	3.3	6.5†	20	II
679.	Seymour	1279	702	50	5.0	—	40	7.2	7.0	6.9	3.5†	5	I
680.	Willoughby	1170	600	81	5.0	—	45	7.3	7.2	11.2	0.7†	3	I
681.	Crystal	970	272	31	5.0	—	38	7.2	6.9	3.3	5.0†	25	I
682.	Casplan	1400	315	42	5.0	—	70	8.0	7.1	6.9	2.0†	5	I
683.	Eligo	881	47	30	6.0	20	85	8.3	7.1	5.1	3.0†	5	II
684.	Rangley	1520	2400	30	7.0	—	10	6.8	6.0	9.0	1.0	5	II
685.	Thompson	500	1740	27	9.0	—	30	6.7	6.0	5.5	2.0†	5	II
686.	Sebago	250	11630	96	4.0	—	35	7.0	6.5	10.5	0.7†	3	I
687.	Winnepesaukee ‡	480	18000	54	5.0	—	20	6.5	6.0	6.5	2.0†	5	I
688.	Sunapee ‡	1200		60	4.5	—	15	6.5	6.0	8.0	1.0†	5	I
689.	Chazy	1531	794	25	6.0	—	24	6.8	6.0	4.2	5.0†	10	II
690.	Upper Chateaugay	1310		20	6.0	—	30	6.7	6.4	3.0	6.0†	20	II
691.	Taylor	1370		27	5.5	—	26	6.6	5.8	5.0	2.5†	5	I
692.	Upper St. Regis	1600		21	6.0	15	15	6.4	5.6	3.0	6.5†	20	II
693.	Placid	1850		42	5.0	—	20	5.8	5.7	10.2	1.0†	5	I
694.	Upper Saranac	1571	2035	27	5.5	—	20	6.8	5.9	3.3	7.0†	20	I
695.	Tupper	1542	1920	27	6.0	—	12	5.5	5.0	3.5	6.5†	15	II
696.	Trout	780		27	7.0	20	36	7.1	6.6	4.0	5.0†	10	I
697.	Massawepie	1550		24	5.0	15	15	6.5	5.5	5.7	4.0†	5	I
698.	Portaferry	859		27	5.5	20	32	6.9	3.6	3.6	6.0†	20	I
699.	Bonaparte	768	512	24	6.5	—	75	7.8	7.3	4.2	4.5†	10	II
700.	Brantingham	1235	138	18	6.5	15	15	6.2	6.0	3.6	7.0†	30	II

No.	Locality	Elev.	A _s	z _m	T _m	zO ₂	TH	E	pH	SDV	O ₂ c	Co.	Order
701.	Fourth Fulton	1707		21	6.0	-	20	6.5	6.3	4.8	5.5†	10	II
702.	White	1422	84	23	6.0	15	20	6.5	6.1	7.8	2.0†	5	II
703.	Schroon	702	1692	45	5.0	-	24	6.8	6.3	4.5	4.4†	10	I
704.	Trout	759	100	23	5.0	20	26	6.0	6.0	4.5	4.5†	10	I
705.	Canada	1542	210	35	5.0	-	12	5.4	5.0	5.1	4.0†	10	I
706.	Hedges	535	30	23	5.5	12	136	8.5	7.1	2.1	5.5†	5	I
707.	Otsego	1194	1600	50	5.0	-	145	8.5	7.9	4.8	3.5†	5	I
708.	Canadice	1012	256	27	6.5	25	60	7.2	6.9	2.0	4.0†	5	II
709.	Sylvan	325		42	5.5	-	144	8.5	7.9	3.6	3.5†	5	I
710.	Gilead	498		35	5.5	-	43	7.0	6.8	7.8	1.0†	5	I
711.	Glenida	508	76	30	6.0	20	80	8.0	7.2	9.0	1.0†	5	II
712.	Sterling	750	126	33	4.5	-	24	6.5	6.2	6.3	2.0†	5	I
713.	Wawayanda	525	90	33	6.0	-	40	7.0	6.6	2.4	5.6†	20	II
714.	Western Brook	125		210	9.2	-	9	7.0	6.7	9.0	3.6	5	II
715.	Rocky Harbour	50		20	11.0	-	70	7.6	7.6	2.7	9.8	20	II
716.	Deer	17		6144	6.0	-	10	6.5	6.0	2.7	9.0	30	I

* A dash signifies that at no depth was oxygen less than 2 ppm at time of survey.

† Estimated by graphical analysis (after Schindler 1971).

‡ Physicochemical data obtained from the literature or regional biologist's files.

Appendix 2

Appendix 2
Average number of *Pontoporeia "affinis"*
per Ekman grab in relation to depth and
water chemistry in nine selected lakes

Round Lake (No. 196), 16 Aug. 1971

Depth (m)	Temp. (°C)	O ₂ (ppm)	pH	<i>Pontoporeia</i>
0	21.0		7.4	0
10	13.0		6.4	12
15	7.5	8	6.3	
20	7.0		6.1	
25	7.0		6.1	
30	7.0		6.1	22
35	7.0		6.1	16
40	6.5		6.1	33
45	6.5		6.0	44
50	6.5	6	6.0	45

Total hardness: 30 ppm

O₂ consumed: 10 m, 5.8 ppm

50 m, 4.0 ppm

Number of Ekman grabs per depth level: 2

Blue Sea Lake (No. 275), 20 Aug. 1971

Depth (m)	Temp. (°C)	O ₂ (ppm)	pH	<i>Pontoporeia</i>
0	21.0		8.5	0
10	18.0		8.5	0
15	8.0	7	7.3	86
20	7.0		7.1	140
25	6.5		7.1	125
30	6.5		7.1	141
35	6.5		7.1	178
40	6.0		7.0	235
45	5.5		7.0	188
50	5.5	6	7.0	161

Total hardness: 90 ppm

O₂ consumed: 15 m, 1.9 ppm

50 m, 2.3 ppm

Number of Ekman grabs per depth level: 2

Canoe Lake (No. 182), 19 Aug. 1971

Depth (m)	Temp. (°C)	O ₂ (ppm)	pH	<i>Pontoporeia</i>
0	22.0		8.5	0
10	12.0		8.5	5
15	8.0	6	7.3	32
20	7.0		7.3	10
25	6.5		7.2	13
30	6.5		7.1	33
35	6.5		7.0	11
40	6.0		7.0	16
45	5.5	6	7.0	1

Total hardness: 90 ppm

O₂ consumed: 15 m, 2.8 ppm

45 m, 1.4 ppm

Number of Ekman grabs per depth level: 2

Bernard Lake (No. 122), 7 Aug. 1972

Depth (m)	Temp. (°C)	O ₂ (ppm)	pH	<i>Pontoporeia</i>
0	22.0		6.7	0
10	13.5		6.0	2.5
15	10.5	9	5.9	38
20	9.0		5.9	42.5
25	8.0		5.8	58.5
30	7.5		5.8	10.5
35	7.0		5.7	21
40	6.0		5.5	24.5
45	6.0		5.5	0
50	6.0	8	5.5	0

Total hardness: 25 ppm

O₂ consumed: 10 m, 2.0 ppm

Number of Ekman grabs per depth level: 2

Buck Lake (No. 140), 1 Aug. 1972

Depth (m)	Temp. (°C)	O ₂ (ppm)	pH	<i>Pontoporeia</i>
0	22.0		5.7	0
10	7.5	9	5.5	0
15	6.5		5.3	0
20	6.0		5.2	0
25	6.0		5.0	0
30	6.0		5.0	0
35	6.0	6	5.0	0

Total hardness: 25 ppm

O₂ consumed: 10 m, 9.2 ppm

Number of Ekman grabs per depth level: 3

Lac Albanel (No. 3), 10 July 1970

Depth (m)	Temp. (°C)	O ₂ (ppm)	pH	<i>Pontoporeia</i>
0	16.0		6.6	0
10	12.0		6.5	0
15	9.0	9	6.3	0
20	8.5		6.3	0
25	8.0		6.3	0
30	8.0		6.3	0
40	8.0	9	6.3	0

Total hardness: 17 ppm

O₂ consumed: 10 m, 5.3 ppm

Number of Ekman grabs per depth level: 2

Appendices

Charleston Lake (No. 190), 18 Aug. 1971

Depth (m)	Temp. (°C)	O ₂ (ppm)	pH	<i>Pontoporeia</i>
0	22.0		8.5	0
10	12.0		7.5	120
20	6.5		7.2	232
30	5.5	6	7.2	263
40	5.0		7.2	231
50	5.0		7.2	96
60	5.0		7.2	134
70	4.5		7.2	147
80	4.5		7.1	145
90	4.0	6	7.0	111

Total hardness: 130 ppm

O₂ consumed: 20 m, 4.3 ppm

90 m, 3.5 ppm

Number of Ekman grabs per depth level: 2

Mazinaw Lake (No. 167), 17 Aug. 1971

Depth (m)	Temp. (°C)	O ₂ (ppm)	pH	<i>Pontoporeia</i>
0	21.5		7.3	0
10	13.0	8	6.6	14
20	6.5		6.6	21
30	5.5		6.6	20
40	5.0		6.6	25
50	5.0		6.6	23
60	4.5		6.6	15
70	4.5		6.6	14
80	4.5		6.6	6
90	4.0		6.6	5
100	4.0	6	6.6	4

Total hardness: 36 ppm

O₂ consumed: 10 m, 5.3 ppm

100 m, 5.9 ppm

Number of Ekman grabs per depth level: 2

Fairy Lake (No. 143), 14 July 1972

Depth (m)	Temp. (°C)	O ₂ (ppm)	pH	<i>Pontoporeia</i>
0	21.5		6.3	0
10	8.0	11	5.6	0
20	5.5		5.6	0
30	5.0		5.6	0
40	5.0		5.5	0
50	4.5		5.4	0
60	4.5		5.3	0
70	4.5	7	5.3	0

Total hardness: 20 ppm

O₂ consumed: 0 m, 5.9 ppm

Number of Ekman grabs per depth level: 3

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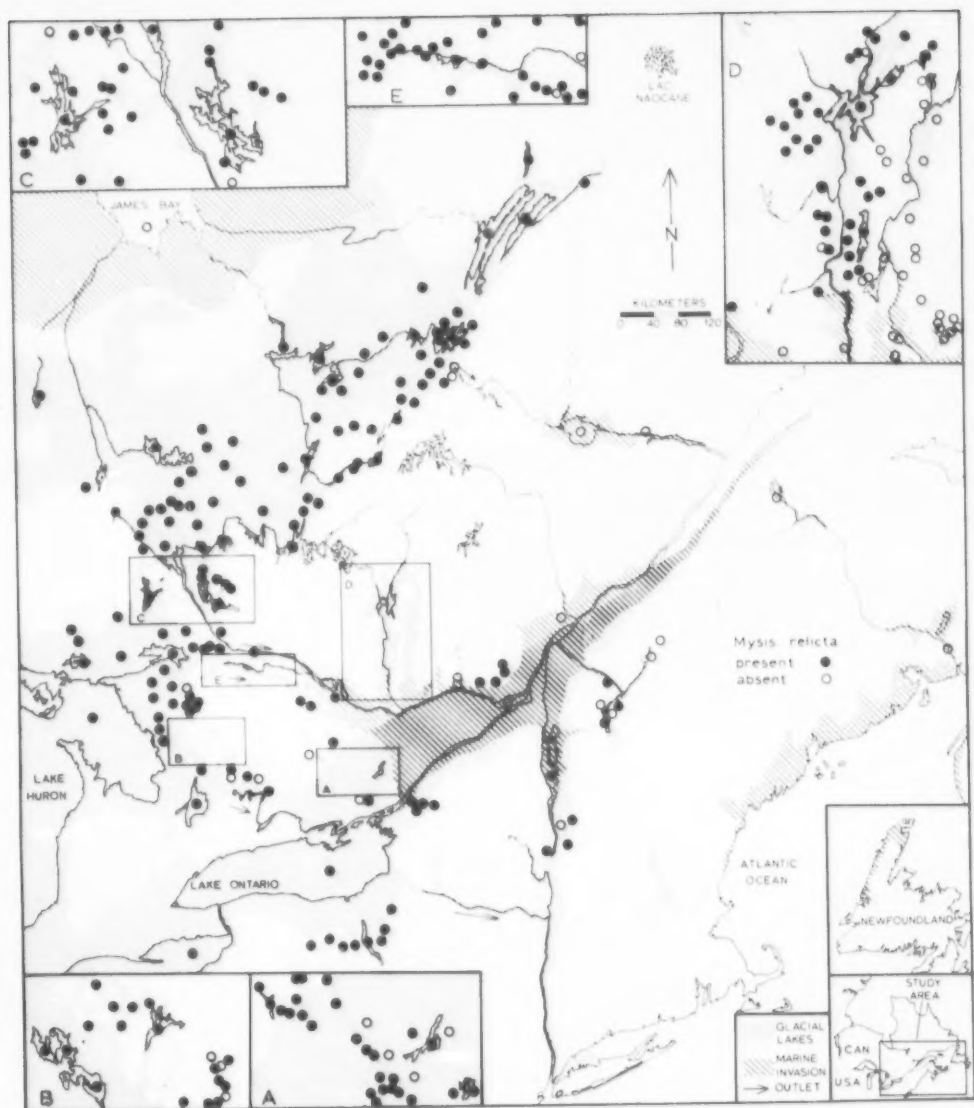
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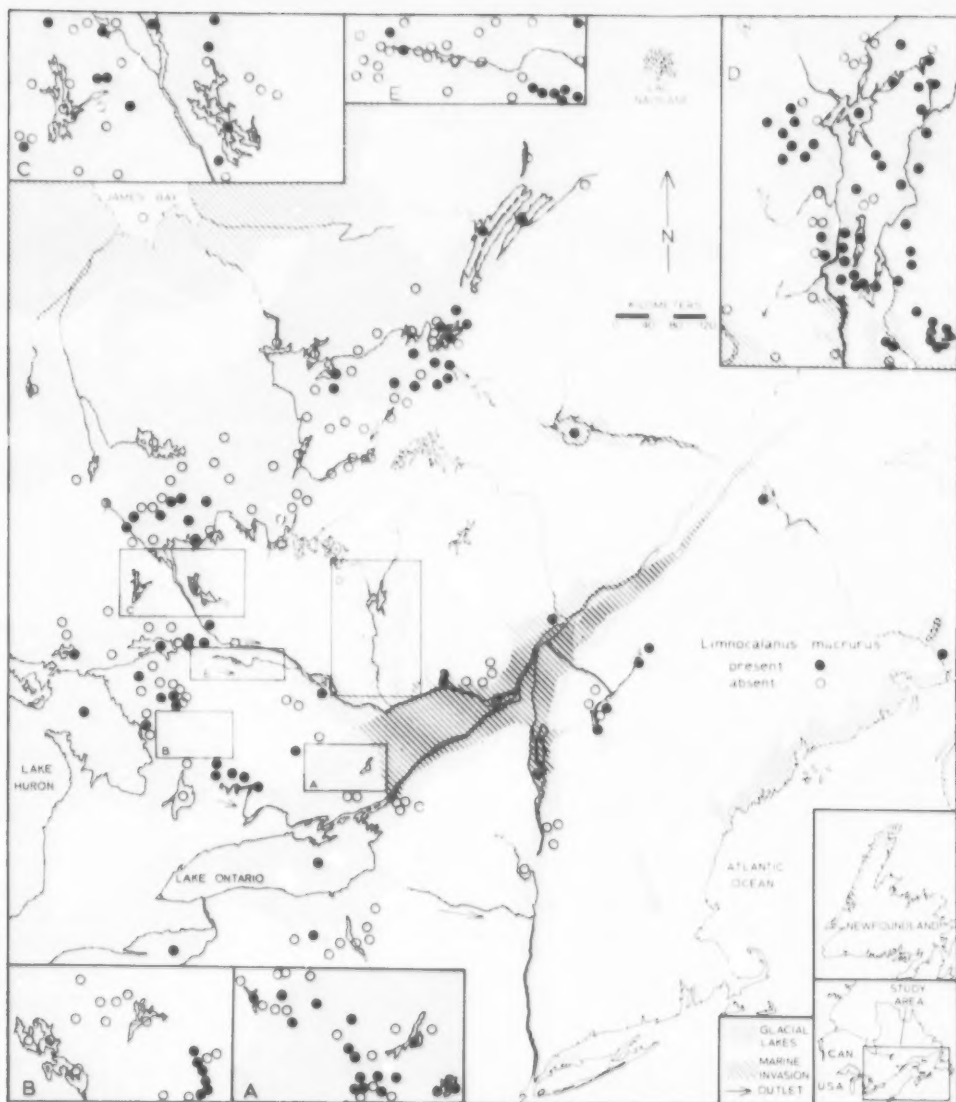
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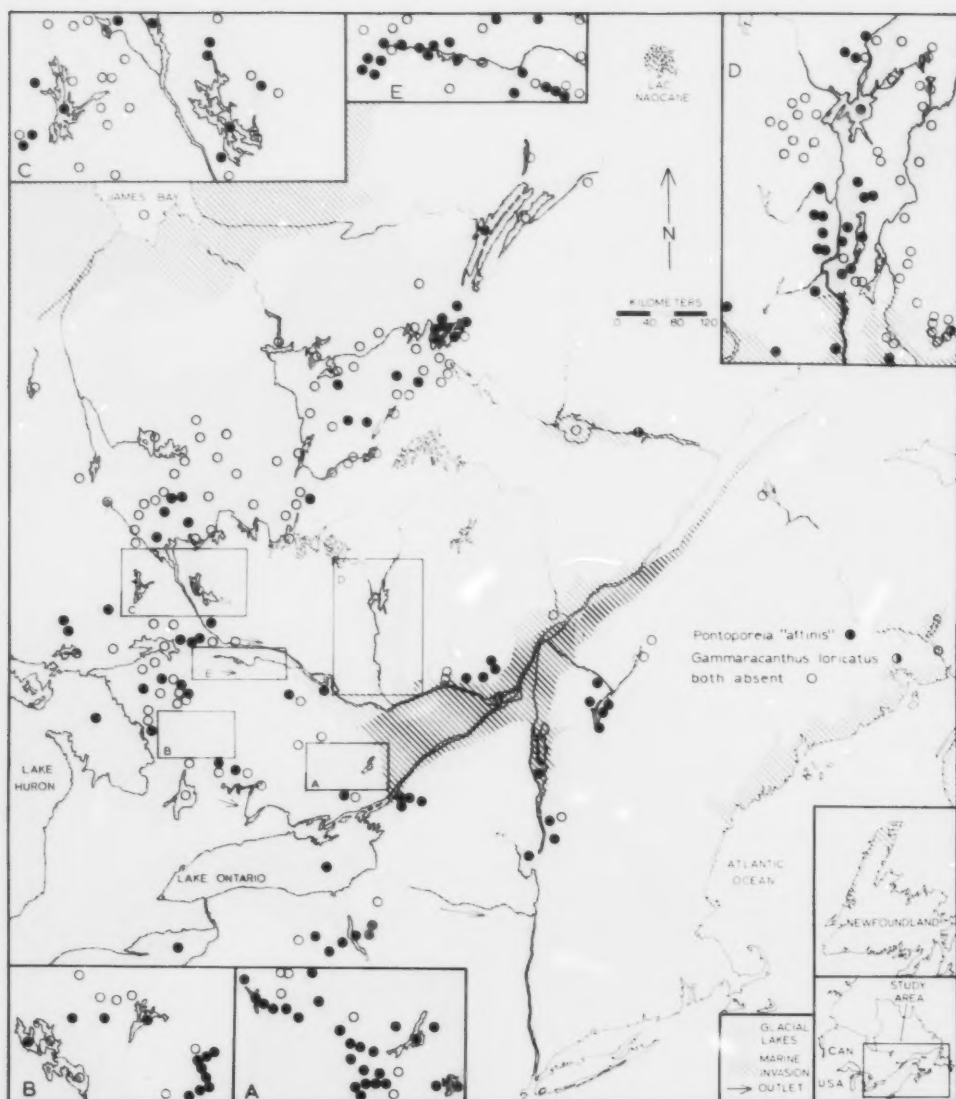
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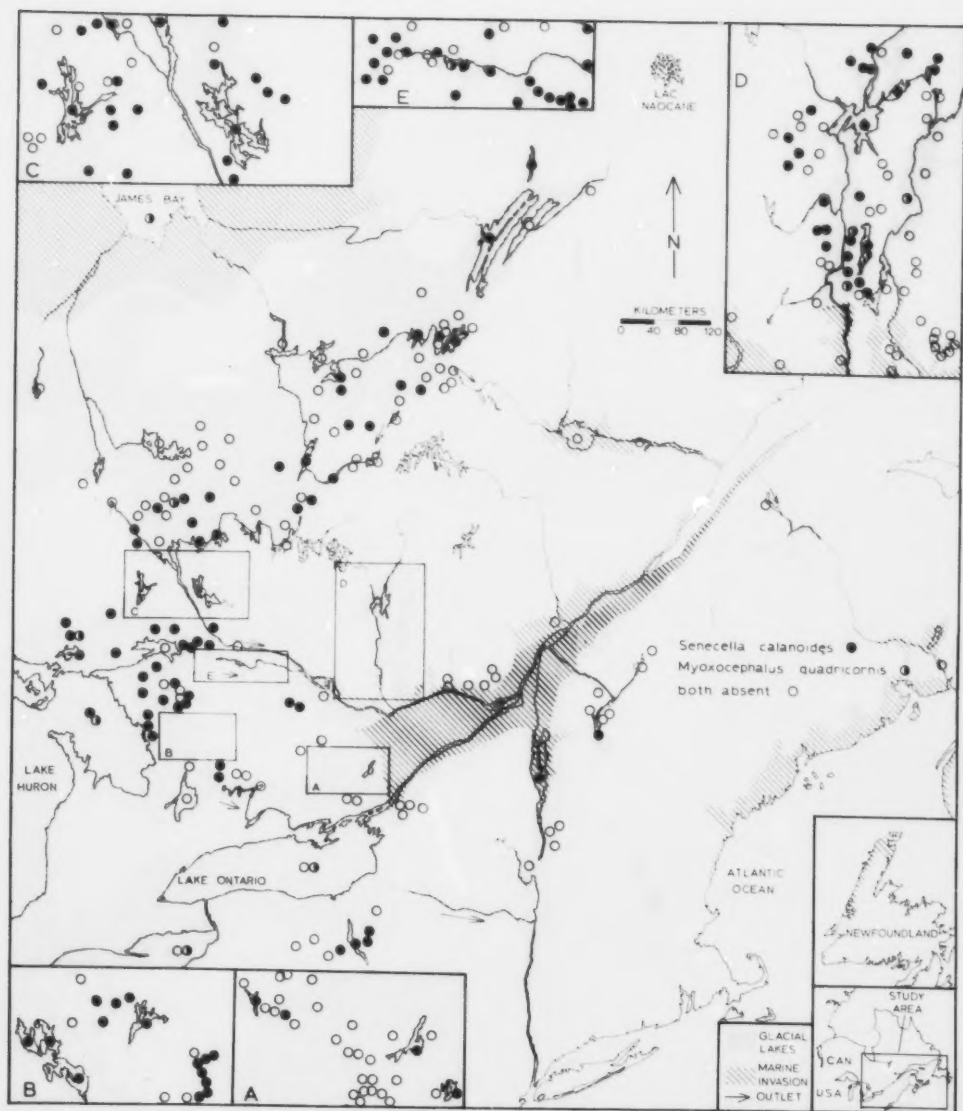
Map 4A
Distribution of *Mysis relicta* in eastern North America. Dots indicate species present; circles indicate species absent.



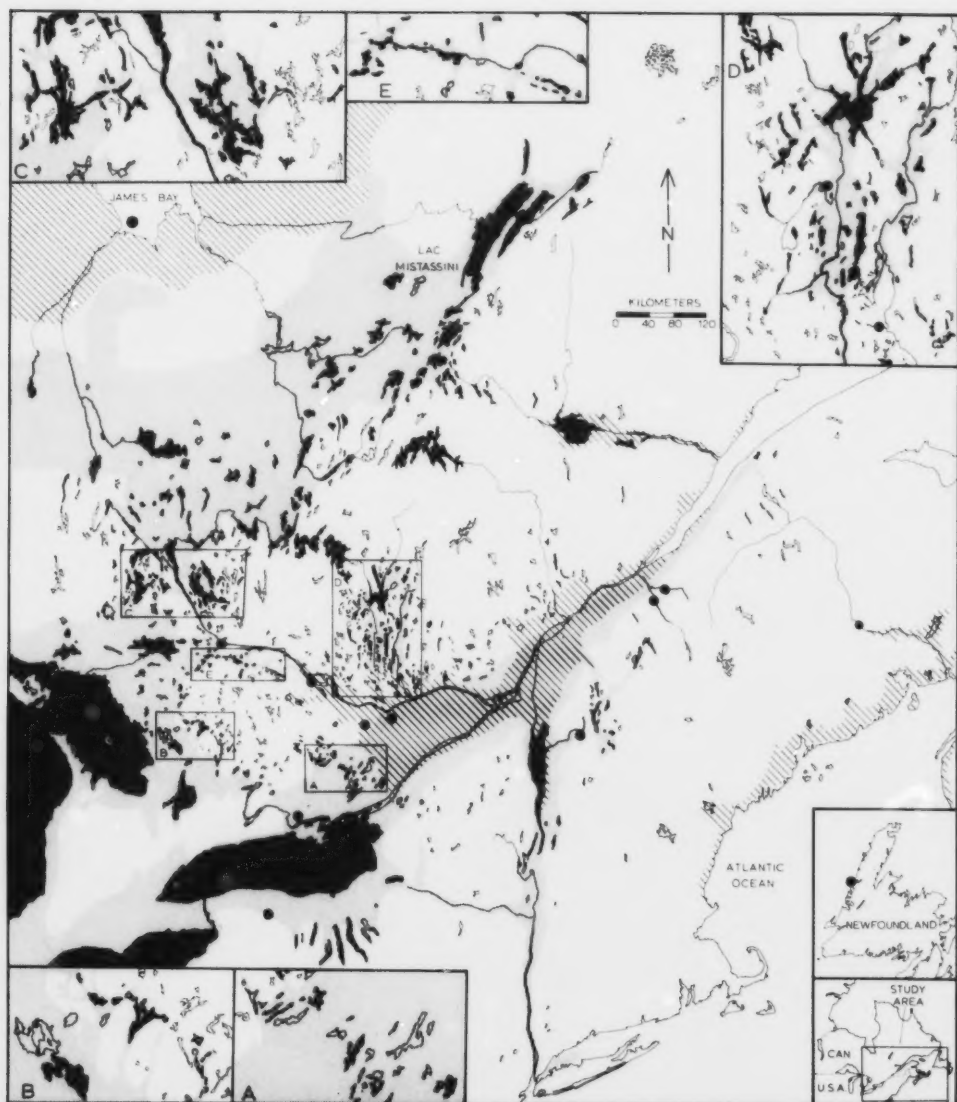
Map 4B
Distribution of *Limnocalanus macrurus* in eastern North America. Dots indicate species present; circles indicate species absent.



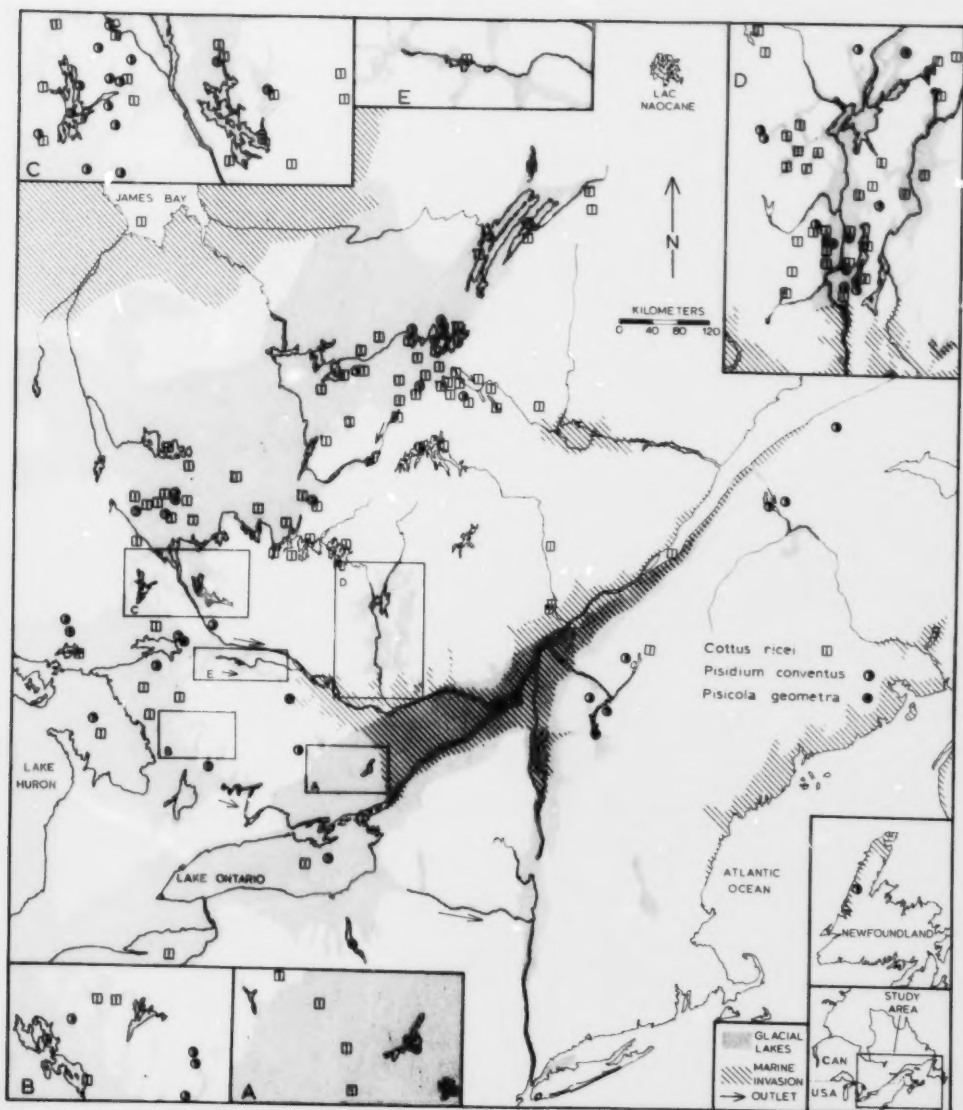
Map 5A
 Distribution of *Pontoporeia* "affinis" and *Gammaracanthus loricatus* in eastern North America. Dots indicate species present; circles indicate species absent.



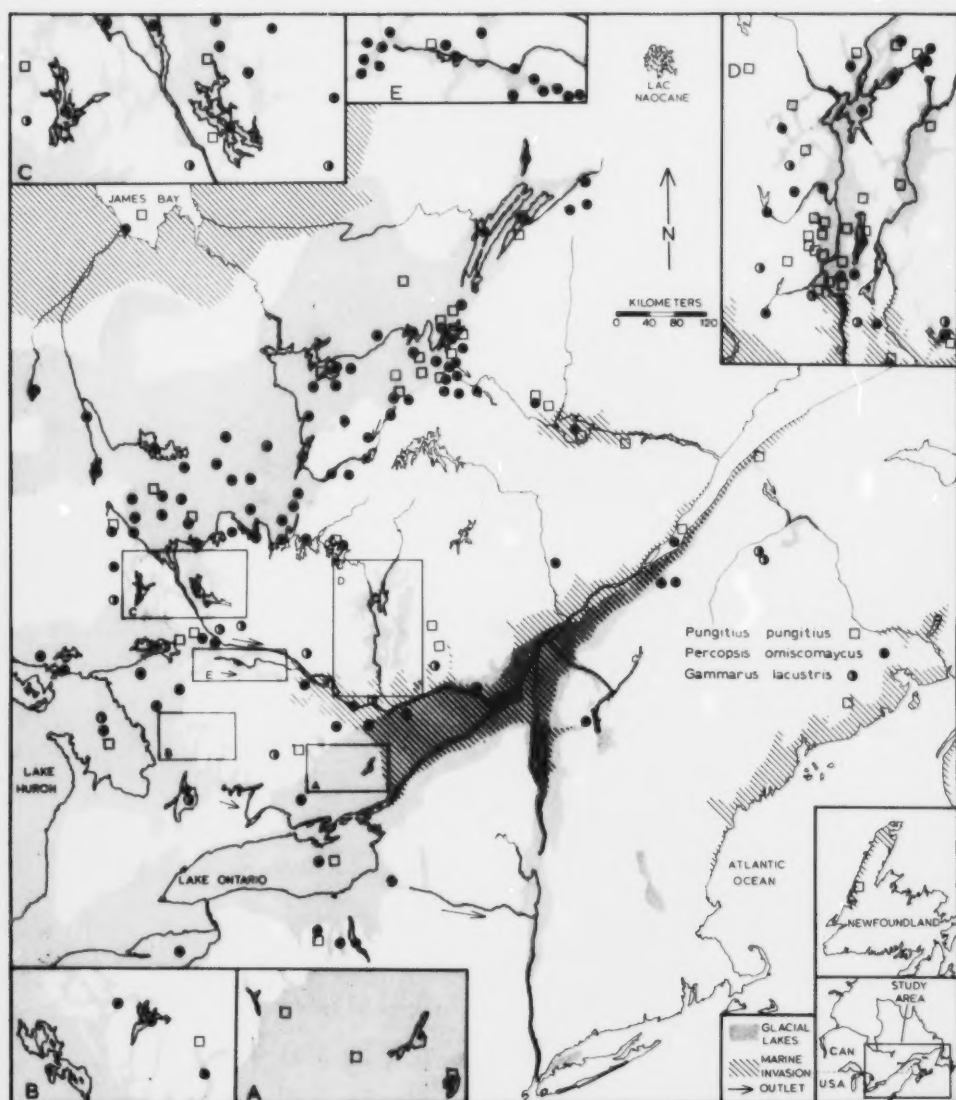
Map 5B
 Distribution of *Senecella calanoides* and *Myoxocephalus quadricornis* in eastern North America. Dots indicate species present; circles indicate species absent.



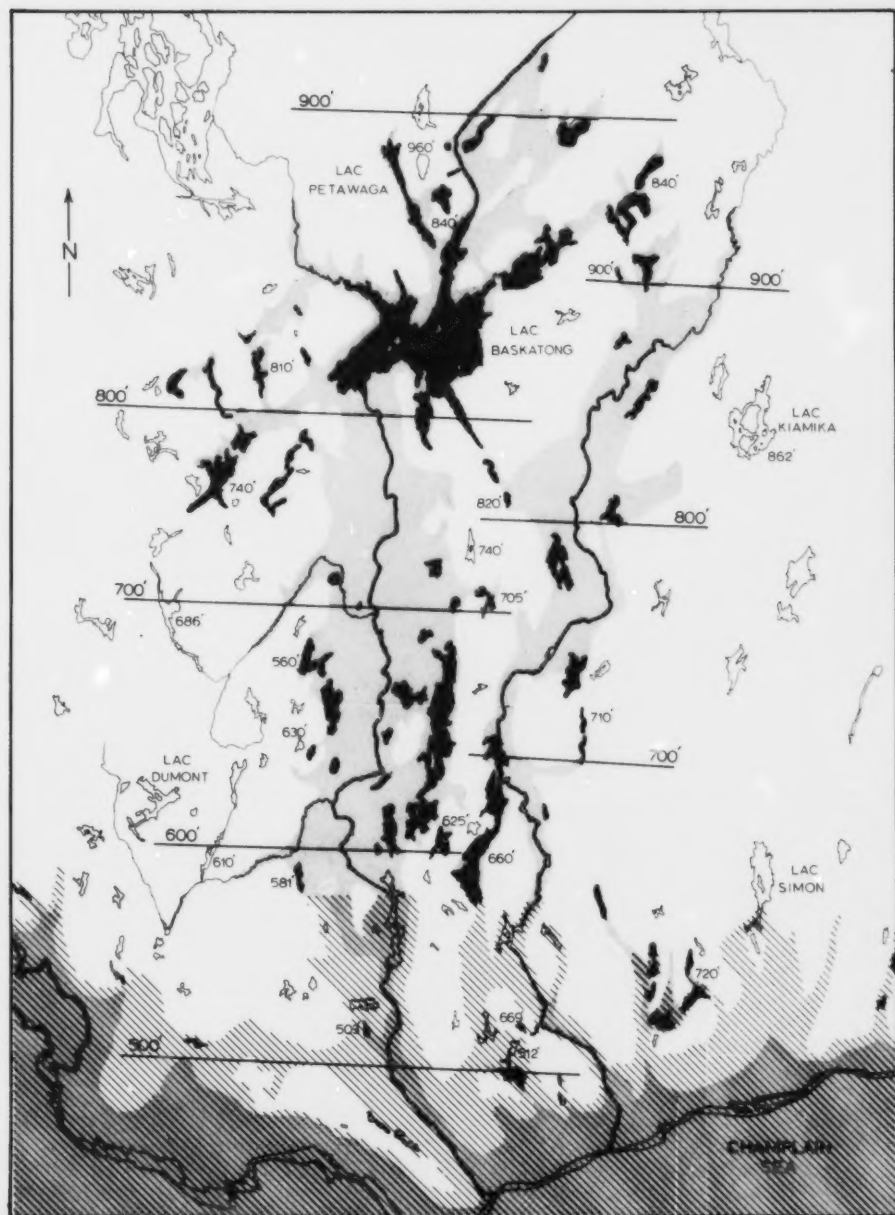
Map 6
Localities (solid lakes and dots) in which one or more of the four fish species are known. Open lakes are those sampled in which none of the fishes were found. Note moderate dispersal of fishes beyond glacial-lake boundaries.



Map 7A
Distribution of fishes and invertebrates whose dispersal was influenced, but not wholly controlled, by glacial lakes

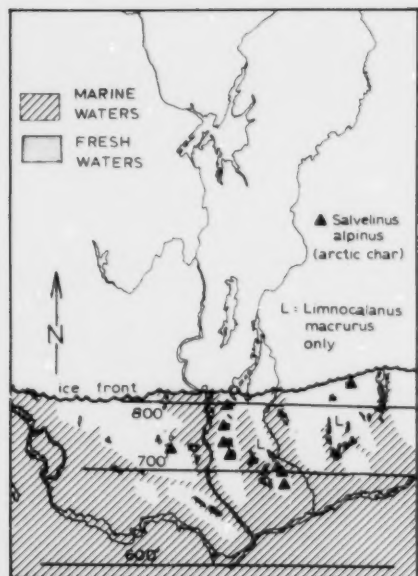


Map 7B
Distribution of fishes and invertebrates whose dispersal was influenced, but not wholly controlled, by glacial lakes

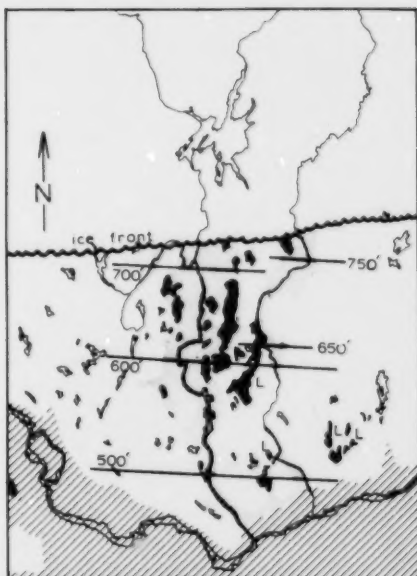


Map 8
Distribution of the community in the Gatineau (left basin) and Lièvre (right basin) valleys. Solid lakes contain one or more of the species; open lakes are ecologically acceptable lakes than contain none. Lines are isobases of maximum occurrence of the

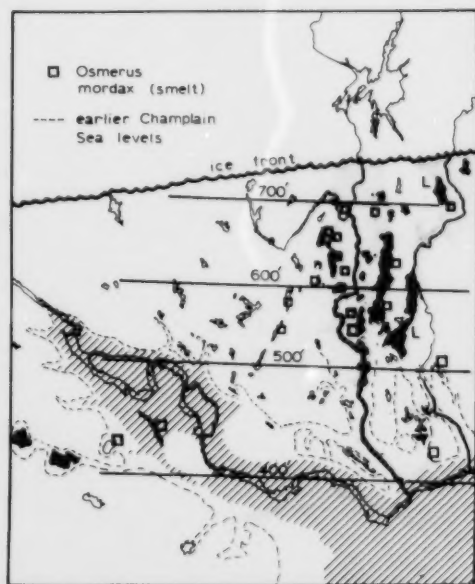
community. Numbers are elevations of lakes and isobases, in feet. Probable maximum extents of the Champlain Sea is indicated by cross-hatching, and of glacial Lake Gatineau by shading.



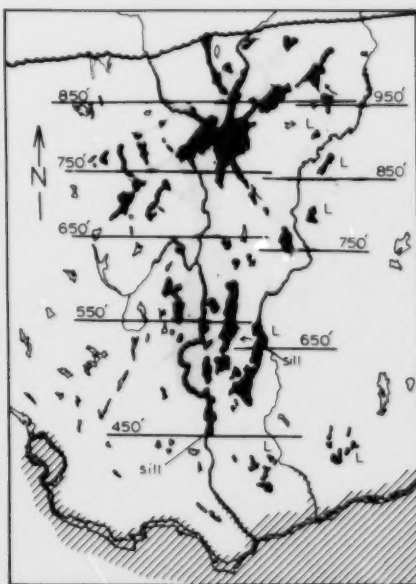
11,800 B.P. INITIAL CHAMPLAIN SEA



11,200 B.P. LAKE GATINEAU BEGINS



11,000 B.P. SMELT INVASION

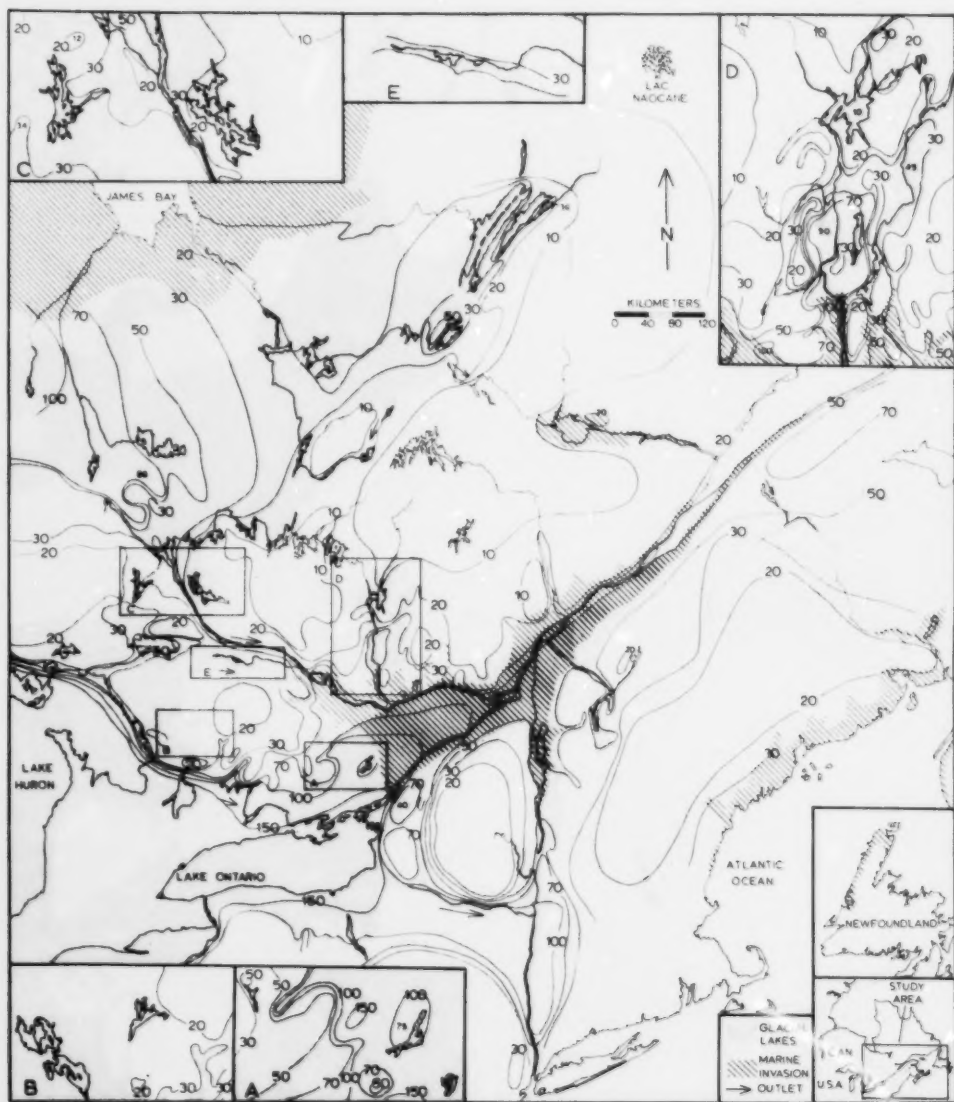


10,500 B.P. SEPARATION from MARINE

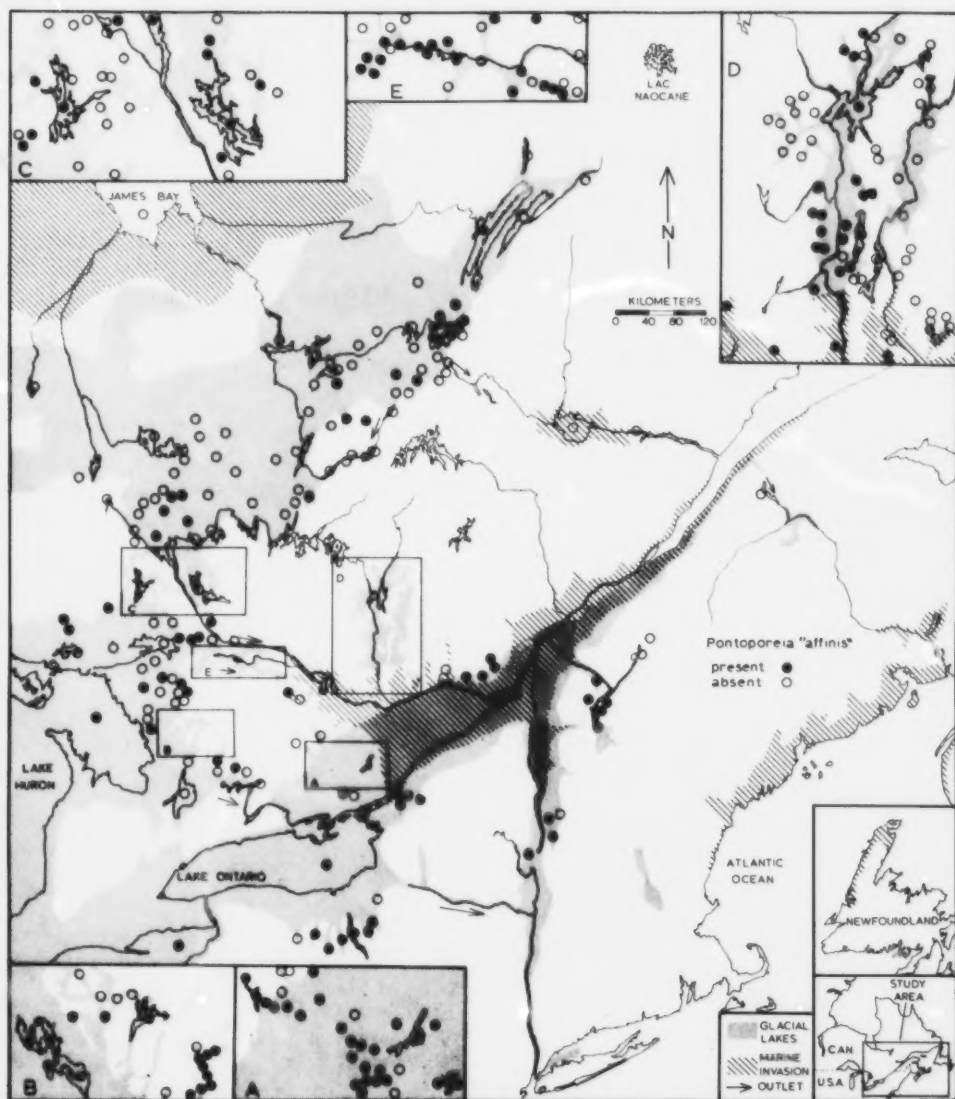
Map 9

Hypothetical stages of the Champlain Sea and glacial Lake Gatineau based on the known shoreline-level times at Kingsmere, and the distance-isobase relationship from Figure 3B. Solid lakes contain the

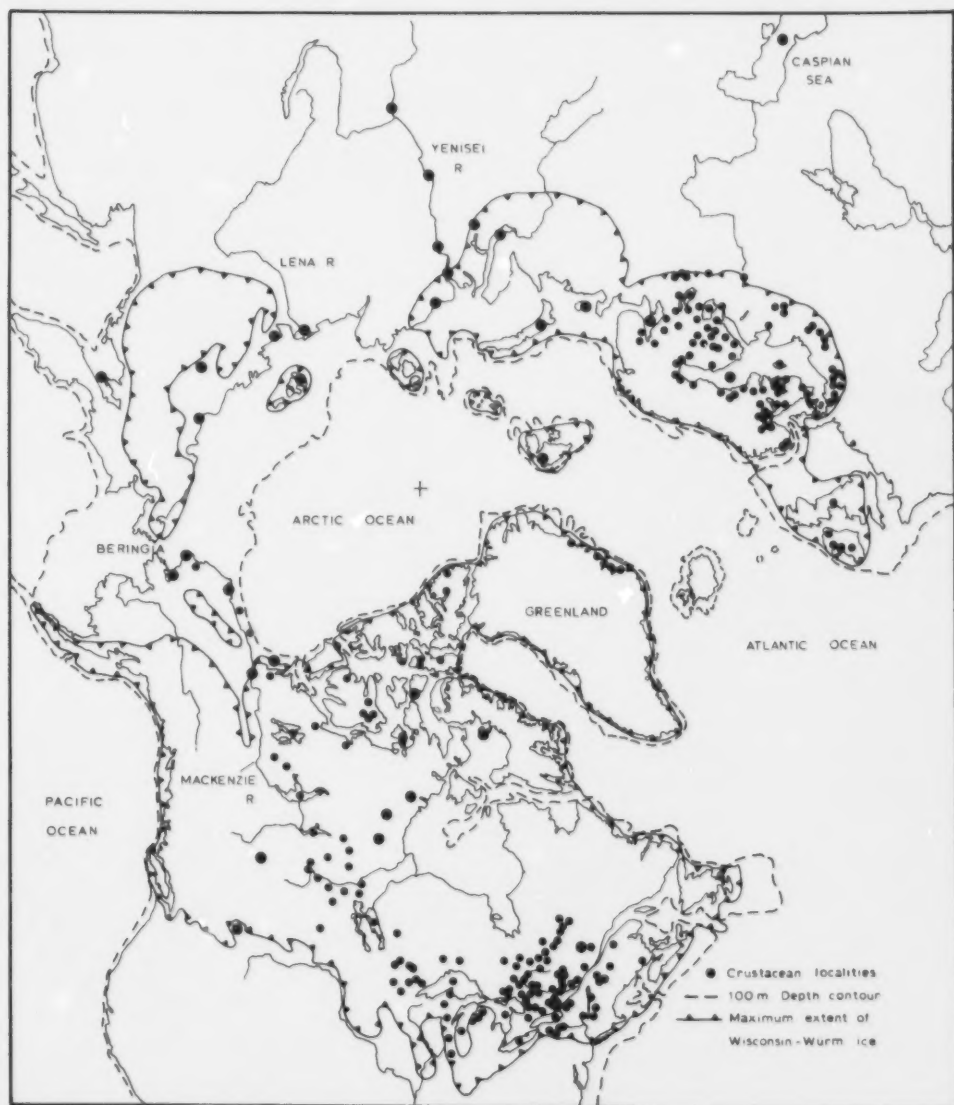
community. ⊗ indicates geographical location of Kingsmere, Quebec. Distribution of smelt is after Delisle and Veilleux (1969), distribution of arctic char after Legendre (1953).



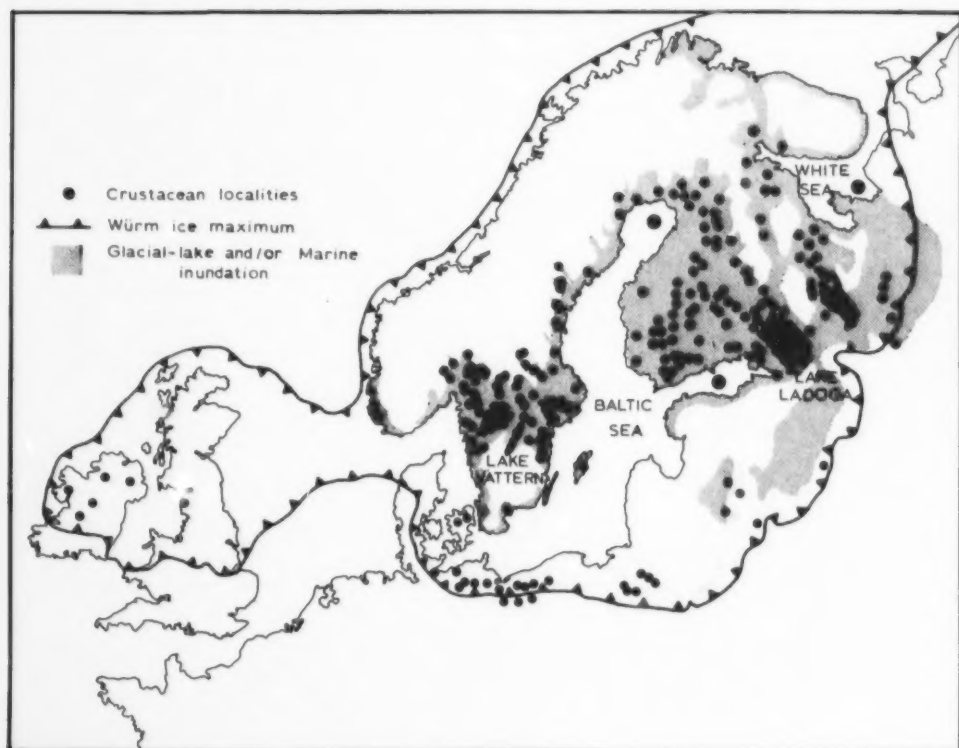
Map 10A
Distribution of total hardness in lakes of eastern
North America



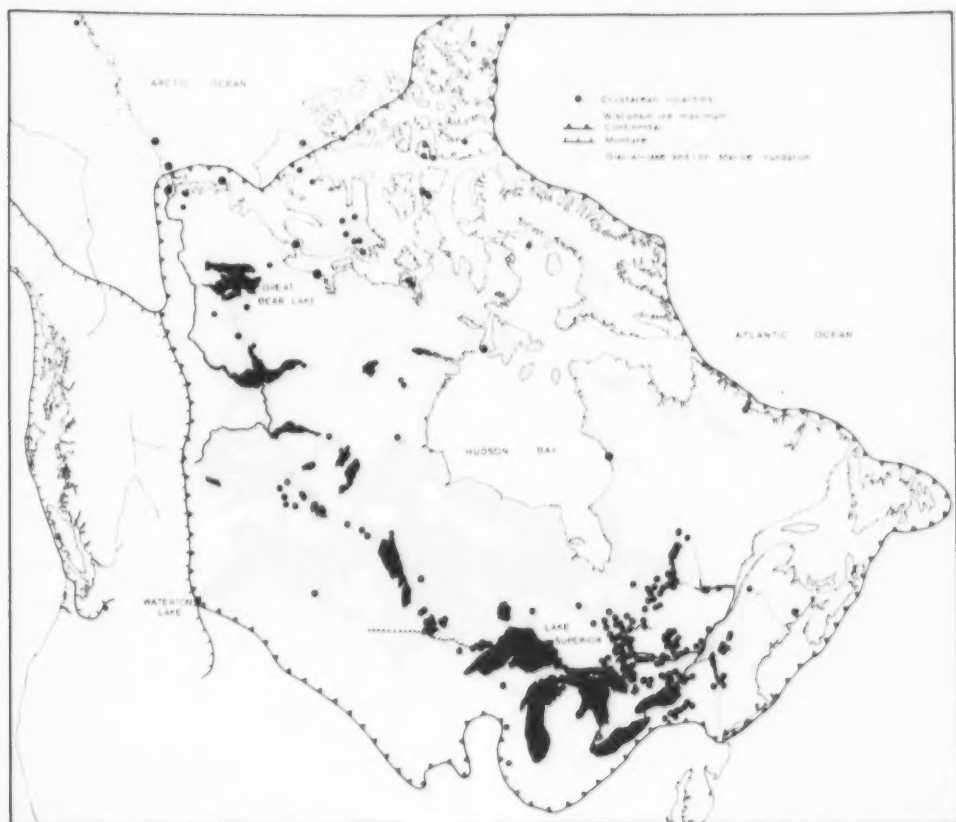
Map 10B
Distribution of *Pontoporeia "affinis"* in eastern North America in relation to total hardness of the lakes. Distribution of total hardness (TH) isobars are in ppm. Note that the frequency of occurrence increases sharply in regions with lakes of high TH.



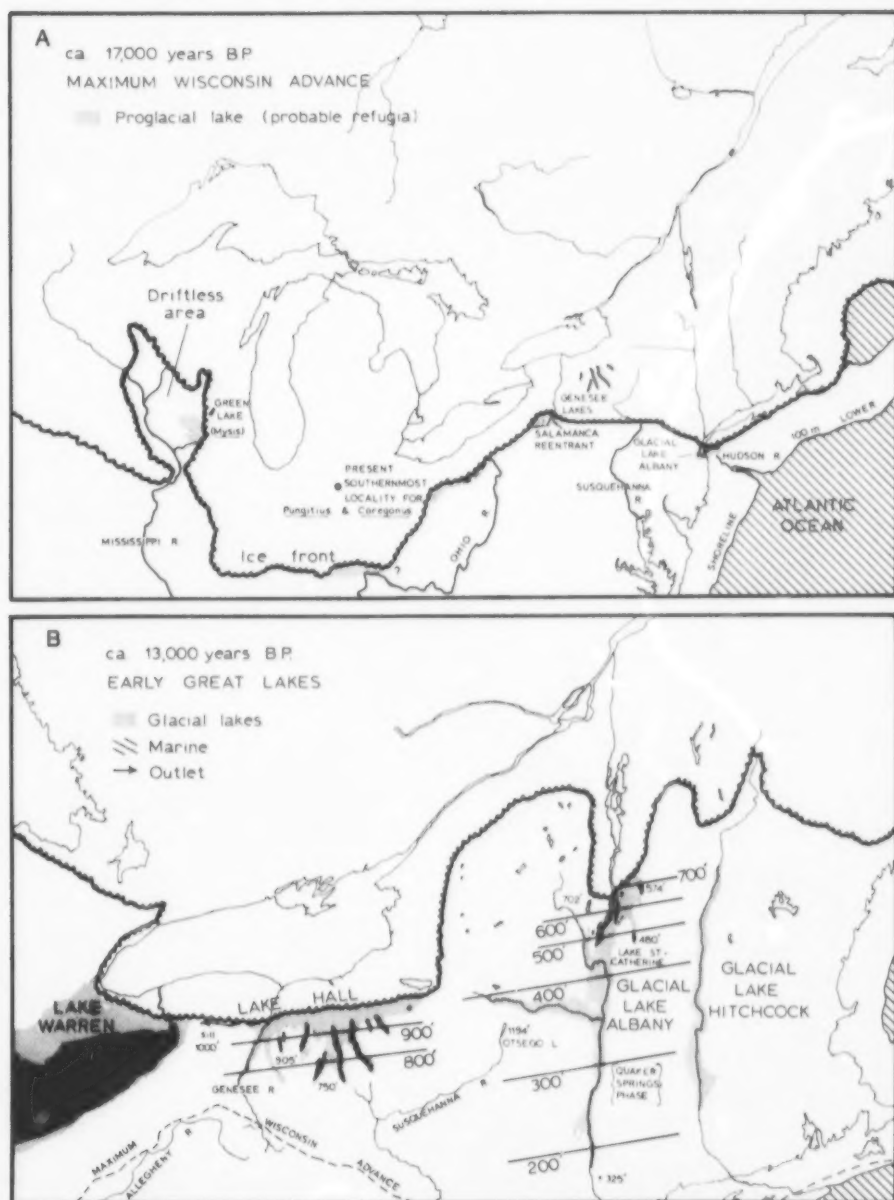
Map 11
 Distribution of the "glaciomarine relicts" in relation to the maximum extent of Wisconsin-Würm glacial ice and the probable minimum size of the Arctic Ocean during maximum glaciation. Maximum influx of fresh water would have been in the western portion of the ocean. Distribution of the crustaceans is after Segerstråle 1957; Ricker 1959; Zenkevitch 1963; Johnson 1964; Holmquist 1966; and Dadswell, this study.



Map 12
Distribution of the "glaciomarine relicts" in relation to the maximum extents of Würm glaciation and the glacial-lake-marine inundations formed during the retreat of the last ice sheet in Europe (after Segerstråle 1957, 1962; Holmquist 1966)

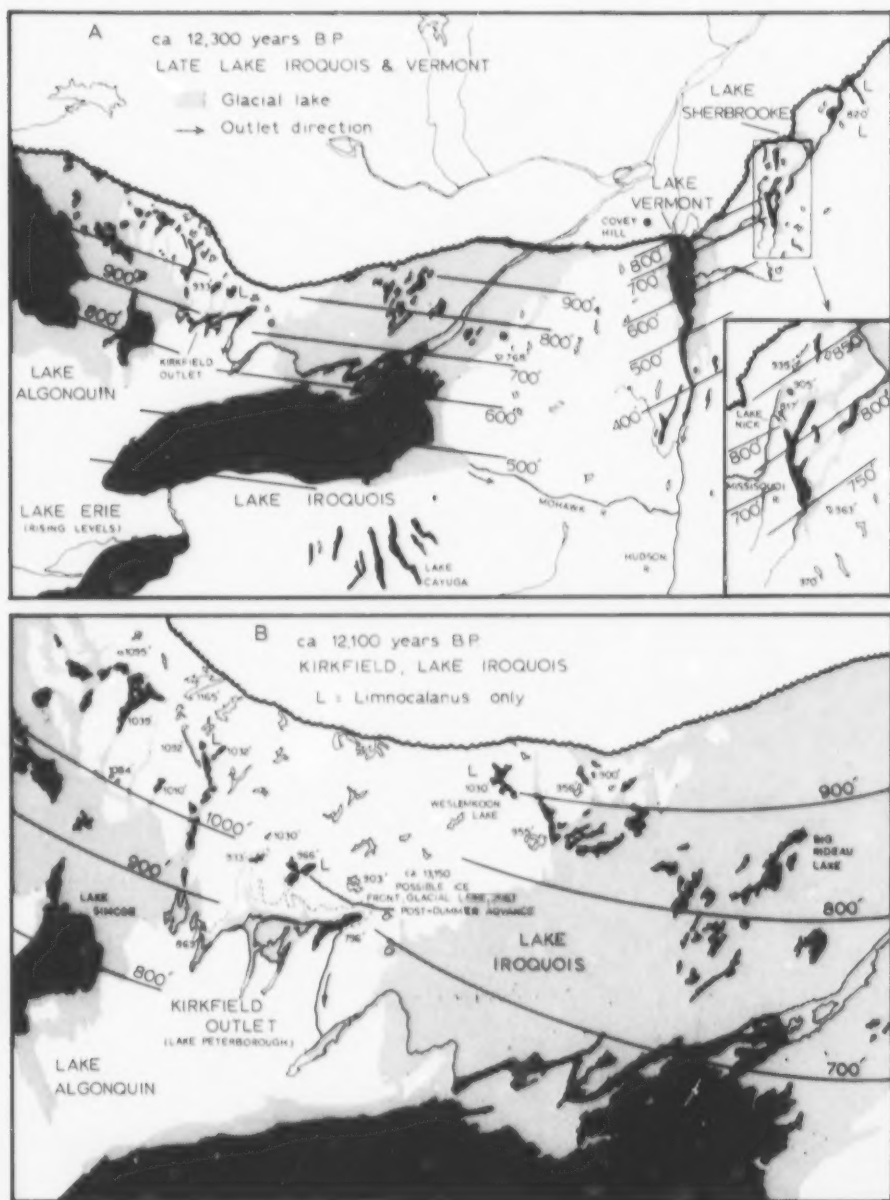


Map 13
 Distribution of the "glaciomarine relicts" in North America in relation to the maximum extents of Wisconsin glaciation and the glacial-lake-marine inundations formed during the retreat of the last ice sheet (after Prest, Grant, and Rampton 1968; Ricker 1959; Reed 1963; Johnson 1964; Holmquist 1966; and Dadswell, this study)



Map 14
A. Maximum extent of the last ice advance in eastern North America (after Prest 1970), and possible refugia for the community (after Fairchild 1932; Goldthwait et al. 1965; Frye, Willman, and Black 1965).

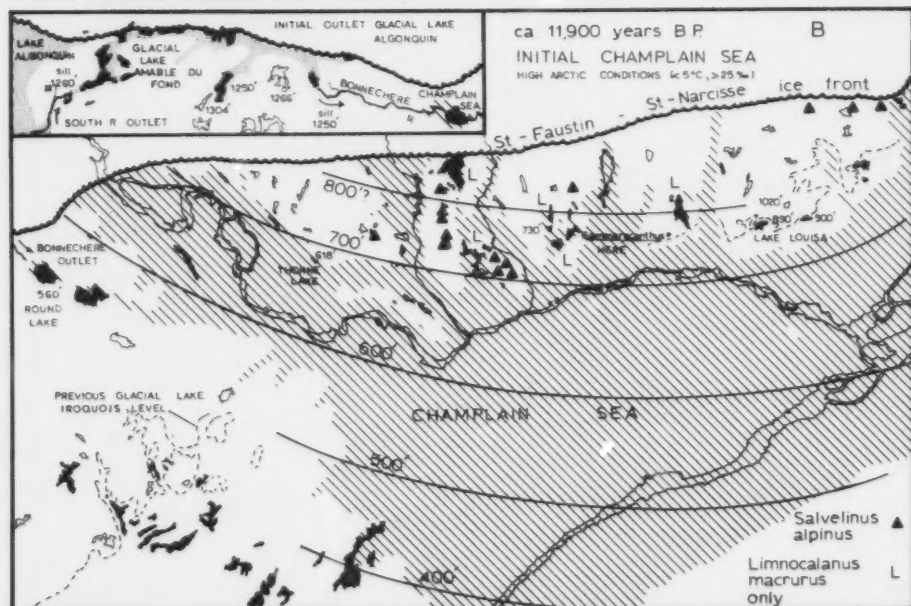
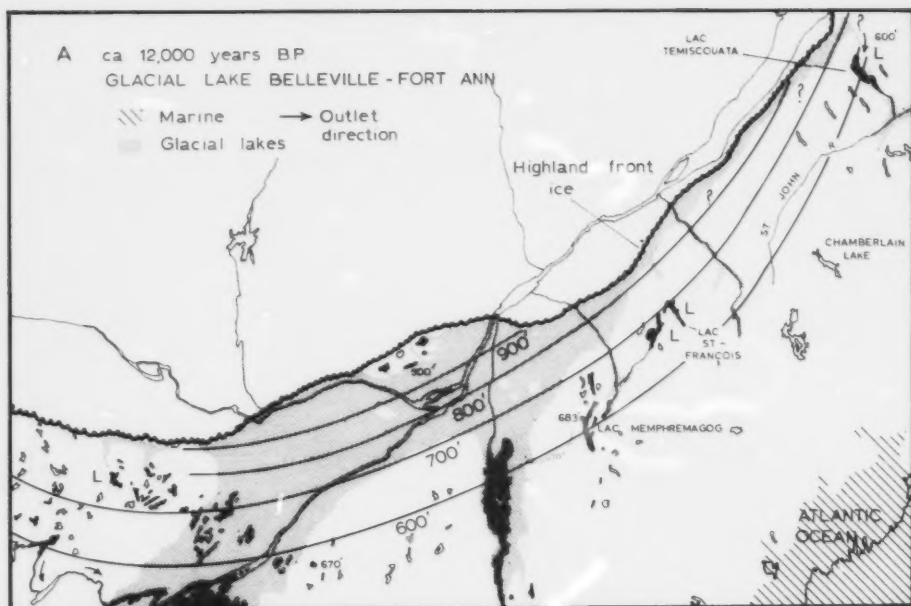
B. Early glacial lakes in eastern North America, with isobases of rebound to present elevation (after Fairchild 1932; Goldthwait et al. 1965; Connally 1972). Elevations of present-day lakes and the isobases are in feet. Solid lakes contain the community.



Map 15

A. Late glacial Lakes Iroquois and Vermont (after Prest 1970; Chapman 1937) and glacial Lake Memphremagog; inset: Sherbrooke phase (after McDonald 1968).

B. Hypothetical late phase of glacial Lake Iroquois and glacial Lake Algonquin (after Martin and Chapman 1965). Dotted line represents previous high level of Lake Peterborough. Isobases and present-day lake elevations are in feet. Solid lakes contain the community.

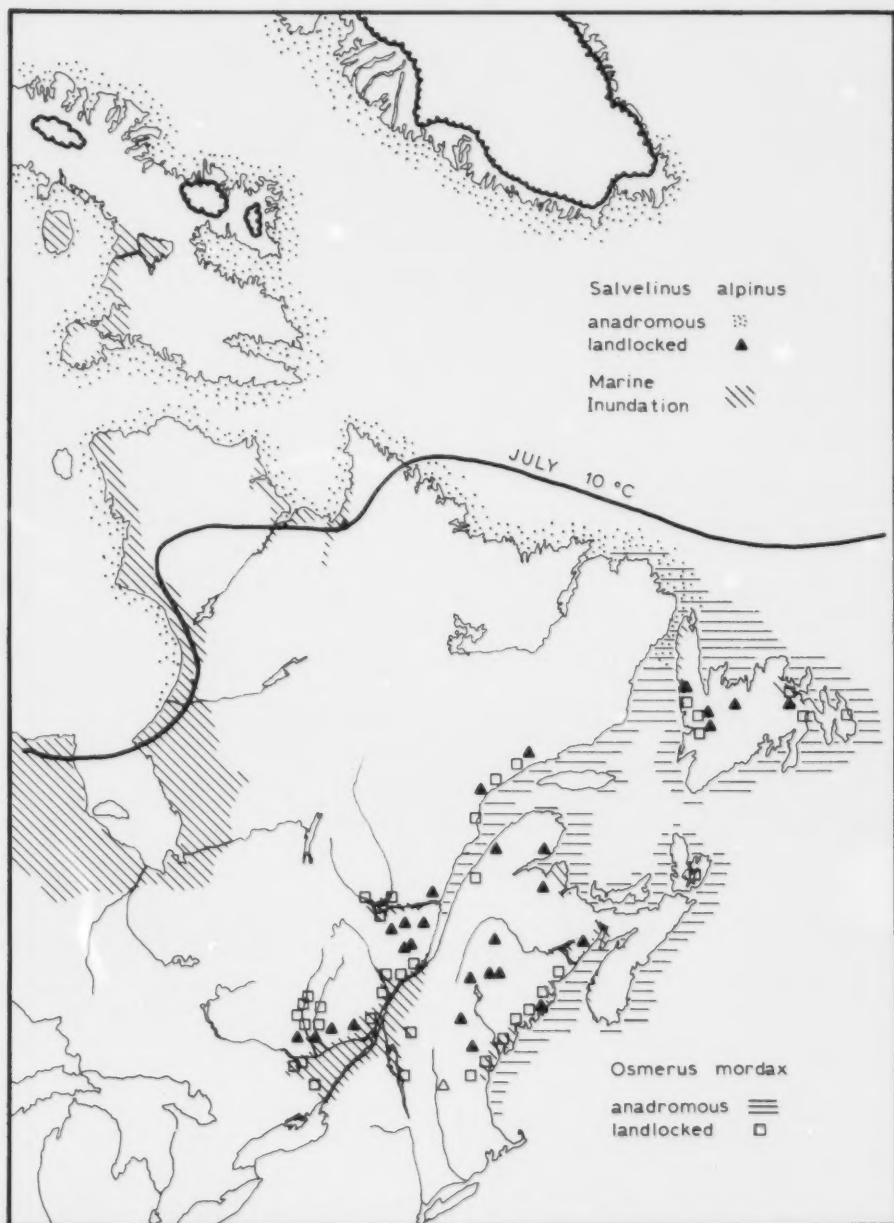


Map 16

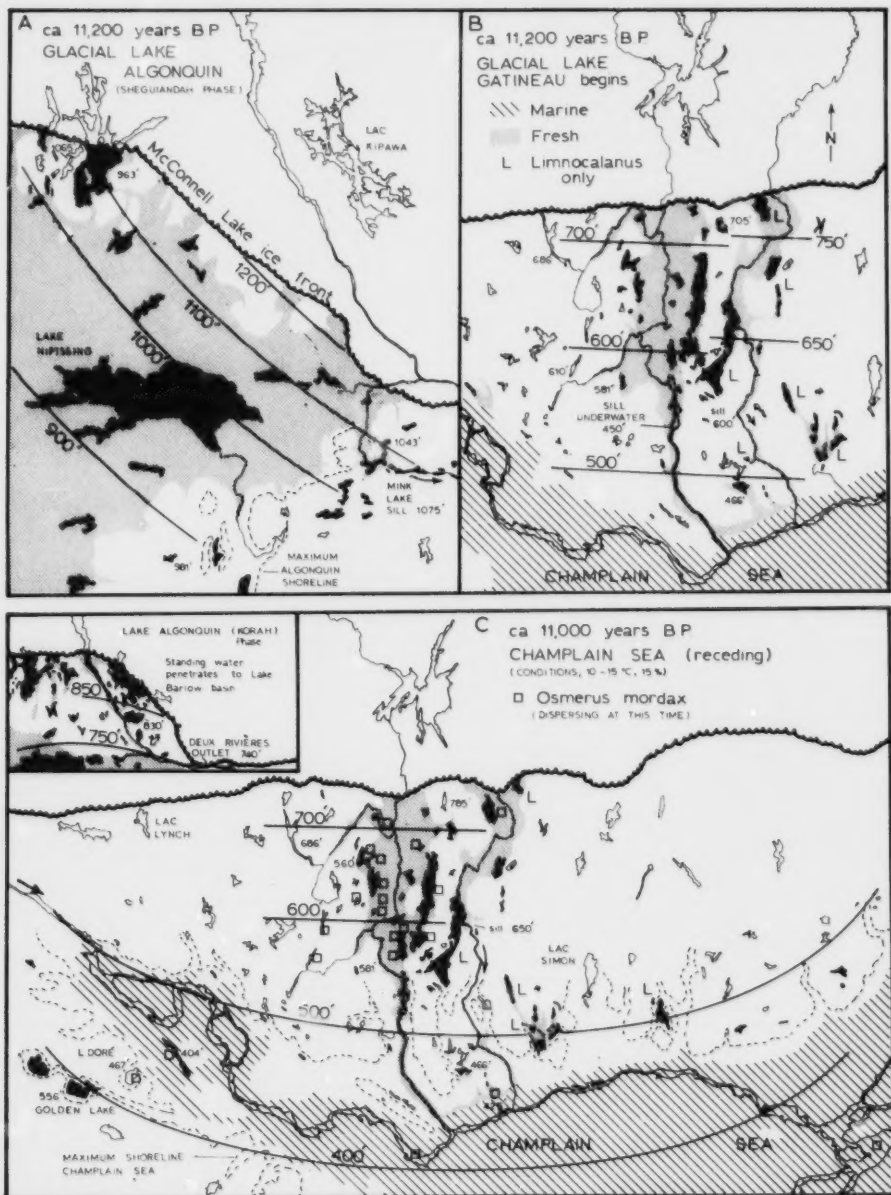
A. Glacial Lake Belleville-Fort Ann just prior to the collapse of the ice dam near Quebec City and the invasion of the sea.

B. Early Champlain Sea in the Ottawa-St. Lawrence valleys; inset: South River Outlet of glacial Lake Algonquin.

Isobases and present-day lake elevations are in feet. Solid lakes contain the community.



Map 17
Present distribution of anadromous and landlocked
populations of arctic char (*Salvelinus alpinus*) and
smelt (*Osmerus mordax*) in eastern North America
(after Legendre 1953; Leim and Scott 1966)

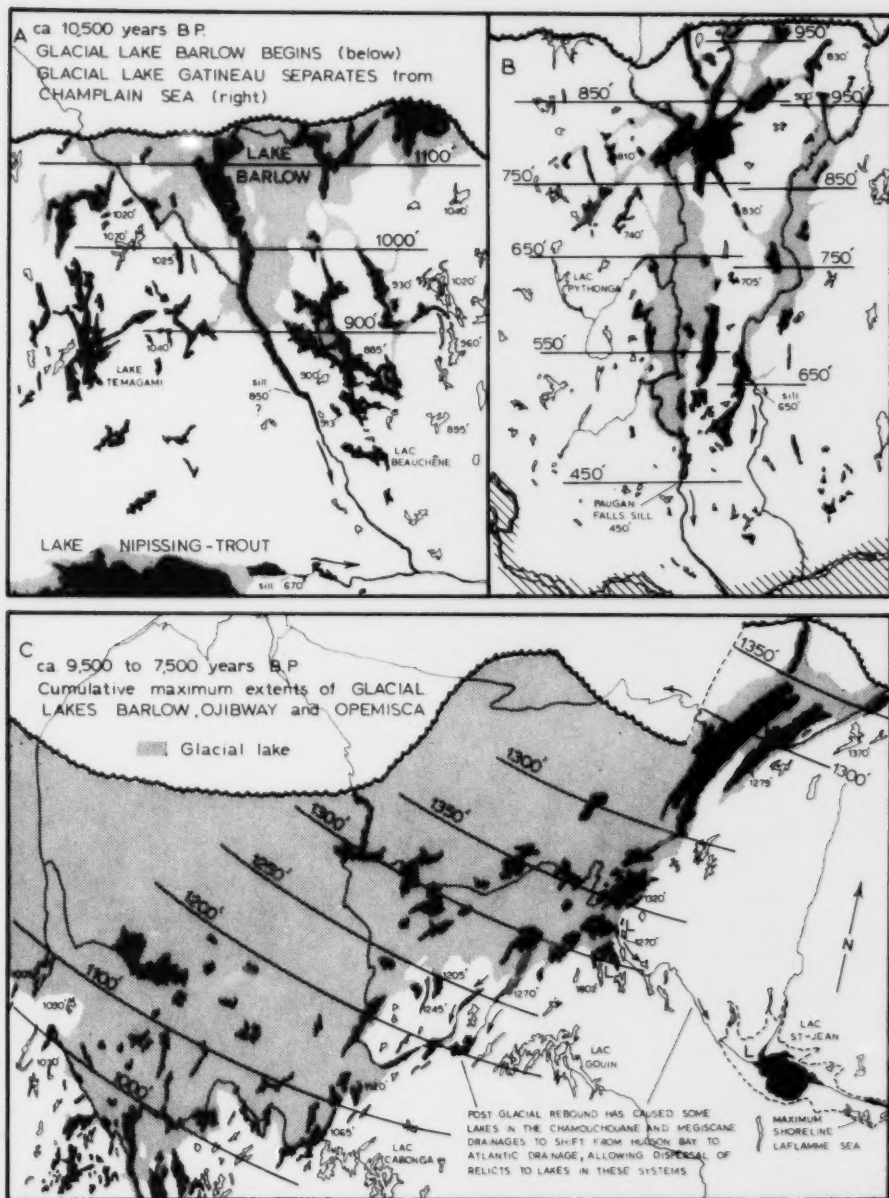


Map 18

A. Events in the Temagami-North Bay area, late glacial Lake Algonquin (after Boissoneau 1968; Harrison 1971).

B. Early phase of glacial Lake Gatineau.

C. Champlain Sea at the time of smelt invasion (smelt distribution after Dymond 1939; Delisle and Veilleux 1969); *inset*: last phase of glacial Lake Algonquin and penetration of the community into the Barlow basin (after Harrison 1971).



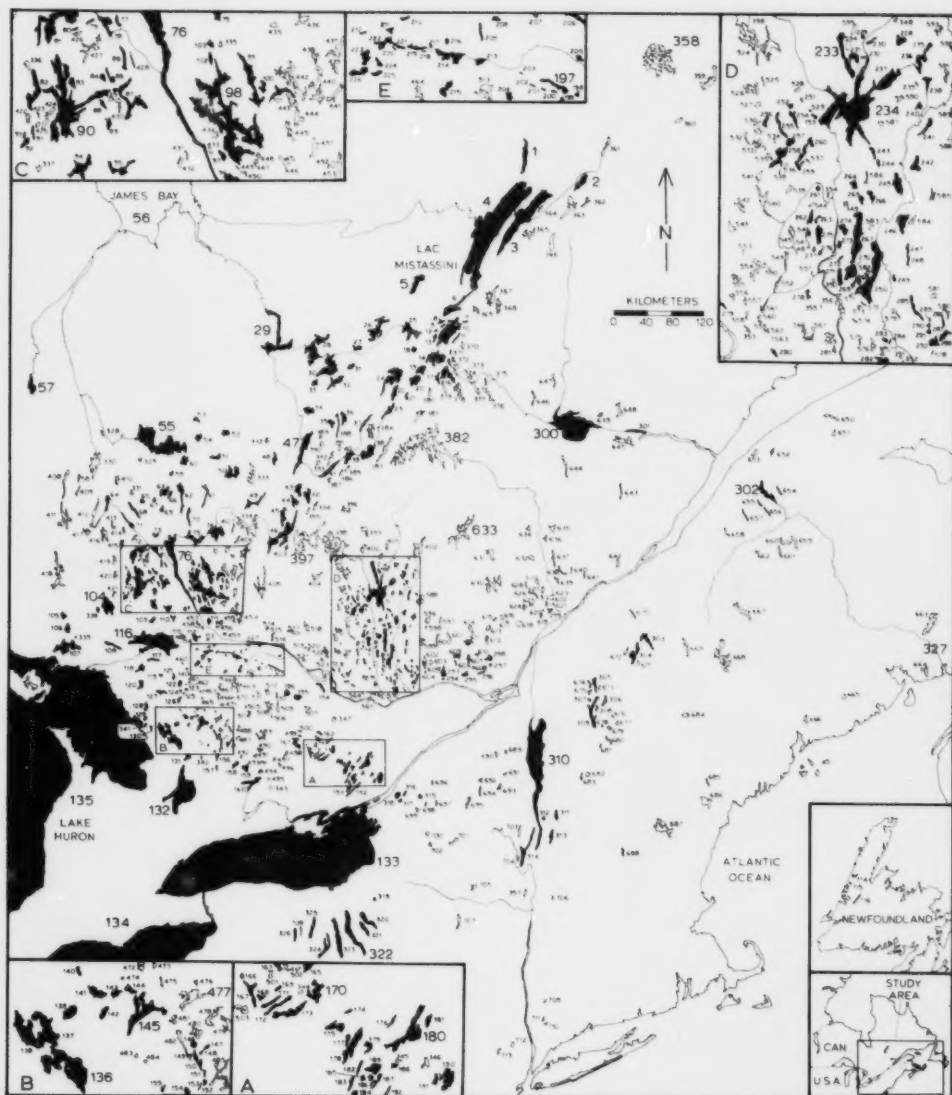
Map 19

A. Hypothetical isobase situation in the early stage of glacial Lake Barlow. Ice front is after Prest (1969).

B. Hypothetical isobases of glacial Lake Gatineau after separation from the Champlain Sea.

C. Maximum extent of glacial lakes in the Abitibi region.

Isobases are based on the northward rise of the community localities. Correlation is close to the glacial-lake outline of Prest, Grant, and Rampton (1968).



Map 20

Base map of study area, illustrating lakes considered in the study. Numbers refer to localities in Appendix 1. Solid lakes and numbers are localities in which one or more species of the deepwater community are known. Open lakes are ecologically suitable lakes lacking the deepwater community. Latitudes and longitudes of study area: 53°N–41°N and 82°W–57°W.